

Ictiofauna estuarina do nordeste do Brasil: variabilidade espacial e indicadores funcionais

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(PPG-RPAq)**

Ictiofauna estuarina do nordeste do Brasil: variabilidade espacial e indicadores funcionais

Rayssa Rayanna Salvador de Siqueira Lima

Dissertação apresentada ao Programa de Pós-Graduação em Recursos Pesqueiros e Aquicultura da Universidade Federal Rural de Pernambuco, como exigência para obtenção do título de Mestre.

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Dedicatória

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“Every day is a new day. I’m thankful for every breath I take.. So, I learn from my mistakes. It’s beyond my control sometimes.. It’s best to let go, whatever happens in this lifetime.. (P.O.D.)”

Resumo

As zonas costeiras são áreas que possuem importantes funções ecológicas e servem de ligação entre os ecossistemas marinhos e terrestres. Esta peculiaridade se deve a inúmeras características, tais como variabilidade da salinidade, suporte à reprodução das espécies, e diversidade de fontes para alimentação e elevada disponibilidade de nutrientes, tornando-as ambientes complexos e de extrema importância para o ciclo de vida de espécies marinhas. Dentre os ecossistemas costeiros tropicais, os estuários e seus manguezais são áreas dinâmicas que desempenham distintos papéis, seja para pesca, turismo, local de proteção contra enchentes e tempestades, além de servir como fonte de renda e de alimento para diversas famílias de pescadores. A diversidade das espécies pode variar em uma escala local ou entre regiões, em uma escala geográfica. Aspectos físicos e biológicos podem influenciar nesses padrões de diversidade de espécies assim como a interação entre os seus componentes, causando diferenças dentro e entre as comunidades. Várias ferramentas podem ser utilizadas para se compreender os padrões de distribuição das espécies e seu funcionamento. Apesar da existência de estudos voltados sobre a biodiversidade da ictiofauna estuarina do Brasil, estes são geralmente pontuais e em reduzido número na região nordeste, não refletindo a quantidade de ambientes estuarinos ao longo do litoral da região, indicando uma deficiência no conhecimento da composição ictiofaunística dos mesmos. Este trabalho tem o objetivo geral de compreender os padrões geográficos da ictiofauna estuarina do Nordeste do Brasil, através do conhecimento dos aspectos estruturais das assembleias e índices funcionais. O primeiro capítulo fornece uma síntese das informações sobre a ictiofauna estuarina de seis dos mais importantes estuários de Pernambuco baseado em coletas entre os anos de 2000 e 2018 correlacionando com dados abióticos, totalizando 198 espécies de peixes. Dentre estas espécies, 80% foram caracterizados como marinhas migrantes e visitantes e 70% como zoobentívoros e piscívoros. O segundo capítulo fornece uma síntese das informações disponíveis sobre a ictiofauna estuarina do Nordeste do Brasil, identificando e caracterizando padrões geográficos, também analisando as similaridades taxonômica entre os 45 estuários estudados, totalizando 339 espécies de peixes. Espécies marinhas migrantes e visitantes foram 77% do número total de espécies e 72% foram consideradas zoobentívoras e piscívoras. Em ambos os artigos, cada espécie foi designada dentro das categorias da lista de espécies ameaçadas de extinção a nível regional (ICMbio) e/ou internacional (IUCN).

Palavras-Chave:

Estuários, lista vermelha, guildas, diversidade

Abstract

Coastal zones are areas that have important ecological functions and serve as a link between marine and terrestrial ecosystems. This peculiarity is due to numerous characteristics, such as salinity variability, support for the reproduction of species, different sources of food, and high availability of nutrients, making them complex environments and of extreme importance for the life cycle of marine species. Among the tropical coastal ecosystems, estuaries and their mangroves are dynamic areas that play important roles, whether on the socioeconomic, as a source of income and food for the communities that live nearby, or on the environmental aspect (e.g. nursing areas, protection against floods and storms). The diversity of species can vary on a local scale or between regions in a geographical scale. Physical and biological aspects can influence these patterns as well as the interaction among its components, causing differences within and among communities. Several tools can be used to understand the patterns of species distribution and function. Despite the existence of studies on the biodiversity of estuarine fish fauna in Brazil, these are generally in a low number and isolated in the northeastern region, not reflecting the number of estuarine environments along the coast of the region. It indicates a deficiency in the knowledge of their ichthyofaunal composition. This study has the general objective of understanding the geographic patterns of the estuarine ichthyofauna of Northeastern Brazil, through the knowledge of the structural aspects of the assemblages and functional indices. The first chapter provides a synthesis of information on the estuarine ichthyofauna of six of the most important estuaries of Pernambuco based on samples between the years 2000 and 2018 totaling 198 fish species and correlating with abiotic data to better understand their geographical patterns. Among these species, 80% were designated as marine migrants and stragglers and 70% as zoobenthivores and piscivores. The second chapter provides a synthesis of the available information on the estuarine ichthyofauna of Northeast Brazil, identifying and characterizing geographical patterns, also analyzing taxonomic similarities among the 45 studied estuaries totaling 339 fish species. Marine migrants and stragglers species were 77% of the total number of species and 72% were considered zoobenthivorous and piscivorous. In both articles, each species was designated a category of the regional (ICMbio) and/or international (IUCN) endangered species list.

Key words:

Estuaries, red list, guilds, diversity

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1. Introdução

As zonas costeiras são áreas que possuem importantes funções ecológicas e servem de ligação entre os ecossistemas marinhos e terrestres. Esta peculiaridade se deve a inúmeras características, tais como variabilidade da salinidade, suporte à reprodução das espécies, e diversidade de fontes para alimentação e elevada disponibilidade de nutrientes, tornando-as ambientes complexos e de extrema importância para o ciclo de vida de espécies marinhas (MMA 2002; Gerling et al. 2016). Dentre os ecossistemas costeiros tropicais, os estuários e seus manguezais são áreas dinâmicas que desempenham distintos papéis, seja para pesca, turismo, local de proteção contra enchentes e tempestades, além de servir como fonte de renda e de alimento para diversas famílias de pescadores (Oliveira 1979; Boerema & Meire 2017). Os mesmos representam um dos ecossistemas mais produtivos, mesmo quando comparados com as florestas tropicais (Longhurst et al. 1995).

A região Nordeste compreende nove estados costeiros com uma extensão de 1.558.196 km², abrangendo uma grande diversidade de características oceanográficas e geomorfológicas, bem como diferentes comunidades de peixes (Mateus et al. 2008; Costanza et al. 2014; Vasconcelos et al. 2015; Loureiro et al. 2016a).

A diversidade das espécies pode variar em uma escala local ou entre regiões. Aspectos físicos e biológicos podem influenciar nos padrões de diversidade de espécies assim como a interação entre os seus componentes, causando diferenças dentro e entre as comunidades. Várias ferramentas podem ser utilizadas para se compreender os padrões de distribuição das espécies e seu funcionamento. Diversidade taxonômica e riqueza funcional são parâmetros úteis para a avaliação da integridade e resiliência dos ecossistemas estuarinos. Tratos funcionais, expressos através de guildas e índices ecológicos, têm sido utilizados na avaliação do papel destes ecossistemas, além de avaliar os possíveis efeitos dos impactos antrópicos e das mudanças climáticas (Menge & Olson 1990; Blaber 2000; Flores-Lopes et al. 2010; Contente 2013). Guildas ecológicas são utilizadas para fornecer informações funcionais, estruturas hierárquicas e conectividade, visando simplificar o entendimento de ecossistemas complexos (Elliott et al. 2007b). Guilda trófica é definida como a forma que um grupo de espécies que, independentemente de sua afiliação taxonômica, exploram uma classe de recursos alimentares de maneira parecida (Elliott et al. 2007b). Guilda reprodutiva caracteriza-se como parte da estratégia reprodutiva, se expressando por diversas maneiras, através de diferentes táticas, como fecundação interna ou externa, diferenciação na idade de maturação, presença e/ou ausência de cuidado parental, entre outros fatores (Vazzoler 1996). Guilda ambiental caracteriza-se na forma como as espécies utilizam o local e qual o impacto que o mesmo tem sobre elas (Elliott et al. 2007b).

Apesar da existência de estudos voltados sobre a biodiversidade da ictiofauna estuarina do Brasil (Paiva et al. 2013a; Vilar & Joyeux 2018a), estes são geralmente pontuais e em reduzido número na região nordeste, (por exemplo Colares et al., 2016; Mérigot et al., 2017; Silva-Júnior et al., 2017; Ferreira et al., 2019), não refletindo a quantidade de ambientes estuarinos ao longo do litoral da região, indicando uma deficiência no conhecimento da composição ictiofaunística dos mesmos. Além da carência desse conhecimento, a maioria dos estudos realizados não contemplam uma abordagem funcional das comunidades ou parâmetros que permitam uma visão comparativa e integradas das mesmas.

De acordo com Cowman (2014) e Pinheiro et al. (2017), entender a biodiversidade e os padrões de endemismo são essenciais para o entendimento dos padrões geográficos e ecologia. Devido à importância que os sistemas estuarinos possuem na manutenção das populações ictiofaunísticas, o estudo sobre a organização funcional das comunidades são informações relevantes visando a conservação desses recursos renováveis, mitigação de impactos e aprimoramento do manejo destes locais (Karr 1991; Loebmann & Vieira 2005; Mouillot et al. 2007; Barletta et al. 2010).

Este trabalho, composto de dois artigos, tem o objetivo geral de compreender os padrões geográficos da ictiofauna estuarina do Nordeste do Brasil, através do conhecimento dos aspectos estruturais das assembleias e índices funcionais. O primeiro capítulo fornece uma síntese das informações sobre a ictiofauna estuarina de Pernambuco, Nordeste do Brasil, correlacionando com dados abióticos e caracterizando padrões geográficos. O segundo capítulo fornece uma síntese das informações disponíveis sobre a ictiofauna estuarina do Nordeste do Brasil, identificando e caracterizando padrões geográficos, também analisando as similaridades taxonômica entre os estuários.

2. Referências

- Barletta M, Jaureguizar AJ, Baigun C, et al (2010) Fish and aquatic habitat conservation in South America: A continental overview with emphasis on neotropical systems. *Journal of Fish Biology* 76:2118–2176. <https://doi.org/10.1111/j.1095-8649.2010.02684.x>
- Blaber SJM (2000) Tropical estuarine fishes. ecology, exploitation and conservation. Blackwell Science 372. [https://doi.org/10.1643/0045-8511\(2001\)001\[0880:\]2.0.CO;2](https://doi.org/10.1643/0045-8511(2001)001[0880:]2.0.CO;2)
- Boerema A, Meire P (2017) Management for estuarine ecosystem services: A review. *Ecological Engineering* 98:172–182. <https://doi.org/10.1016/j.ecoleng.2016.10.051>

- Colares MCDS, Pinheiro LDS, Menezes MOB, Morais JO de (2016) Caracterização sedimentar do Canal do estuário do Rio Coreaú, estado do Ceará, Brasil. *Arquivos de Ciências do Mar* 49:5–12. <https://doi.org/10.32360/acmar.v49i1.4728>
- Contente RF (2013) Padrões ecológicos locais e multidecadais da ictiofauna do estuário Cananéia-Iguape. 186
- Costanza R, de Groot R, Sutton P, et al (2014) Changes in the global value of ecosystem services. *Global Environmental Change*. <https://doi.org/10.1016/j.gloenvcha.2014.04.002>
- Cowman PF (2014) Historical factors that have shaped the evolution of tropical reef fishes: A review of phylogenies, biogeography, and remaining questions. *Frontiers in Genetics* 5:1–15. <https://doi.org/10.3389/fgene.2014.00394>
- Elliott M, Whitfield AK, Potter IC, et al (2007) The guild approach to categorizing estuarine fish assemblages: A global review. *Fish and Fisheries* 8:241–268. <https://doi.org/10.1111/j.1467-2679.2007.00253.x>
- Ferreira V, le Loc'h F, Ménard F, et al (2019) Composition of the fish fauna in a tropical estuary: the ecological guild approach. *Scientia Marina* 83:133. <https://doi.org/10.3989/scimar.04855.25a>
- Flores-Lopes F, Cetra M, Malabarba LR (2010) Utilização de índices ecológicos em assembleias de peixes como instrumento de avaliação da degradação ambiental em programas de monitoramento. *Biota Neotropica* 10:184–193
- Gerling C, Ranieri C, Fernandes L, et al (2016) Manual de ecossistemas marinhos e costeiros para educadores. Editora Comunicar, Santos
- Karr JR (1991) Biological Integrity: A Long-neglected aspect of water resource management. *Ecological Applications* 1:66–84. [https://doi.org/10.1016/S0960-8524\(96\)00113-7](https://doi.org/10.1016/S0960-8524(96)00113-7)
- Loebmann D, Vieira JP (2005) Distribuição espacial e abundância das assembleias peixes no Parque Nacional da Lagoa do Peixe, Rio Grande do Sul, Brasil. *Revista Brasileira de Zoologia* 22:667–675
- Longhurst A, Sathyendranath S, Platt T, Caverhill C (1995) An estimate of global primary production in the ocean from satellite radiometer data. *Journal of Plankton Research* 17:1245–1271. <https://doi.org/10.1093/plankt/17.6.1245>
- Loureiro SN, Reis-Filho JA, Giarrizzo T (2016) Evidence for habitat-driven segregation of an estuarine fish assemblage. *Journal of fish biology* 89:804–820. <https://doi.org/10.1111/jfb.13017>
- Mateus M, Mateus S, Barrett (2008) Basic concepts of estuarine ecology. In: *Perspectives on Integrated Coastal Zone Management in South America*. IST Press, Lisboa, pp 3–14
- Menge BA, Olson AM (1990) Role of scale and environmental factors in regulation of community structure. *Trends in Ecology & Evolution* 5:52–57. [https://doi.org/10.1016/0169-5347\(90\)90048-I](https://doi.org/10.1016/0169-5347(90)90048-I)
- Mérigot B, Frédoú FL, Viana AP, et al (2017) Fish assemblages in tropical estuaries of northeast Brazil: A multi-component diversity approach. *Ocean and Coastal Management* 143:175–183. <https://doi.org/10.1016/j.ocecoaman.2016.08.004>

- MMA (2002) Avaliação e identificação de áreas e ações prioritárias para conservação, utilização sustentável e repartição dos benefícios da biodiversidade nos biomas brasileiros. In: Biodiversidade brasileira. MMA/SBF, Brasília, p 404
- Mouillot D, Dumay O, Tomasini JA (2007) Limiting similarity, niche filtering and functional diversity in coastal lagoon fish communities. *Estuarine, Coastal and Shelf Science* 71:443–456. <https://doi.org/10.1016/j.ecss.2006.08.022>
- Oliveira AME de (1979) Distribuição dos peixes nos estuários do nordeste brasileiro de acordo com a salinidade da água. 88
- Paiva ACG de, Chaves P de T, Araújo ME de (2013) Distribution of estuarine fish fauna along coast of Brazil. *Tropical Oceanography Online* 41:1–36
- Pinheiro HT, Bernardi G, Simon T, et al (2017) Island biogeography of marine organisms. *Nature* 549:82–85. <https://doi.org/10.1038/nature23680>
- Silva-Júnior CAB, Mérigot B, Lucena-Frédou F, et al (2017) Functional diversity of fish in tropical estuaries: A traits-based approach of communities in Pernambuco, Brazil. *Estuarine, Coastal and Shelf Science* 198:413–420. <https://doi.org/10.1016/j.ecss.2016.08.030>
- Vasconcelos RP, Henriques S, França S, et al (2015) Global patterns and predictors of fish species richness in estuaries. *Journal of Animal Ecology* 84:1331–1341. <https://doi.org/10.1111/1365-2656.12372>
- Vazzoler AEA de M (1996) *Biologia da reprodução de peixes teleósteos: Teoria e prática*. Editora da Universidade Estadual de Maringá, Maringá
- Vilar CC, Joyeux J-C (2018) Synopsis of the knowledge on the Brazilian estuarine fishes. *PeerJPrints* 37. <https://doi.org/https://doi.org/10.7287/peerj.preprints.27085v1> |

3. Artigo científico 1

3.1. Neotropical estuarine fishes of the Northeast Brazil (Pernambuco): ecology, conservation, and updated checklist

Neotropical estuarine fishes of Northeast Brazil (Pernambuco): ecology, conservation, and updated checklist

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Abstract

Estuaries are productive environments and suppliers of many ecosystem processes, such as coastal protection and fisheries resources. However, while being severely impacted by anthropogenic

activities, information for their conservation is still fragmented and lacking, especially in tropical areas such as Brazil. Fish biodiversity and its relationship with abiotic variables in six estuaries of northeastern Brazil (Pernambuco) was analyzed. Based on 97 surveys (from 2000 to 2018), each fish species was classified according to its feeding mode, estuarine functional group, and IUCN conservation status at a regional level. A total of 197 fish species among 57 families were identified. Marine stragglers and migrants represented 80% of the species identified, while 70% of them were zoobenthivorous and piscivorous. Additionally, commercially important fishes and nine IUCN risk-categorized species were found. Together, these findings reinforce the role of estuaries in supporting coastal fish communities, including socioeconomically relevant and endangered species. Multivariate analysis evidenced taxonomic similarities in three groups: I- Sirinhaém, Santa Cruz Channel, and Rio Formoso; II- Jaguaribe; and III- Maracaípe and Suape, which were influenced by river mouth size, depth, and rainfall. Finally, a list of the main fish species was provided, highlighting the region highest biodiversity compared to estuaries at similar latitudes.

Resumo

Os estuários são ambientes produtivos e possuem diversos papéis para os ecossistemas associados. Apesar de serem severamente impactados, informações sobre conservação ainda são fragmentadas e carentes, especialmente em áreas tropicais como o Brasil. A ictiofauna estuarina de seis estuários de Pernambuco (Brasil) foi analisada e correlacionada com variáveis abióticas baseado em 97 coletas (de 2000 a 2018). As espécies foram classificadas de acordo com seu modo de alimentação, grupo funcional e status de conservação a nível regional (IUCN). Ao todo, 197 espécies de peixes entre 57 famílias foram identificadas. Marinhos visitantes e migrantes representaram 80% das espécies identificadas, enquanto zoobentívoros e piscívoros representaram 70%. Além disso, foram encontrados peixes de importância comercial e nove espécies classificadas pela IUCN como de risco. Juntas, estas descobertas reforçam o papel dos estuários no apoio às assembleias de peixes costeiros, incluindo espécies socioeconomicamente relevantes e ameaçadas de extinção. Análises multivariadas evidenciaram similaridades taxonômicas em três grupos: Sirinhaém, Canal Santa Cruz e Rio Formoso; Jaguaribe; e Maracaípe e Suape, que foram influenciados pelo tamanho da foz, profundidade e pluviosidade. Finalmente, foi fornecida uma lista das principais espécies de peixes, destacando a região estudada como a maior biodiversidade quando comparados com estuários em latitudes semelhantes.

Keywords, Palavras-chave

Keywords: Biodiversity, Coastal habitats, Fish assemblage, Guilds, IUCN

Palavras-chave: Assembleia de peixes, Biodiversidade, Guildas, Habitats costeiros, IUCN

Running Head

Estuarine fish species of Pernambuco (NE, Brazil)

Introduction

Estuaries are productive environments and suppliers of many ecosystem services, such as raw materials, coastal protection, nutrients recycling, fisheries resources, carbon sequestration, and tourism (Costanza et al. 2014; Boerema, Meire, 2017). These ecosystems are sources of income and food, being directly responsible for maintaining the daily lives of traditional populations worldwide (Barletta, Costa, 2009). They are also one of the most biologically valuable areas in the world (Costanza et al. 2014), providing essential habitats for the growth, feeding, and breeding ground of many fish species at different life stages (Elliott et al. 2007; Potter et al. 2015).

Estuarine fishes have an essential role in conducting energy throughout the trophic levels or exchanging energy with neighboring environments (Blaber, 2000; Potter et al. 2011). Different tools can be used to understand species distribution patterns and their functioning. Functional diversity and taxonomic richness are useful parameters for assessing the functionality, integrity, and resilience of estuarine ecosystems, which can also be expressed through ecological guilds, useful for the evaluation of the role of this ecosystem (Menge, Olson, 1990; Blaber, 2000; Elliott et al. 2007; Flores-Lopes et al. 2010; Contente, 2013).

The Brazilian coastline has over one hundred estuaries, with differential freshwater inputs, tidal regimes, and geomorphological characteristics (Dominguez, 2009; Bernadino et al. 2016). In Pernambuco state, northeast of Brazil, estuaries are highly threatened by deforestation, erosion, pollutants from industrial and agricultural discharge, sewage, and shrimp's farming. In addition, the lack of proper data hampers the assessment of the real impact of these activities and the elaboration of conservation policies (CPRH, 2001; Souza et al. 2007; Pelage et al. 2019).

There has been considerable progress in recent years regarding the knowledge on estuarine fishes of Pernambuco, with an increase in the number of publications on biodiversity (Ferreira et al. 2019), trophic ecology (Lira et al. 2018), functional diversity (Mérigot et al. 2017), contamination (Silva et al. 2018), population dynamics (Aschenbrenner, Ferreira, 2015), environmental connectivity (Falcão, 2012; Santana et al. 2018) and use of living resources (Barletta, Costa, 2009), even if it is still incipient compared to other regions of Brazil and the world (Whitfield 1994, 1999; Gillanders et al. 2011; James, Harrison, 2016; Nakayama et al. 2020). Moreover, for a long time,

most of the information has been scattered and limited to single estuaries, and/or restricted to the “grey literature” (*e.g.*, thesis, dissertations, monographs, abstracts, and technical reports) (Paiva, Araújo, 2010), with no integrated analysis or considering the abiotic estuarine characteristics. Physical and biological aspects can influence the patterns of species diversity and the interaction between its components (Fernandes, 2003).

The aquatic biodiversity is responsible for integrating the effects caused by different abiotic factors, being a good indicator of the ecosystem's health (Wannamaker, Rice, 2000). According to Whitfield, Elliot (2002), using fishes as bioindicators is important because they have a range of available information about their life history, including different species from various trophic levels, which can reflect on the effects of stressors within that studied ecosystem. However, one of the major bottlenecks in estuarine conservation policies is the lack of information on ecology and biophysical knowledge and the degree of fish dependence on such environments (Pikitch et al. 2004; Miloslavich et al. 2011). Such knowledge is essential to support management strategies, especially for threatened, endemic, or commercially targeted species (Vilar, Joyeux, 2018).

In this study, we provide ecological fish descriptors (richness, functional guilds, and risk of extinction) along six estuaries spread along the coast of Pernambuco, describing their relationship with environmental characteristics. Additionally, we compile a large dataset and provide a list of the main estuarine fishes of Pernambuco, comparing the biodiversity of this region with those estuaries at similar latitudes. This information is relevant to access changes that can be caused by anthropogenic impacts like fisheries, pollution, climate change, and management strategies and conservation of socially-economically important species.

Material and Methods

Sampling. The study area comprehends six estuaries located along the coastal region of Pernambuco state, northeastern Brazil: the estuarine complex of Santa Cruz channel and the estuary of Jaguaribe (north coast of Pernambuco) and Suape, Maracaípe, Sirinhaém, and Rio Formoso (south coast of Pernambuco) (Fig. 1). These estuarine systems exhibit different geomorphologies and are subject to distinct degrees of anthropogenic pressures (CPRH, 2003). Sampling in each area was carried out using passive (block nets) and active fishing gears (trawling and seine nets); between 2000 and 2018 (S. Tab. 2).

The estuarine complex of Santa Cruz channel is the larger estuarine ecosystem in the state. The substrate is from clay and silt to sand along with its connections with the adjacent sea (Silva, 2004; Teixeira, Galvncio, 2010; Albuquerque et al. 2019). The Jaguaribe estuary is 9 km long and

distinguishes it from the estuarine complex of Santa Cruz by discharging directly into the ocean, being restrained to the Itamaracá Island (Paiva et al. 2009; Lima et al. 2019). The right margin of this estuary is covered by small-sized mangroves and the anthropic action is evident by the withdrew of this vegetation for the construction of shrimp's farms (El-deir 2005; Lima et al. 2019). In this region, there is Marine Protected Areas (MPAs) that includes the municipalities of Itamaracá, Itapissuma, Igarassu, and Goiana.

In the south region, Suape was initially covered by a compact coastal rainforest including vast mangroves (Lima, Costa, 1978). However, this landscape suffered several impacts, owing to the rapid growth of sugar cane plantations. Four main rivers discharge into the bay of Suape, but their course, geomorphology, and hydrodynamics have changed after the construction of the Suape port (Muniz et al. 2005). The Maracaípe estuary is bordered by dense mangroves and sandbanks, and discharges near the Pontal de Maracaípe (CPRH, 2001; Paiva et al. 2009). Sirinhaém has a complex estuary with numerous islands, lagoons, and extensive mangrove forest (CPRH, 2001). Rio Formoso estuary is formed by channels, tidal creeks, sandbanks, dense mangroves, and reefs near the coastline acting as natural barriers (Paiva et al. 2009; Lima et al. 2019). The municipality where it is located is part of the two implemented Marine Protected Areas (MPAs): Guadalupe and Costa dos Corais. Abiotic factors that may influence the distribution of species were considered in this study (Tab. 1).

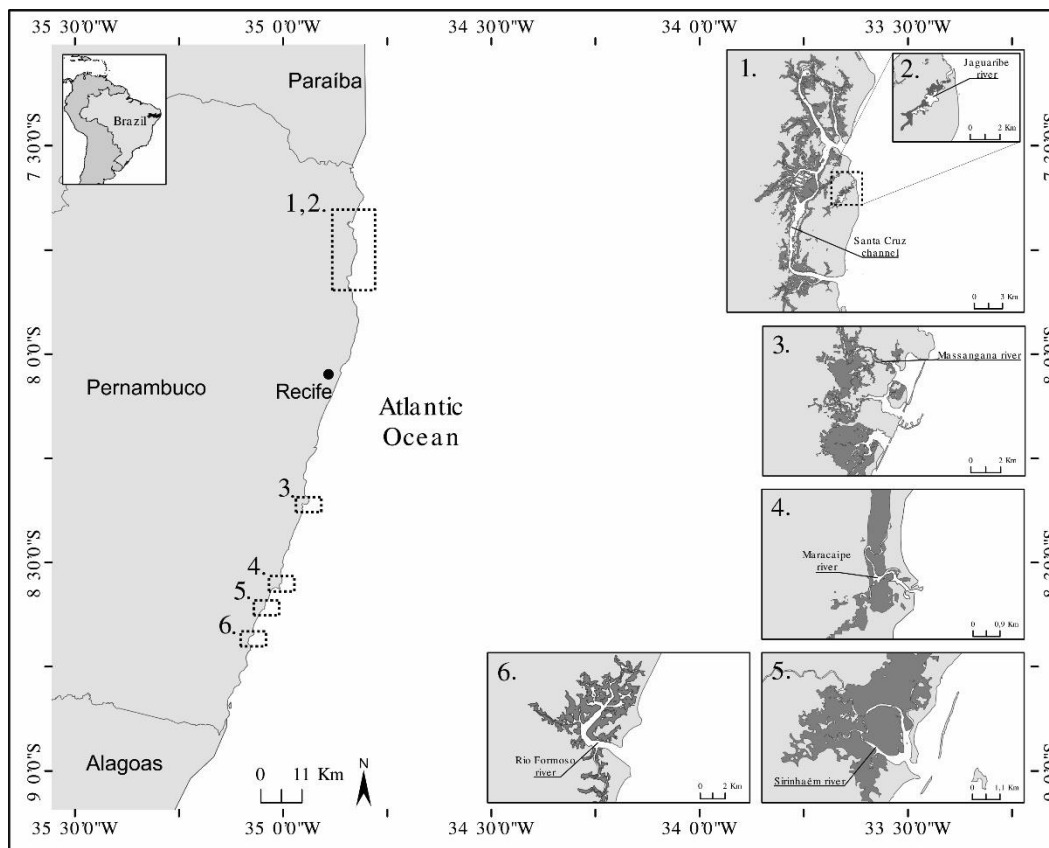


Figure I. Estuaries of Santa Cruz Channel (1), Jaguaribe (2), Suape (3), Maracaípe (4), Sirinhaém (5) and Rio Formoso (6) located in northeast Brazil. Mangrove cover is shown as darker grey areas in each panel.

Table I: Morphologic features of estuarine systems addressed in the study; northeast Brazil used in the analysis (adapted from Gonzalez et al. (2019)).

Characteristics	Estuary					
	Santa Cruz	Jaguaribe	Suape	Maracaípe	Sirinhaém	Rio Formoso
Type	Ria	Coastal Plain	Coastal Lagoon	Coastal Plain	Coastal Plain	Coastal Plain
Estuarine area (km ²) [†]	73.5	2.15	29.7	5.5	18.7	27.7
Vegetated area (km ²) [†]	48.0	1.0	23.1	5.0	17.0	19.9
Watershed area (km ²) [†]	25.5	1.15	6.6	0.5	1.7	7.8
Depth (m)	3	1.5	3.1	1.4	2.6	4.7
Mouth width (km) [†]	0.90	0.10	0.28	0.16	0.60	1.07
Numb. of mouths	2	1	1	1	1	1
Tide (mean)	1.4	2.5	3	1.5	1.2	1.5
Precipitation (2000-2018) (mean - mm)	127.7	127.7	171.8	156.8	181.9	141.2
Source	Flores Montes et al. 1998; Medeiros et al. 2001; Silva et al. 2003; Silva et al. 2011; Lana, Bernardino, 2018; APAC, 2020	Silva et al. 2011; APAC, 2020	Borges, 2011; Silva et al. 2011; Cordeiro et al. 2019; APAC, 2020	Bastos et al. 2011; Silva et al. 2011; Favero et al. 2019; APAC, 2020	Mello, 2009; Silva et al. 2009; Silva et al. 2011; APAC, 2020	Honorato da Silva et al. 2004; Silva et al. 2011; Lima, 2016; APAC, 2020

[†] Data obtained with ArcGis 10.1 based on satellite images LANDSAT 7. Type - geomorphological classification of estuaries according to Pritchard (1952). The estuary's area is considered the sum of the watershed and vegetated areas. Depth - mean depth during low tide. Mouth width - width of the

connection to the sea; mean width was considered when the estuary exhibits more than one connection to the sea. Numb. of mouths - number of connections to the sea. Tide – mean tide reported for all areas.

Data collection. The species were identified following dichotomous keys such as Araújo et al. (2004), Carpenter et al. (2003) and Figueiredo, Menezes (1980, 2000). Species richness was presented according to the systematic organization proposed by Nelson et al. (2016) with 3 levels: order, family, and species. The nomenclature of species was revised through the Eschmeyer Fish Catalogue and, when necessary, specific literature (Fricke et al. 2020). Fish species were analyzed as presence/absence data and classified to their use of estuaries based on the functional guild approach (Elliott et al. 2007).

Statistical analyses. The Estuarine Use Functional Group (EUFG) represents the use of estuaries and adjacent environments by a species over its life cycle and was classified as marine stragglers (MS), marine migrants (MM), estuarine (ES) or freshwater species (FS). The Feeding Mode Functional Group (FMFG) categorizes the feeding habit of a species: detritivore (DV), herbivore (HV), zooplanktivore (ZP), zoobenthivore (ZB), omnivore (OV), piscivore (PV), and opportunist (OP). An extensive literature review based on articles, books, and reports totalizing more than 200 references was used to define the functional guilds classification (S. Tab. 1 see Franco et al. (2008) for a detailed description of each category).

Species were assigned a conservation status according to the categories based in the International Union for Conservation of Nature (IUCN) Red List criteria of threatened species, categorized at Brazilian level (ICMbio 2018) in 10 categories: extinct (EX), regionally extinct (RE), extinct in the wild (EW), critically endangered (CR), endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

Functional guilds (EUFG and FMFG) proportions (% of species) were graphically displayed through a heatmap (estuaries x guild proportion) along with an Agglomerative Hierarchical Cluster (AHC) performed using the Ward's minimum variance method based on a Euclidian distance matrix. The linear correlation between the cophenetic and Euclidean distance matrix was calculated to validate the chosen cluster method (Pearson correlation > 0.8). These analyses were used aiming to see the similarity between estuaries based on the use of the estuary by the species as well as their feeding behavior.

A principal components analysis (PCA) was used to analyze associations between the environmental variables (Tab. 1) and the estuaries. In addition, to investigate the similarities among the fish assemblages and estuaries, a Non-metric Multidimensional Scaling (nMDS) using a Sorensen similarity matrix from species presence/absence data was performed as a cluster analysis using the average linkage method. To determine the subset of abiotic variables linked to the fish community structure, a BioEnv analysis was carried out (Oksanen et al. 2009). All data were standardized. A permutational multivariate analysis of variance (PERMANOVA) was used to test richness differences among estuaries (Clarke, Warwick, 2001). All statistical analyses were performed using the *vegan* (Oksanen et al. 2019), *ggplot2* (Wickham, 2016), and *stats* (R Core, 2019) packages from R Core Team (2020).

Results

A total of 197 fish species, distributed in 30 orders and 57 families were identified in the studied estuaries (Tab. 3). Sciaenidae was the most diverse family, comprehending 13 species, followed by Carangidae (12 species), Engraulidae (11), and Gobiidae (11). The remaining families had less than 10 species each. The estuarine complex of Santa Cruz channel exhibited higher species richness than the other estuaries, with 107 species, followed by Rio Formoso (90 species), Maracaípe (88), Sirinhaém (85), Suape (83), and Jaguaribe (72). Twenty-two species were recorded in all locations: *Achirus lineatus*, *Anchovia clupeioides*, *Archosargus rhomboidalis*, *Bairdiella ronchus*, *Caranx latus*, *Centropomus parallelus*, *C. undecimalis*, *Chaetodipterus faber*, *Diapterus auratus*, *D. rhombeus*, *Eucinostomus argenteus*, *E. melanopterus*, *Eugerres brasiliensis*, *Harengula clupeiola*, *Hyporhamphus unifasciatus*, *Lutjanus alexandrei*, *L. jocu*, *Mugil curema*, *Oligoplites saurus*, *Sphoeroides testudineus*, *Sphyraena barracuda* and *Strongylura timucu*.

Most fishes (79.8%) were classified as Least Concern according to the ICMbio conservation status (Tab. 3). However, nine species were identified under some degree of threat: one as CR (*Epinephelus itajara* (Rio Formoso)) and nine as VU (*Achirus mucuri* (Sirinhaém), *Hippocampus erectus* (Santa Cruz Channel, Suape and Maracaípe), *Hippocampus reidi* (Maracaípe and Rio Formoso), *Megalops atlanticus* (Jaguaribe, Sirinhaém, and Rio Formoso), *Mycteroperca bonaci* (Santa Cruz Channel), *Lutjanus cyanopterus* (Jaguaribe, Sirinhaém, Maracaípe, and Rio Formoso), *Sciades parkeri* (Sirinhaém), and *Sparisoma axillare* (Suape and Rio Formoso)). It was also observed nine species classified as NT: *Balistes vetula* (Maracaípe), *Cynoscion acoupa* (Sirinhaém), *Hyporhamphus unifasciatus* (in all studied estuaries), *Mugil liza* (Santa Cruz Channel, Jaguaribe, Suape, Maracaípe and Rio Formoso), *Lutjanus analis* (Santa Cruz Channel and Suape),

L. jocu (in all studied estuaries), *L. synagris* (Santa Cruz Channel, Jaguaribe, Suape and Rio Formoso), *Ocyurus chrysurus* (Rio Formoso), and *Sparisoma amplum* (Jaguaribe). In addition, eighteen species were recognized as DD (Tab. 3).

Fish species exhibited a strong affinity to the marine ecosystem, most of them belonging to the categories marine stragglers (46.6%) and marine migrants (32.3% of the total number of species) (S. Tab. 1, Fig. 2), a pattern observed in every estuary. The contribution of marine stragglers and marine migrants to the overall fish composition varied between 75 to 79% among estuaries. Estuarine species were 20.5% of the total number and only one freshwater species (*Poecilia vivipara*) was observed in one of the studied locations (Jaguaribe).

Most of the species fed on invertebrates associated to the substrate (*i.e.* zoobenthivorous, 50.7%), followed by piscivorous (21.4%) and zooplanktivorous fishes (10.4%; Supplementary Material Tab. 1) in all estuaries. This pattern of feeding habits was observed for each estuary alone, with small differences between them and, whenever present, it was associated to variations in the number of species in the detritivore, zooplanktivore, and omnivore categories (Fig. 3).

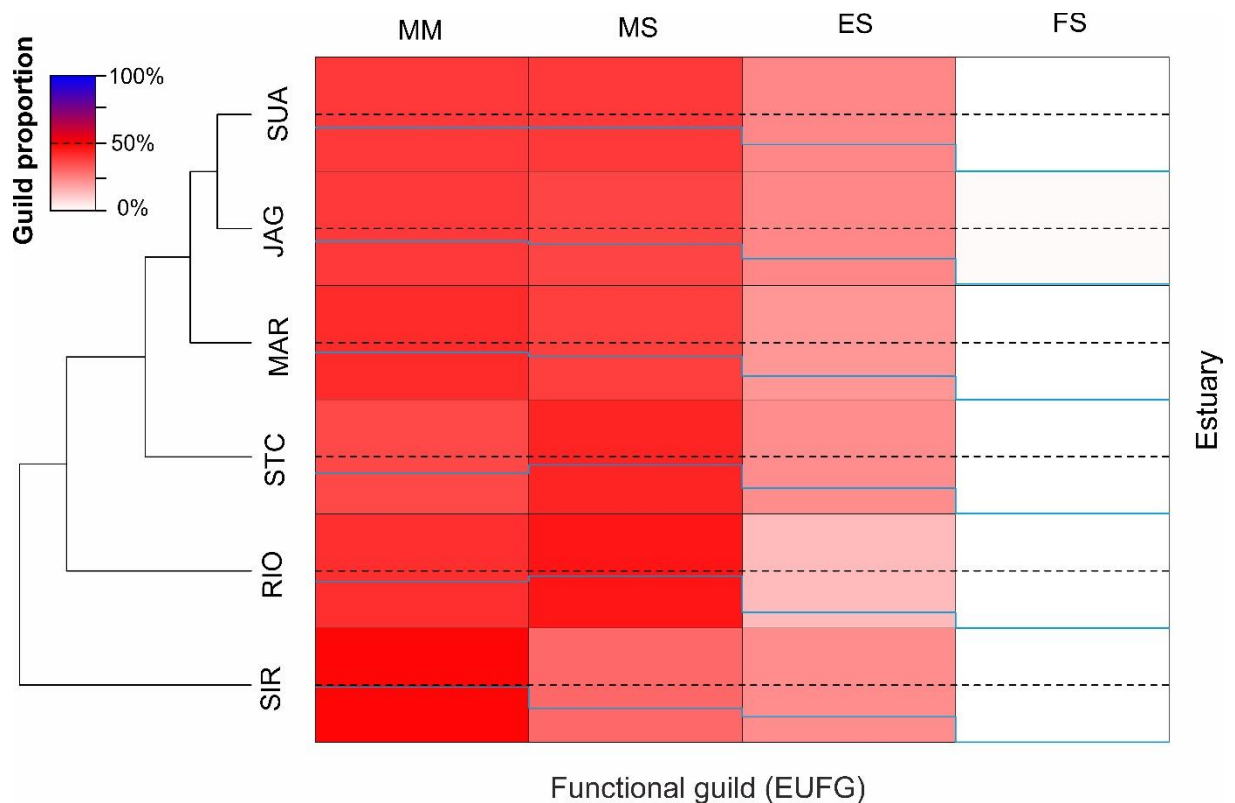


Figure II. Percentage of each EUFG category marine migrant (MM), marine stragglers (MS), estuarine fishes (ES) and freshwater species (FS) for the estuaries of Sirinhaém (SIR), Rio Formoso (RIO), Santa Cruz channel (STC), Maracaípe (MAR), Jaguaribe (JAG) and Suape (SUA) located in

northeast Brazil. Blue line: percentage of each category within the estuaries. Dotted line: 50 percentage of the total number.

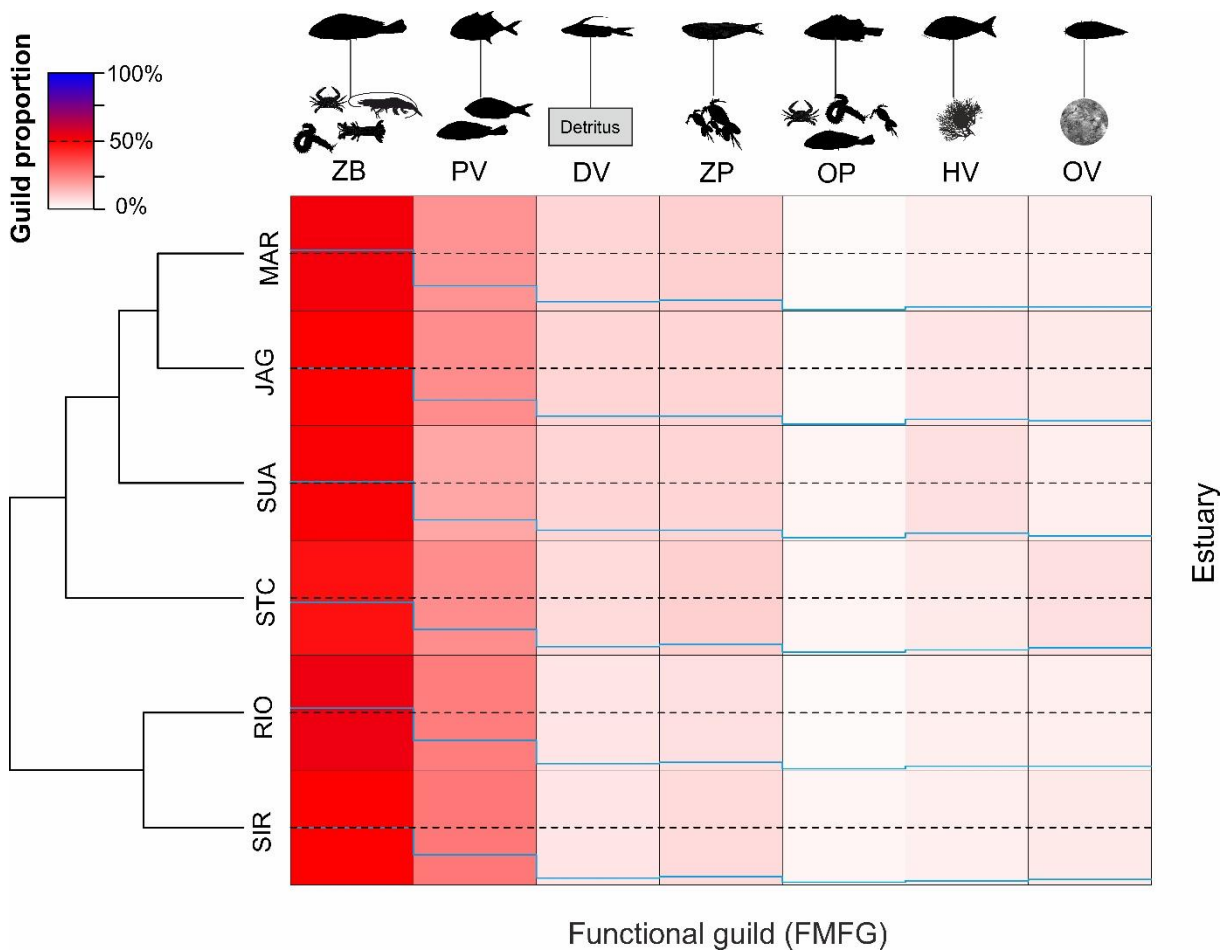


Figure III. Percentage of each FMFG category zoobenthivore (ZB), piscivore (PV), zooplanktivore (ZP), herbivore (HV), omnivore (OV), detritivore (DV) and opportunistic (OP) for the estuaries of Sirinhaém (SIR), Rio Formoso (RIO), Santa Cruz channel (STC), Maracaípe (MAR), Jaguaribe (JAG) and Suape (SUA) located in northeast Brazil. Blue line: percentage of each category within the estuaries. Dotted line: 50 percentage of the total number.

A total of 76.5% of the variance was explained by the two first axis (PC1 – 58.5% and PC2 – 17.9%) of the PCA analysis. The greater proportion of explanation for the PC1 was related to three variables (watershed, estuarine and vegetated areas) that are positively associated with the Santa Cruz Channel's estuary. At the same time, mouth width and depth are positively linked to the Rio Formoso estuary. On the other hand, the tide and precipitation variables have little contribution to the explanation of PC1 being negatively correlated with the estuaries of Santa Cruz channel and Rio Formoso in overall. In the PC2, the largest contribution is related to the variables tide (PC2;

eigenvalues= 0.48) associated with the estuaries of Jaguaribe and Suape, and the precipitation (PC2; eigenvalues= -0.57) to Maracaípe and Sirinhaém (Fig. 4).

The nMDS and the Cluster analysis (Fig. 5) showed similar results, both based on the taxonomic composition of each estuary correlated with the abiotic data. The cluster analysis exhibited three groups with a resemblance level of 41%, which showed a significant difference between the Jaguaribe estuary and the other ones. Also, it is possible to distinguish the formation of two groups: the first one comprised the estuaries of Santa Cruz, Rio Formoso, and Sirinhaém, and the second one, Maracaípe, and Suape. The nMDS plotted with the abiotic characteristics (Fig. 5) showed the relationship between these attributes and the patterns observed in the cluster analysis (Fig. 5), evidencing that the Jaguaribe estuary differed from the other estuaries mainly by the tide. The BioEnv showed that the best correlation within the abiotic and biotic characteristics was between tide and precipitation (Tab. 2).

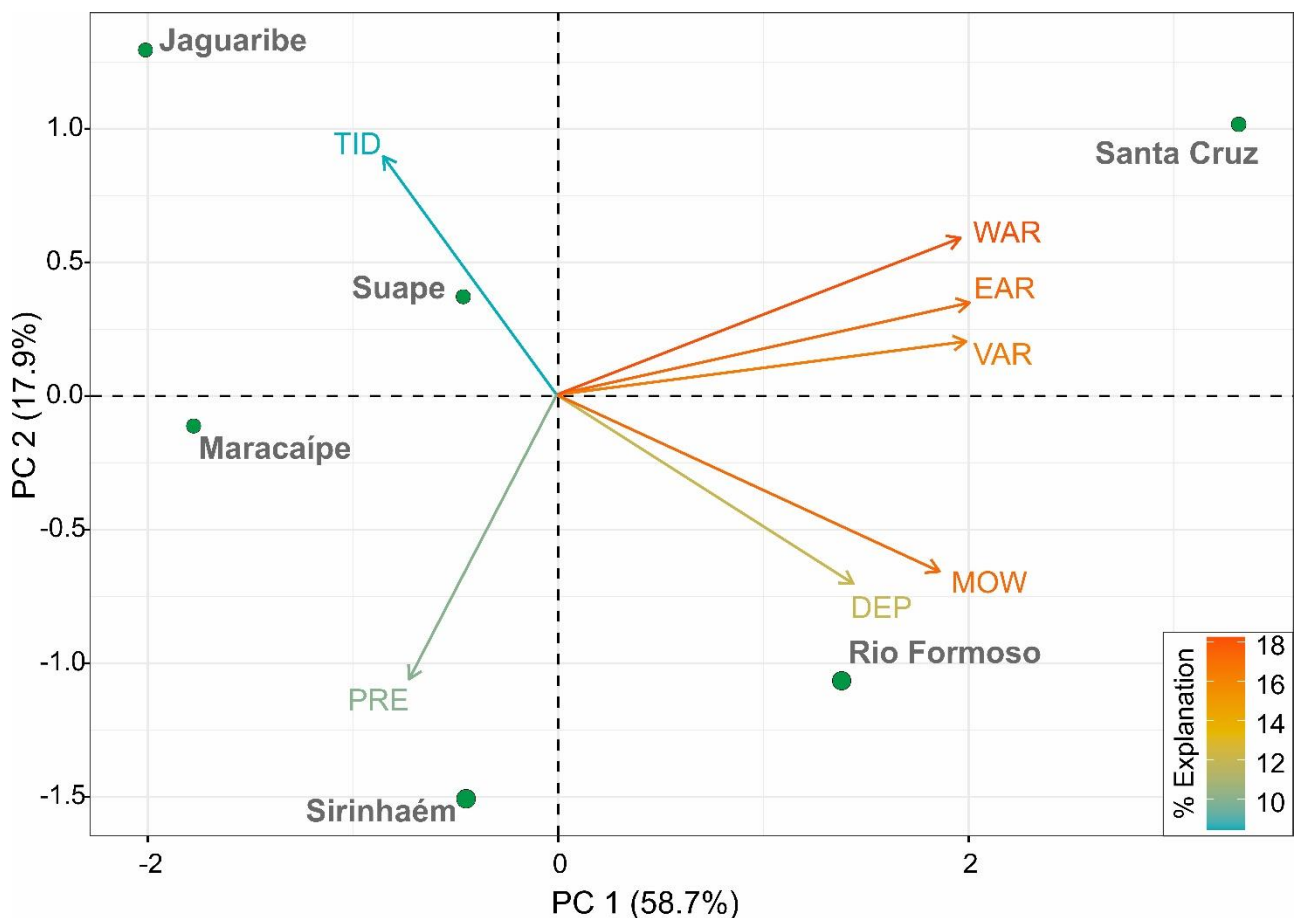


Figure IV. Principal components analysis (PCA) based on abiotic data of each estuary. TID: tide; PRE: precipitation; WAR: watershed area; EAR: estuarine area; VAR: vegetated area; MOW: mouth width; DEP: depth

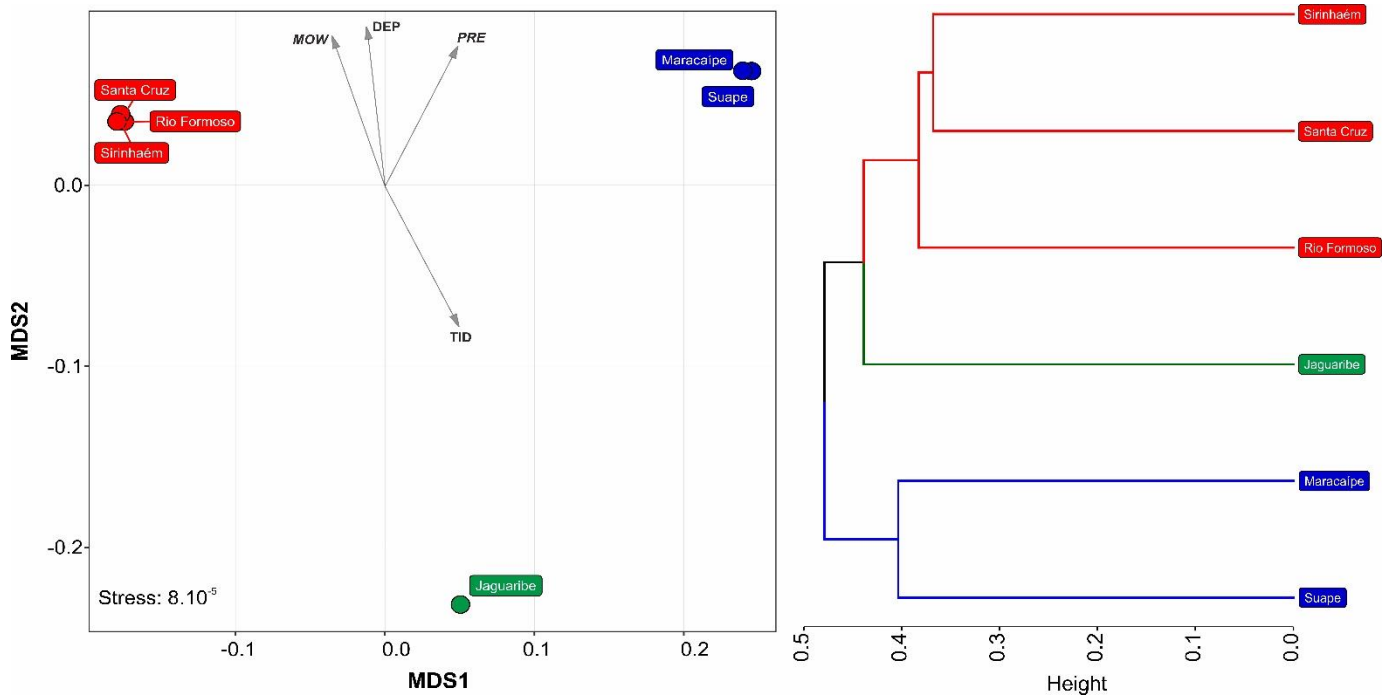


Figure V. Non-metric Multidimensional Scaling (nMDS) based on species records and abiotic characteristics of the estuaries and a cluster analysis based on species records by estuaries. MOW: mouth width; DEP: depth; PRE: precipitation; TID: tide

Table II: Results from the BioEnv – Abiotic and Biotic Matrixes

* Best correlation	Size	Correlation
Tide and Precipitation*	2	0.5079
Tide	1	0.3790
Depth; Tide and Precipitation	3	0.2439
Depth; Mouth Width; Tide and Precipitation	4	0.0409

Discussion

Among several definitions, estuaries have been commonly regarded as partly enclosed areas with a free connection with the open sea, creating salinity gradients along the mixing zone (Able, Fahay,

2010). Estuaries provide a wide diversity of habitats and wildlife with significant commercial and recreational benefits for the population (Blaber, 2000). They are highly susceptible to different variations in chemical or physical factors, which may affect the composition and distribution of local assemblages and critical transitional areas for the ichthyofauna (Blaber, 2000; Ray, 2005; Favero et al. 2019).

Pernambuco has around fifteen estuaries along its coast (Silva et al. 2011; Pelage et al. 2019), where six of the most productive are investigated in this present study. Based on our findings, the estuarine ichthyofauna of Pernambuco is rich and diverse, with the occurrence of 197 species and 57 families. Other studies analyzed or compiled information about the estuaries in Pernambuco: Vasconcelos Filho, Oliveira (1999) studied a single estuary and observed 145 species; Mérigot et al. (2017) in four estuaries reported 122 species, and Paiva, Araújo (2010) analyzed 17 estuaries, found a similar number (210) and dominance of species to the present study. Nonetheless, the number of species observed in adjacent areas such as Mamanguape estuary (Paraíba, Brazil, 66 species and 33 families; Medeiros et al. 2018) and Santo Antônio River estuary (Alagoas, Brazil, 34 species and 16 families; da Silva et al. 2018). The variation found in the species number observed at these locations may be attributed to the variability on estuary's size and sampling strategies, as well as some possible divergence in taxonomic classification. Some common families such as Mugilidae are speciose and species composition has only recently been revised (Menezes et al. 2015).

The use of multiple fishing gear can improve the catch of species and thus the observed image of biodiversity (Gotelli, Colwell, 2011). Therefore, to provide a piece of robust information on the estuarine biodiversity of Pernambuco, we combined information of multiple fishing gears (passive and active) used in different habitats, such as channels, flooded mangroves, shallow coastal areas, and sand or mud banks.

Abiotic conditions of the estuaries also play an essential role in the fish assemblage. The studied estuaries totalized 158 km of extension and have different sizes and morphology, relatively smaller compared to other Brazilian estuaries. For instance, the largest estuary in the state of Pernambuco (Santa Cruz) covers an area of approximately 73.5 km² and exhibits a total number of 107 species. On the other hand, for one of the largest estuaries in northeast Brazil, the Jaguaribe – Ceará, with an area of approximately 641,216 km², only 129 species have been reported (ZEE, 2005; Costa, 2013). Such richness found in Pernambuco can be explained by their different areas and habitats (Ferreira et al. 2019).

Among the estuaries of Pernambuco, the most common fish species are mullets (Mugilidae), catfishes (Ariidae), jacks (Carangidae), snooks (Centropomidae), and anchovies (Engraulidae) (Lessa et al. 2006; Lira et al. 2018). The present study found 22 common species between all studied sites mainly belonging to Gerreidae (5), Engraulidae (2), Carangidae (2) and Centropomidae (2), similar to what it was reported in the same estuaries or nearby ones: Castro (2001) (Paciência River, Maranhão); Paiva et al. (2008) (Rio Formoso, Pernambuco); Neta and Castro (2008) (Ilha de Caranguejos, Maranhão); Lira et al. (2018) (Sirinhaém estuary, Pernambuco); Favero et al. (2019) (Maracaípe estuary, Pernambuco) and Ferreira et al. (2019) (Itapissuma/Itamaracá complex, Pernambuco). However, three clusters divided the studied estuaries based in the fish community. The first group was Sirinhaém, Rio Formoso, and Santa Cruz, while the second was formed by Jaguaribe estuary and the third by Suape and Maracaípe. The climate pattern has a higher potential to control biological community dynamics (Vasconcelos et al. 2015; Fujiwara et al. 2019). The precipitation was larger in the south area (Sirinhaém, Rio Formoso, Suape and Maracaípe) than in the north area (Santa Cruz and Jaguaribe) and, according to our BIOENV data, in general, the fish assemblage was correlated mainly with the precipitation and tide patterns. The control of rainfall is not only driven by the increased freshwater inflow, but also by indirect processes like a high concentration of nutrients and consequently the highest primary productivity (PP) during the rainy seasons (Caffrey et al. 2007; Parizzi et al. 2016). The effect of the rainfall in regions of minor tidal variations may favor the development of the PP and consequently in the biotic community (Bucci et al. 2012). This can be observed in the estuaries of Sirinhaém, Rio Formoso, and Santa Cruz. Although these estuaries have morphological structure differences and dimensions, the rainfall rates and/or the reduced tidal variations were considered crucial in defining the species richness.

The results discussed in the previous paragraph were different from what reported by Vorwerk et al. (2003), which attested that the mouth size of an estuary mainly influences the structure of a fish assemblage in an estuary. According to Kuparinen et al. (2009) and Bezerra et al. (2012), lunar cycles can directly influence the tide changes, affecting the feeding behavior and movement of fishes, such as their vertical distribution. The river flow may also drive fish assemblages since it maintains the detrital and nutrient inputs into estuaries, preventing hypersaline conditions and providing a favorable environment to these species (Blaber, Blaber, 1980; Whitfield, 1994; Meynecke et al. 2006). The size of an estuary can also directly influence the fish assemblages (Mérigot et al. 2017). Jaguaribe is the smallest one in terms of extension and mangrove coverage within all the assessed estuaries. Also, it is known to have suffered strong siltation processes in its

mouth over time (Silva-Falcão et al. 2007), reducing its depth and making it difficult to exchange fresh and seawater, therefore, potentially decreasing the diversity of marine species. The lowest species richness was reported in the Jaguaribe estuary.

Suape and Maracaípe had a higher number of common species (52 species), thus, forming the last cluster. However, these estuaries also had exclusive species. Morphologically, these estuaries are different, and their rivers have a low potential for water discharge (Bastos et al. 2011). Maracaípe is a coastal plainland river with an open estuary and a direct connection to a coral reef barrier.

Therefore, reef specie, such as *Acanthurus bahianus*, *Balistes vetula*, and *Pomacanthus paru* are more frequent and thus represented the catches. On the other hand, Suape is a coastal lagoon that suffered a high deforestation impact due to the construction of a large industrial port complex (chemical, shipping, and logistics companies), with predictable adverse effects on biodiversity and fisheries productivity (Paiva, Araújo, 2010). However, this was not reflected in the total observed species richness, which was high (83 species), similar to the other estuaries.

The guild approach, particularly the trophic and estuarine use guilds, have been widely applied for understanding the structure and functioning of aquatic ecosystems, the movement pattern between environments, and their use as feeding, breeding, or development grounds (Elliott et al. 2007; Passos et al. 2013). In this study, most species were marine stragglers and migrants, given the permanent connection between the estuaries and the Atlantic Ocean over the year (Andrade-Tubino et al. 2008). Moreover, marine stragglers and migrants are commonly represented along estuaries around the world (Franco et al. 2008), since estuaries have an essential role as breeding areas, and fish use these environments to spawn, to complete their life cycles, or even to do migratory movements (Vendel, Chaves, 2006).

According to Duarte, Andreato (2003), the type of sediment found in an estuary has a huge influence in the distribution and regulation of organisms and their community. Demersal fishes have unique importance in marine ecosystems due to their feeding and reproductive behavior being correlated with the benthic ecosystem. Furthermore, the zoobenthivores fishes perform a critical role within the primary consumers and the higher levels found in the food web, which can be explained by a high abundance of benthic fauna (Silva, 2013; Passarone et al. 2019). Benthos is largely mainly responsible for structuring the food web and having an important role in the system dynamics (Herman et al. 1999). Benthivores dominate in all types of estuaries (Blaber, 2000).

Most of the species reported in the estuaries of Pernambuco were classified as least concern (LC) according to the IUCN conservation status. However, important commercial species were classified

as vulnerable (VU), near threatened (NT), data deficient (DD), or even not evaluated (NE). The estuary of Rio Formoso presented most of the species (24) classified within the previously mentioned categories. Within the important commercial species, additional attention should be given to *E. itajara* (CR), since it is under an active fishing prohibition until 2023 (Pereira 2018); and to *Megalops atlanticus* (VU), *Lutjanus cyanopterus* (VU), *Mugil liza* (NT), *Hyporhamphus unifasciatus* (NT) and *Mycteroperca bonaci* (VU), given their economic importance (Lessa and Nóbrega 2000; Gaspar 2006; Falcão 2007; Favero et al. 2019). *Sphoeroides testudineus* (DD), *Pomacanthus arcuatus* (DD), and *Pomacanthus paru* (DD) are notable components of coral reefs and seagrass ecosystems (Marty, 2014; Eduardo et al., 2018; Eduardo et al. 2020) and exemplifies poorly studied species deserve further attention for conservation and management, even though they are not considered threatened species.

Currently, most results about the biodiversity found in estuaries worldwide are still restricted to 'grey literature' or scattered into different data types (Blaber, 2000; König et al. 2019). Compiling this data is important not only to improve access to information, but also to carry out integrated studies that allow understanding the taxonomic and functional variation and how estuaries provide advantages for the protection and development of species (Medeiros et al. 2001; König et al. 2019). A compilation of the fish biodiversity found at six of the most productive estuaries in Pernambuco and its correlation between some abiotic characteristics were provided, focusing on the contribution to fisheries management and the conservation of the ichthyofauna. Our findings indicate that the Santa Cruz Complex had the highest species richness, while Jaguaribe had the lowest. Even though the abiotic characteristics morphology and size are significant, the variables precipitation and tide range were crucial to identify richness patterns within the studied estuaries. Also, estuarine fish assemblages mainly comprise species that spend only part of their lives in the estuaries, mainly feeding on benthic invertebrates. This reinforces the contribution role of marine waters in maintaining the biodiversity in estuaries and how possible changes in morphology or weather patterns can affect the estuaries showing its connectivity.

Estuaries consist of biologically varied subsystems, one of the most natural and productive systems in the biosphere (Dame, 2008). Although it is considered resilient, this ecosystem is affected by changes in marine or terrestrial environments, such as variations in wind patterns, evaporation, precipitation, and temperatures, which may significantly impact species distribution and assemblages (James et al. 2013). Even though the estuaries in Pernambuco are highly compromised because of the daily impacts like aquaculture and industries (Santa Cruz), port (Suape), and sugarcane plantation (Sirinhaém), they still have a high ichthyofaunal richness.

However, the functionality and the dominance of fishes can be altered because of the direct anthropogenic impacts, which changes the geomorphology and hydrodynamic conditions of the areas and alters the number and composition of species (Kennish, 2005; Pelage et al. 2019), hence the necessity of the validation of environmental degradation in monitoring programs.

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Author contributions

RSL, JGG, LNE, ASL, LP, VF, CC, and IMSA conducted the sampling and laboratory analysis; RSL, JGG and ASL performed the statistical analysis; RSL, ASL, JGG, VF, LP and CCM prepared the figures and tables; BP, MEA and ACAE contributed with data from some of the estuaries; and RSL, ASL, JGG, LNE, FLF, TF, BP and WS wrote the manuscript.

Table III: List of fish species recorded in six estuaries in north-eastern Brazil, along with their conservation status (IUCN list) and respective feeding mode (FMFG) and estuarine use (EUFG) functional groups (Elliot et al., 2007). Estuaries: Santa Cruz channel (STC), Jaguaribe (JAG), Suape (SUA), Maracaípe (MAR), Sirinhaém (SIR) and Rio Formoso (RIO). IUCN classification: NE - not evaluated; DD - data deficient; LC - least concern; VU - vulnerable; NT - near threatened; CR - critically endangered; feeding mode functional groups (FMFG): ZB - zoobenthivore; PV - piscivore; ZP - zooplanktivore; HV - herbivore; OV - omnivore; DV - detritivore; OP - opportunistic, estuarine use functional group (EUFG): MM - marine migrants; MS - marine straggler; ES - estuarine species; FS - freshwater species.

Order	Family	Species	IUCN †	FMFG ‡	EUFG ‡	Estuary						
						STC	JAG	SUA	MAR	SIR	RIO	
Myliobatiformes	Dasyatidae	<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)	DD	ZB	MM						X	
		<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	LC	ZB	MS	X	X			X	X	
Elopiformes	Elopidae	<i>Elops saurus</i> Linnaeus, 1758	NE	PV	MS	X	X					
		<i>Megalops atlanticus</i> Valenciennes, 1847	VU	PV	MM		X			X	X	
Albuliformes	Albulidae	<i>Albula vulpes</i> (Linnaeus, 1758)	DD	ZB	MM		X		X		X	
		<i>Gymnothorax funebris</i> Ranzani, 1839	DD	ZB	MS		X			X	X	
		<i>Gymnothorax ocellatus</i> Agassiz, 1831	DD	ZB	MS	X						
Anguilliformes	Ophichthidae	<i>Ahlia egmontis</i> (Jordan, 1884)	LC	ZB	MS			X				
		<i>Myrichthys ocellatus</i> (Lesueur, 1825)	LC	ZB	MS	X					X	
		<i>Myrophis punctatus</i> Lütken, 1852	LC	ZB	MM		X		X			
		<i>Ophichthus cylindroideus</i> (Ranzani, 1839)	LC	PV	ES				X			
		<i>Ophichthus ophis</i> (Linnaeus, 1758)	LC	PV	MS				X			
Clupeiformes	Congridae	<i>Ariosoma balearicum</i> (Delaroche, 1809)	LC	-	MS			X				
	Pristigasteridae	<i>Chirocentron bleekermanus</i> (Poey, 1867)	LC	ZB	MS	X						
	Engraulidae	<i>Anchoa filifera</i> (Fowler, 1915)	LC	ZB	ES			X				
		<i>Anchoa januaria</i> (Steindachner, 1879)	LC	ZP	ES			X		X		
		<i>Anchoa lyolepis</i> (Evermann & Marsh, 1902)	LC	ZP	MS	X						
		<i>Anchoa marinii</i> Hildebrand, 1943	LC	ZP	MS	X						
		<i>Anchoa spinifer</i> (Valenciennes, 1848)	LC	PV	MM	X		X	X	X		
		<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)	LC	ZB	MM	X		X	X	X		
		<i>Anchovia clupeoides</i> (Swainson, 1839)	LC	ZP	MM	X	X	X	X	X	X	
		<i>Anchoviella lepidentostole</i> (Fowler, 1911)	LC	ZB	MM		X		X			
		<i>Cetengraulis edentulus</i> (Cuvier, 1829)	LC	ZP	MM	X			X	X	X	
		<i>Engraulis anchoita</i> Hubbs & Marini 1935	LC	ZP	MS	X						
		<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	LC	PV	ES	X	X	X		X	X	
		<i>Harengula clupeola</i> (Cuvier, 1829)	LC	ZP	MS	X	X	X	X	X	X	
		<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)	LC	HV	MM	X	X	X	X	X		
		<i>Opisthonema oglinum</i> (Lesueur, 1818)	LC	ZP	MS	X	X			X	X	
		<i>Platanichthys platana</i> (Regan, 1917)	LC	ZP	ES	X		X				
		<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)	LC	ZP	ES	X						
		Ariidae	<i>Aspistor luniscutis</i> (Valenciennes, 1840)	LC	OV	MS	X				X	
			<i>Aspistor quadriscutis</i> (Valenciennes, 1840)	LC	ZB	MS					X	
<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)	LC		ZB	ES	X				X	X		
<i>Cathorops spixii</i> (Agassiz, 1829)	LC		ZB	ES				X	X			
<i>Notarius grandicassis</i> (Valenciennes, 1840)	LC		ZB	MM					X			
<i>Sciades herzbergii</i> (Bloch, 1794)	LC		ZB	ES	X				X	X		
<i>Sciades parkeri</i> (Traill, 1982)	VU		ZB	ES					X			
<i>Sciades proops</i> (Valenciennes, 1840)	DD		ZB	ES						X		

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Aulopiformes	Synodontidae	<i>Synodus foetens</i> (Linnaeus, 1766)	LC	PV	MS	X		X	X		
Holocentriformes	Holocentridae	<i>Holocentrus adscensionis</i> (Osbeck, 1765)	LC	ZB	MS			X			
Batrachoidiformes	Batrachoididae	<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)	LC	ZB	MS	X					
		<i>Thalassophryne nattereri</i> Steindachner, 1876	LC	ZB	MS	X	X	X	X		X
Kurtiformes	Apogonidae	<i>Phaeoptyx pigmentaria</i> (Poey, 1860)	LC	ZP	MS			X			
Gobiiformes	Eleotridae	<i>Eleotris pisonis</i> (Gmelin, 1789)	LC	PV	ES						X
		<i>Erotelis smaragdus</i> (Valenciennes, 1837)	LC	-	ES			X			
		<i>Guavina guavina</i> (Valenciennes, 1837)	LC	ZB	ES	X	X			X	X
	Gobiidae	<i>Bathygobius soporator</i> (Valenciennes, 1837)	LC	PV	ES	X	X	X	X		
		<i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882)	LC	DV	ES	X	X	X	X	X	
		<i>Ctenogobius saepepallens</i> (Gilbert & Randall, 1968)	LC	ZB	MS			X			
		<i>Ctenogobius shufeldti</i> (Jordan & Eigenmann, 1887)	LC	OV	ES	X					
		<i>Ctenogobius smaragdus</i> (Valenciennes, 1837)	LC	DV	ES	X	X	X			
		<i>Ctenogobius stigmaticus</i> (Poey, 1860)	LC	DV	ES	X					
		<i>Evorthodus lyricus</i> (Girard, 1858)	LC	DV	MS	X					
		<i>Gobioides broussonnetii</i> Lacepède, 1800	LC	DV	ES				X		
		<i>Gobionellus oceanicus</i> (Pallas, 1770)	LC	DV	ES	X	X		X	X	X
		<i>Gobionellus stomatus</i> Starks, 1913	LC	DV	ES	X	X	X			
		<i>Microgobius meeki</i> Evermann & Marsh, 1899	LC	ZB	MS	X					
Mugiliformes	Mugilidae	<i>Mugil brevirostris</i> (Ribeiro, 1915)	NE	-	MM			X			
		<i>Mugil curema</i> Valenciennes, 1836	DD	DV	MM	X	X	X	X	X	X
		<i>Mugil curvidens</i> Valenciennes, 1836	DD	DV	-			X	X	X	X
		<i>Mugil incilis</i> Hancock, 1830	LC	DV	MM			X	X	X	
		<i>Mugil liza</i> Valenciennes, 1836	NT	DV	MM	X	X	X	X		X
		<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007	DD	-	MM	X		X			X
Blenniiformes	Dactyloscopidae	<i>Dactyloscopus crossotus</i> Starks, 1913	LC	-	MS				X		
Atheriniformes	Atherinopsidae	<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)	LC	OV	ES	X	X	X	X	X	
		<i>Atherinella cf. blackburni</i> (Schultz, 1949)	LC	ZP	MM				X		
Beloniformes	Hemiramphidae	<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)	LC	HV	MS	X		X		X	
		<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)	LC	HV/OP	MM		X	X			
		<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)	NT	OP	MM	X	X	X	X	X	X
	Belonidae	<i>Ablennes hians</i> (Valenciennes, 1846)	LC	PV	MS				X		
		<i>Strongylura marina</i> (Walbaum, 1792)	LC	PV	MM				X	X	
		<i>Strongylura timucu</i> (Walbaum, 1792)	LC	PV	MM	X	X	X	X	X	X
		<i>Tylosurus acus acus</i> (Lacepède, 1803)	LC	PV	MS	X			X		
		<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)	LC	PV	MM				X		

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Cyprinodontiformes	Poeciliidae	<i>Poecilia vivipara</i> (Bloch & Schneider, 1801)	LC	ZP	FS			X					
Carangiformes	Echeneidae	<i>Echeneis naucrates</i> Linnaeus, 1758	LC	DV	MS							X	
	Carangidae	<i>Carangoides bartholomaei</i> (Cuvier, 1833)	LC	PV	MS				X	X	X	X	X
		<i>Caranx crysos</i> (Mitchill, 1815)	LC	PV	MS	X	X			X	X	X	X
		<i>Caranx hippos</i> (Linnaeus, 1766)	LC	PV	MS	X	X			X	X	X	X
		<i>Caranx latus</i> Agassiz, 1831	LC	ZB	MS	X	X	X	X	X	X	X	X
		<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	LC	ZB	MS	X					X		
		<i>Oligoplites palometa</i> (Cuvier, 1832)	LC	PV	MM	X	X				X	X	X
		<i>Oligoplites saliens</i> (Bloch, 1793)	LC	PV	MM	X					X	X	X
		<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	LC	PV	MM	X	X	X	X	X	X	X	X
		<i>Selene brownii</i> (Cuvier, 1816)	LC	ZB	MS								X
		<i>Selene vomer</i> (Linnaeus, 1758)	LC	PV	MS	X	X				X		X
		<i>Trachinotus carolinus</i> (Linnaeus, 1766)	LC	ZB	MM								X
		<i>Trachinotus falcatus</i> (Linnaeus, 1758)	LC	ZB	MS						X		X
Istiophoriformes	Sphyraenidae	<i>Sphyraena barracuda</i> (Edwards, 1771)	LC	PV	MM	X	X	X	X	X	X	X	
		<i>Sphyraena guachancho</i> Cuvier, 1829	LC	PV	MS	X				X		X	
Pleuronectiformes	Paralichthyidae	<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900	LC	ZB	ES		X	X	X			X	
		<i>Citharichthys macrops</i> Dresel, 1885	LC	ZP	MS					X			
		<i>Citharichthys spilopterus</i> Günther, 1862	LC	ZB	ES	X		X	X	X	X		
		<i>Etropus crossotus</i> Jordan & Gilbert, 1882	LC	ZB	MM	X		X	X				
		<i>Etropus longimanus</i> Norman, 1933	LC	ZB	MS	X							
		<i>Paralichthys brasiliensis</i> (Ranzani, 1842)	LC	ZB	MM	X	X	X	X			X	
		<i>Syacium micrurum</i> Ranzani, 1842	LC	ZB	MM			X					
		Bothidae	<i>Bothus ocellatus</i> (Agassiz, 1831)	LC	ZB	MM			X	X			X
			<i>Bothus robinsi</i> Topp & Hoff, 1972	LC	ZB	MS				X			
		Achiriidae	<i>Achirus achirus</i> (Linnaeus, 1758)	LC	ZB	ES	X					X	X
			<i>Achirus declivis</i> Chabanaud, 1940	LC	ZB	ES	X			X	X	X	X
			<i>Achirus lineatus</i> (Linnaeus, 1758)	LC	ZB	ES	X	X	X	X	X	X	X
			<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009	VU	-	-						X	
<i>Trinectes microphthalmus</i> Chabanaud, 1928	LC		ZB	MM						X			
<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	LC		ZB	MM	X			X	X	X			
Cynoglossidae	<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)	LC	ZB	MM			X						
	<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)	LC	ZB	ES		X		X					
	<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)	LC	ZB	ES	X		X	X	X	X	X		
Syngnathiformes	Syngnathidae	<i>Bryx dunckeri</i> (Metzelaar, 1919)	LC	ZP	ES				X				
		<i>Cosmocampus elucens</i> (Poey, 1868)	LC	ZB	MS		X	X					
		<i>Hippocampus erectus</i> Perry, 1810	VU	ZB	MS	X		X	X	X			
		<i>Hippocampus reidi</i> Ginsburg, 1933	VU	ZP	MM				X		X		

Microphis brachyurus lineatus (Kaup, 1856)

NE

ZP

ES

X

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			NE	ZP	ES	X					
Syngnathiformes	Syngnathidae	<i>Syngnathus pelagicus</i> Linnaeus, 1758	LC	ZB	MS		X	X	X		
	Fistulariidae	<i>Fistularia petimba</i> Lacepède, 1803	LC	PV	ES		X	X			
		<i>Fistularia tabacaria</i> Linnaeus, 1758	LC	PV	MS		X	X			
	Dactylopteridae	<i>Dactylopterus volitans</i> Linnaeus, 1758	LC	ZB	MS		X		X	X	
Scombriformes	Trichiuridae	<i>Trichiurus lepturus</i> Linnaeus, 1758	LC	PV	MS					X	X
	Scombridae	<i>Scomberomorus brasiliensis</i> Collete, Russo & Zavala-Camin, 1978	LC	PV	MS		X				X
		<i>Scomberomorus regalis</i> (Bloch, 1793)	LC	PV	MS						X
	Stromateidae	<i>Peprilus paru</i> (Linnaeus, 1758)	LC	OV	MS	X					
Trachiniformes	Uranoscopidae	<i>Astroscopus y-graecum</i> Cuvier, 1829	LC	PV	MS				X		
Labriiformes	Scaridae	<i>Sparisoma amplum</i> (Ranzani, 1842)	NT	HV	MS		X				
		<i>Sparisoma axillare</i> (Steindachner, 1878)	VU	HV	MS				X		X
		<i>Sparisoma radians</i> (Valenciennes, 1840)	LC	HV	MS	X			X		X
Perciformes	Centropomidae	<i>Centropomus ensiferus</i> Poey, 1860	LC	PV	-					X	
		<i>Centropomus mexicanus</i> Bocourt, 1868	LC	PV	-				X		X
		<i>Centropomus parallelus</i> (Poey, 1860)	LC	PV	MM	X	X	X	X	X	X
		<i>Centropomus pectinatus</i> Poey, 1860	LC	PV	MM	X		X	X	X	X
		<i>Centropomus undecimalis</i> (Bloch, 1792)	LC	PV	MM	X	X	X	X	X	X
	Gerreidae	<i>Diapterus auratus</i> Ranzani, 1842	LC	ZB	MM	X	X	X	X	X	X
		<i>Diapterus rhombeus</i> (Cuvier, 1829)	LC	ZP	MM	X	X	X	X	X	X
		<i>Eucinostomus argenteus</i> Baird & Girard, 1855	LC	ZB	MM	X	X	X	X	X	X
		<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	LC	ZB	MM	X	X	X	X	X	
		<i>Eucinostomus havana</i> (Nichols, 1912)	LC	ZB	MM	X		X	X		
		<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	LC	ZB	MM	X	X	X	X	X	X
		<i>Eugerres brasilianus</i> (Cuvier, 1830)	LC	OV	MM	X	X	X	X	X	X
		<i>Gerres cinereus</i> (Walbaum, 1792)	LC	ZB	MM	X				X	X
		<i>Ulaema lefroyi</i> (Goode, 1874)	LC	ZB	ES		X	X	X		
	Mullidae	<i>Pseudupeneus maculatus</i> (Bloch, 1793)	LC	ZB	MS				X		
	Serranidae	<i>Alphesthes afer</i> (Bloch, 1793)	DD	ZB	MS						X
		<i>Epinephelus adscensionis</i> (Osbeck, 1765)	DD	ZB	MS	X		X	X		X
		<i>Epinephelus itajara</i> (Lichtenstein, 1822)	CR	PV/OP	MS						X
		<i>Mycteroperca bonaci</i> (Poey, 1860)	VU	PV	MS	X					
		<i>Rypticus randalli</i> Courtenay, 1967	LC	PV	MS	X				X	X
		<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)	LC	ZB	MS			X	X		X
		<i>Serranus flaviventris</i> (Cuvier, 1829)	LC	OP	MS			X			
	Chaetodontidae	<i>Chaetodon ocellatus</i> (Bloch, 1787)	DD	ZB	MS	X					
		<i>Chaetodon striatus</i> Linnaeus, 1758	LC	ZP	MS			X			
	Pomacanthidae	<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	DD	ZB	MS						X
		<i>Pomacanthus paru</i> (Bloch, 1787)	DD	ZB	MS				X		

Haemulidae *Conodon nobilis* (Linnaeus, 1758) LC ZB MM | X

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Order	Family	Species (Author, Year)	IUCN	FMFG	EUFG	STC	JAG	SUA	MAR	SIR	RIO	
Perciformes	Haemulidae	<i>Genyatremus luteus</i> (Bloch, 1790)	LC	OP	MS	X					X	
		<i>Haemulon aurolineatum</i> Cuvier, 1830	LC	ZB	MS			X	X			
		<i>Haemulon parra</i> (Desmarest, 1823)	LC	ZB	MS	X		X			X	
		<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)	LC	ZB	MS		X		X			
		<i>Haemulopsis corvinaeformis</i> (Steindachner, 1868)	LC	ZB	MS		X		X	X	X	
		<i>Pomadasys croco</i> (Cuvier, 1830)	LC	ZB	MS	X	X		X	X	X	
		<i>Pomadasys ramosus</i> Poey, 1860	LC	ZB	MM		X					
		Lutjanidae	<i>Lutjanus alexandrei</i> Moura & Lindeman, 2007	LC	ZB	MS	X	X	X	X	X	X
			<i>Lutjanus analis</i> (Cuvier, 1828)	NT	ZB	MS	X		X			X
			<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	VU	ZB	MS		X		X	X	X
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	NT		ZB	MS	X	X	X	X	X	X		
Scorpaeniformes	Polynemidae	<i>Lutjanus synagris</i> (Linnaeus, 1758)	NT	ZB	MS	X	X	X			X	
		<i>Ocyurus chrysurus</i> (Bloch, 1791)	NT	ZB	MS						X	
	Scorpaenidae	<i>Polydactylus virginicus</i> (Linnaeus, 1758)	LC	ZB	MM	X			X	X	X	
		<i>Scorpaena plumieri</i> Bloch, 1789	LC	ZB	MS			X			X	
	Moroniformes	Triglidae	<i>Prionotus punctatus</i> (Bloch, 1793)	LC	ZB	MS	X	X	X			X
			<i>Chaetodipterus faber</i> (Broussonet, 1782)	LC	OV	MM	X	X	X	X	X	X
	Acanthuriformes	Sciaenidae	<i>Bairdiella ronchus</i> (Cuvier, 1830)	LC	ZB	MM	X	X	X	X	X	X
			<i>Cynoscion acoupa</i> (Lacepède, 1801)	NT	PV	MM						X
			<i>Cynoscion leiarchus</i> (Cuvier, 1830)	LC	ZB	MM	X					X
			<i>Cynoscion virescens</i> (Cuvier, 1830)	LC	-	MM	X					
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)			LC	PV	MM	X				X	X	
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)			LC	PV	MM	X						
<i>Menticirrhus americanus</i> (Linnaeus, 1758)			DD	ZB	MM	X	X			X	X	
<i>Micropogonias furnieri</i> (Desmarest, 1823)			LC	ZB	MM	X				X	X	
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925			DD	ZB	MM					X		
<i>Stellifer brasiliensis</i> (Schultz, 1945)			LC	ZB	MM						X	
Spariformes	Sparidae	<i>Stellifer microps</i> (Steindachner, 1864)	LC	ZB	ES	X				X		
		<i>Stellifer rastrifer</i> (Jordan, 1889)	LC	ZB	MM					X		
		<i>Stellifer stellifer</i> (Bloch, 1790)	LC	ZB	ES	X						
		<i>Acanthurus bahianus</i> Castelnau, 1855	LC	HV	MS				X			
		<i>Acanthurus chirurgus</i> (Bloch, 1787)	LC	HV	MS	X		X				
		<i>Archosargus probatocephalus</i> (Walbaum, 1792)	DD	OV	MS	X					X	
		<i>Archosargus rhomboidalis</i> Linnaeus, 1758	LC	ZB	MS	X	X	X	X	X	X	
		<i>Lobotes surinamensis</i> (Bloch, 1790)	LC	PV	MS	X						
		Lophiiformes	Antennariidae	<i>Stellifer stellifer</i> (Bloch, 1790)	LC	ZB	MS				X	
				<i>Antennarius striatus</i> (Shaw, 1794)	DD	ZB	MS					X
	Ogcocephalidae	<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	LC	ZB	MS						X	

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Tetraodontiformes	Balistidae	<i>Balistes vetula</i> Linnaeus, 1758	NT	ZB	MS						X	
	Tetraodontidae	<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	LC	ZB	ES			X				
		<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	LC	HV	MM		X	X				X
		<i>Sphoeroides greeleyi</i> Gilbert, 1900	LC	ZB	ES		X	X		X		X
		<i>Sphoeroides spengleri</i> (Bloch, 1785)	LC	ZB	ES				X	X		
		<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	DD	ZB	ES		X	X		X	X	X
	Diodontidae	<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	LC	-	MS				X			
		<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)	LC	ZB	MS		X	X				

† Categories based on the regional IUCN red list of threatened species (ICMBio, 2018).

‡ References used for FMFG and EUFG classification are shown in the Supplementary Material Table I.

References

- Able KW, Fahay MP. Ecology of estuarine fishes: Temperate waters of the Western North Atlantic. Baltimore: Johns Hopkins University Press; 2010.
- Albuquerque PTF, Frédoú T, Arruda GN, Filho CAS, Nascimento AF, da Silva MJ, et al. Tracking Hg historical inputs by Pb-210 geochronology for the Itapessoca Estuarine Complex, Pernambuco, Brazil. J Radioanal Nucl Chem. 2019; 321:875–83. doi: <https://doi.org/10.1007/s10967-019-06665-9>.
- Andrade-Tubino MF, Ribeiro ALR, Vianna M. Organização Espaço-Temporal Das Ictiocenoses Demersais Nos Ecossistemas Estuarinos Brasileiros: Uma Síntese. Oecologia Australis. 2008; 12(04):640–61. doi: <https://doi.org/10.4257/oeco.2008.1204.05>.
- APAC. Monitoramento Pluviométrico. 2020. <http://www.apac.pe.gov.br/meteorologia/monitoramento-pluvio.php> (accessed May 14, 2020).
- Araújo ME, Teixeira JM c, Oliveira AME. Peixes Estuarinos Marinhos do Nordeste do Brasileiro. Fortaleza: Editora UFC; 2004.
- Aschenbrenner A, Ferreira BP. Age, growth and mortality of *Lutjanus alexandrei* in estuarine and coastal waters of the tropical south-western Atlantic. J Appl Ichthyol. 2015; 31(1):57–64. doi: <https://doi.org/10.1111/jai.12633>.
- Barletta M, Costa MF. Living and non-living resources exploitation in a tropical semi-arid estuary. J Coastal Res. 2009(January).
- Bastos RB, Feitosa FA do N, Koenig ML, Machado RC de A, Muniz K. Caracterização de uma zona costeira tropical (Ipojuca-Pernambuco-Brasil): Produtividade fitoplanctônica e outras variáveis ambientais. Braz J Aquat Sci Technol. 2011; 15(1):1. doi: <https://doi.org/10.14210/bjast.v15n1.p1-10>.
- Bernardino AF, Pagliosa PR, Christofolletti RA, Barros F, Netto SA, Muniz P, et al. Benthic estuarine communities in Brazil: moving forward to long term studies to assess climate change impacts. Braz J Ocean. 2016; 64:81–96. doi: <https://doi.org/http://dx.doi.org/10.1590/S1679-875920160849064sp2>.
- Bezerra DMM, Nascimento DM, Ferreira EN, Rocha PD, Mourão JS. Influence of tides and winds on fishing techniques and strategies in the mamanguape River Estuary, Paraíba State, NE Brazil. An Acad Bras Cienc. 2012; 84(3):775–87. doi: <https://doi.org/10.1590/S0001-37652012005000046>.
- Blaber SJM. Tropical estuarine fishes. ecology, exploitation and conservation. Blackwell Sci. 2000:372. doi: [https://doi.org/10.1643/0045-8511\(2001\)001\[0880:\]2.0.CO;2](https://doi.org/10.1643/0045-8511(2001)001[0880:]2.0.CO;2).
- Blaber SJM, Blaber TG. Factors affecting the distribution of juvenile estuarine and inshore fish. J Fish Biol. 1980; 17:143–62. doi: <https://doi.org/10.1111/j.1095-8649.1980.tb02749.x>.
- Boerema A, Meire P. Management for estuarine ecosystem services: A review. Ecol Eng. 2017; 98:172–82. doi: <https://doi.org/10.1016/j.ecoleng.2016.10.051>.
- Borges GCP. Comunidade fitoplanctônica do estuário do Rio Massangana (Pernambuco - Brasil).
- Bucci AF, Ciotti ÁM, Pollery RCG, de Carvalho R, de Albuquerque HC, Simões LTS. Temporal variability of chlorophyll-a in the São Vicente estuary. Braz J Oceanogr. 2012; 60(4):485–99. doi: <https://doi.org/10.1590/S1679-87592012000400007>.
- Caffrey JM, Chapin TP, Jannasch HW, Haskins JC. High nutrient pulses, tidal mixing and biological response in a small California estuary: Variability in nutrient concentrations from decadal to hourly time scales. Estuar Coast Shelf Sci. 2007; 71(3–4):368–80. doi: <https://doi.org/10.1016/j.ecss.2006.08.015>.
- Carpenter KE. The living marine resources of the Western Central Atlantic. Volume 1: Introduction, molluscs, crustaceans, hagfishes, sharks, batoid fishes, and chimaeras. vol. 1. Virginia: FAO; 2003.
- Castro ACL. Diversidade da assembléia de peixes em igarapés do estuário do Rio Paciência (MA - Brasil). Atlântica. 2001; 23:61–72.

- Clarke K, Warwick R. Change in marine communities: an approach to statistical analysis and interpretation. 2nd edition. Primer-E, Plymouth. 2001. doi: <https://doi.org/10.1017/S0025315418000176>.
- Contente RF. Padrões ecológicos locais e multidecadais da ictiofauna do estuário Cananéia-Iguape 2013:186.
- Cordeiro IA, Feitosa FADN, Flores Montes MDJ, Otsuka AY, Costa Silva A. Environmental conditions of the Suape estuarine-port complex area (Pernambuco, Brazil): Phytoplankton biomass and hydrological parameters. *J Mar Biol Assoc UK*. 2019; 99(2):291–301. doi: <https://doi.org/10.1017/S0025315418000176>.
- Costa CGF. Estudo da ecologia da paisagem no estuário do rio Jaguaribe no litoral do Ceará (Brasil) numa perspectiva geoambiental. *Rev Bras Ges Amb*. 2013; 7(2):24–32.
- Costanza R, de Groot R, Sutton P, van der Ploeg S, Anderson SJ, Kubiszewski I, et al. Changes in the global value of ecosystem services. *G Environ Chan*. 2014. doi: <https://doi.org/10.1016/j.gloenvcha.2014.04.002>.
- CPRH. Hidrografia. Diagnostico Socioambiental Do Litoral Sul de Pernambuco. Recife: Companhia Pernambucana do Meio Ambiente; 2003.
- CPRH. Diagnóstico socioambiental - Litoral Sul de Pernambuco. Recife: CPRH; 2001.
- Dame RF. Estuaries. *Encyclopedia of Ecology, Five-Volume Set*. Elsevier Inc.; 2008. p.1407–13. doi: <https://doi.org/10.1016/B978-008045405-4.00329-3>.
- Dominguez JML. The Coastal Zone of Brazil. In: Dillenburg SR, Hesp PA, editors. *Geology and Geomorphology of Holocene Coastal Barriers of Brazil*. Berlin: Springer-Verlag Berlin Heidelberg; 2009. p.17–51.
- Duarte GAS, Andreatta J v. Hábito alimentar das espécies de Achiridae e Cynoglossidae que ocorrem na Baía da Ribeira, Angra dos Reis, Rio de Janeiro, Brasil. *Bioikos*. 2003; 17:39–48.
- Eduardo LN, Bertrand A, Frédo T, Lira AS, Lima RS, Ferreira BP, et al. Biodiversity, ecology, fisheries, and use and trade of Tetraodontiformes fishes reveal their socio-ecological significance along the tropical Brazilian continental shelf. *Aquat Conser*. 2020; 1(January):1–14. doi: <https://doi.org/10.1002/aqc.3278>.
- Eduardo LN, Frédo T, Lira AS, Ferreira BP, Bertrand A, Ménard F, et al. Identifying key habitat and spatial patterns of fish biodiversity in the tropical Brazilian continental shelf. *Cont Shelf Res*. 2018. doi: <https://doi.org/10.1016/j.csr.2018.07.002>.
- El-deir ACA. Composição e Distribuição Espaço-Temporal de Formas Iniciais de Peixes no Estuário do Rio Jaguaribe, Itamaracá, Litoral Norte de Pernambuco, Brasil.
- Elliott M, Quintino V. The Estuarine Quality Paradox, Environmental Homeostasis and the difficulty of detecting anthropogenic stress in naturally stressed areas. *Mar Pollut Bull*. 2007; 54(6):640–5. doi: <https://doi.org/10.1016/j.marpolbul.2007.02.003>.
- Elliott M, Whitfield AK, Potter IC, Blaber SJM, Cyrus DP, Nordlie FG, et al. The guild approach to categorizing estuarine fish assemblages: A global review. *Fish Fish*. 2007; 8(3):241–68. doi: <https://doi.org/10.1111/j.1467-2679.2007.00253.x>.
- Falcão ECS. Evidências de conectividade entre habitats costeiros tropicais através do estudo de peixes em fases iniciais do ciclo de vida.
- Falcão ECS. Estrutura Da Comunidade De Formas Iniciais De Peixes Em Uma Gamboa Do Estuário Do Rio Catuama, Pernambuco - Brasil.
- Favero F de LT, Araujo IM da S, Severi W. Structure of the fish assemblage and functional guilds in the estuary of Maracaípe, Northeastern coast of Brazil. *Bol Inst Pesca*. 2019; 45(1):417. doi: <https://doi.org/10.20950/1678-2305.2019.45.1.417>.
- Fernandes MFB. Os manguezais da costa norte brasileira. Belém: Fundação Rio Bacanga; 2003.
- Ferreira V, le Loc'h F, Ménard F, Frédo T, Frédo FL. Composition of the fish fauna in a tropical estuary: the ecological guild approach. *Sci Mar*. 2019; 83(2):133. doi: <https://doi.org/10.3989/scimar.04855.25a>.

- Figueiredo JL, Menezes NA. Manual de Peixes Marinhos do Sudeste do Brasil. São Paulo: Museu de Zoologia- USP; 2000.
- Flores-Lopes F, Cetra M, Malabarba LR. Utilização de índices ecológicos em assembléias de peixes como instrumento de avaliação da degradação ambiental em programas de monitoramento. *Biota Neotrop.* 2010; 10(4):184–93.
- Flores Motes MDJ, Macêdo SJ de, Koenig ML, Lins Correia I. Variação nictmeral do fitoplâncton e elementos nutrientes no canal de Santa Cruz, Itamaracá – PE – Brasil. *Trab Oceanograf Univ Fed PE.* 1998; 26(1):16-26.
- Franco A, Elliott M, Franzoi P, Torricelli P. Life strategies of fishes in European estuaries: The functional guild approach. *Mar Ecol Prog Ser.* 2008; 354:219–28. doi: <https://doi.org/10.3354/meps07203>.
- Fricke, R., Eschmeyer, W. N. & Van der Laan R. Eschmeyer's Catalog of Fishes: Genera, Species, References 2020. <http://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp> (accessed October 21, 2020).
- Fujiwara M, Martinez-Andrade F, Wells RJD, Fisher M, Pawluk M, Livernois MC. Climate-related factors cause changes in the diversity of fish and invertebrates in subtropical coast of the Gulf of Mexico. *Communications Biology.* 2019; 2(1):1–9. doi: <https://doi.org/10.1038/s42003-019-0650-9>.
- Gaspar ALB. Idade, crescimento e padrões de recrutamento do bobó, *Sparisoma axillare*, na Apa Costa dos Corais. doi: <https://doi.org/10.1590/s0102-33061995000200015>.
- Gillanders BM, Elsdon TS, Halliday IA, Jenkins GP, Robins JB, Valesini FJ. Potential effects of climate change on Australian estuaries and fish utilising estuaries: A review. *Mar Freshw Res.* 2011; 62(9):1115–31. doi: <https://doi.org/10.1071/MF11047>.
- Gonzalez JG, Ménard F, le Loc'h F, Andrade HA de, Viana AP, Ferreira V, et al. Trophic resource partitioning of two snook fish species (Centropomidae) in tropical estuaries in Brazil as evidenced by stable isotope analysis. *Est Coast Shelf Sci.* 2019. doi: <https://doi.org/10.1016/j.ecss.2019.106287>.
- Gotelli NJ, Colwell RK. Biological diversity: frontiers in measurement and assessment. *Challenges.* 2011:39–54.
- Herman PMJ, Middelburg JJ, van de Koppel J, Heip CHR. Ecology of estuarine macrobenthos. *Adv Ecol Res.* 1999; 29:195–240.
- Honorato da Silva M, Passavante JZ de O, Silva-Cunha M da GG da, Nascimento Vieira DA do, Grego CK da S, Muniz K. Distribuição espacial e sazonal da biomassa fitoplanctônica e dos parâmetros hidrológicos no estuário do Rio Formoso (Rio Formoso, Pernambuco, Brasil). *Trop Oceanog.* 2004; 32(1):89–106.
- James NC, Harrison TD. A preliminary fish survey of the estuaries on the southeast coast of south africa, kayser's beach - Kei mouth: A comparative study. *Water SA.* 2016; 42(1):82–101. doi: <https://doi.org/10.4314/wsa.v42i1.10>.
- James NC, van Niekerk L, Whitfield AK, Potts WM, Götz A, Paterson AW. Effects of climate change on South African estuaries and associated fish species. *Clim Res.* 2013; 57(3):233–48. doi: <https://doi.org/10.3354/cr01178>.
- Kennish MJ. Estuaries, Anthropogenic Impacts. Schwartz ML (Eds) *Ency Coast Scie.* 2005. doi: https://doi.org/https://doi.org/10.1007/1-4020-3880-1_140.
- König C, Weigelt P, Schrader J, Taylor A, Kattge J, Kreft H. Biodiversity data integration—the significance of data resolution and domain. *PLoS Biol.* 2019; 17(3):1–16. doi: <https://doi.org/10.1371/journal.pbio.3000183>.
- Kuparinen A, O'Hara RB, Merilä J. Lunar periodicity and the timing of river entry in Atlantic salmon *Salmo salar*. *J Fish Biol.* 2009; 74(10):2401–8.
- Lana P da C, Bernardino AF. Brazilian Estuaries: A Benthic Perspective. Cham: Spri Internat Publis; 2018. doi: <https://doi.org/doi.org/10.1007/978-3-319-77779-5>.

Lessa R, Nóbrega MF de. Guia de Identificação de Peixes Marinhos da Região Nordeste. vol. 1. Recife: Revizee; 2000.

Lessa R, Viera AC de S, Monteiro A, Santos JS, Lima MM de, Cunha EJ da, et al. Diagnóstico da Pesca no Litoral do Estado de Pernambuco. A pesca Marinha e Estuarina do Brasil no Início do século XXI: recursos, tecnologias, aspectos socioeconômicos e institucionais. Belém: Editora Universitária UFPA; 2006. p.67–91.

Lima ARB, Torres RA, Jacobina UP, Pinheiro MAA, Adam ML. Genomic damage in *Mugil curema* (Actinopterygii: Mugilidae) reveals the effects of intense urbanization on estuaries in northeastern Brazil. *Mar Pollut Bull.* 2019; 138(July 2018):63–9. doi: <https://doi.org/10.1016/j.marpolbul.2018.07.037>.

Lima DA, Costa JTM. Flora e vegetação terrestres da área Programa Suape (Comunicação Técnica No.5) 1978:27.

Lira AS, Angelini R, Le Loc'h F, Ménard F, Lacerda C, Frédou T, et al. Trophic flow structure of a neotropical estuary in northeastern Brazil and the comparison of ecosystem model indicators of estuaries. *J Mar Syst.* 2018; 182(August 2017):31–45. doi: <https://doi.org/10.1016/j.jmarsys.2018.02.007>.

Marty E. Características ecológicas de espécies de peixes marinhos brasileiros com interesse para aquarofilia. 2014. p.21.

Medeiros APM, Xavier JH de A, da Silva MB, Aires-Souza L, Rosa IM de L. Distribution patterns of the fish assemblage in the Mamanguape River Estuary, North-eastern Brazil. *Mar Biol Res.* 2018; 14(5):524–36. doi: <https://doi.org/10.1080/17451000.2018.1459724>.

Medeiros C, Kjerfve B, Araujo M, Neumann-Leitão S. The Itamaracá Estuarine Ecosystem, Brazil. *Coast Mar Ecosys Lat Amer.* 2001;144:71–82. doi: <https://doi.org/10.1007/978-3-662-04482-7>.

Mello MVL de. Parâmetros hidrológicos correlacionados com a biomassa e composição fitoplanctônica na região costeira adjacente a desembocadura do rio Sirinhaém (Pernambuco - Brasil). 2009:125.

Menezes NA, Figueiredo JL. Manual de peixes marinhos do sudeste do Brasil. IV. Teleostei (3). São Paulo: Museu de Zoologia- USP; 1980.

Menezes NA, Nirchio M, de Oliveira C, Siccharamirez R. Taxonomic review of the species of *Mugil* (Teleostei: Perciformes: Mugilidae) from the Atlantic South Caribbean and South America, with integration of morphological, cytogenetic and molecular data. *Zootaxa.* 2015; 3918(1). doi: <https://doi.org/10.11646/zootaxa.3918.1.1>.

Menge BA, Olson AM. Role of scale and environmental factors in regulation of community structure. *Trends Ecol Evol.* 1990; 5(2):52–7. doi: [https://doi.org/10.1016/0169-5347\(90\)90048-I](https://doi.org/10.1016/0169-5347(90)90048-I).

Mérigot B, Frédou FL, Viana AP, Ferreira BP, do Nascimento Costa Junior E, Beserra da Silva Júnior CA, et al. Fish assemblages in tropical estuaries of northeast Brazil: A multi-component diversity approach. *Ocean Coas Manag.* 2017; 143(October 2017):175–83. doi: <https://doi.org/10.1016/j.ocecoaman.2016.08.004>.

Meynecke JO, Lee SY, Duke NC, Warnken J. Effect of rainfall as a component of climate change on estuarine fish production in Queensland, Australia. *Est Coast Shelf Sci.* 2006; 69(3–4):491–504. doi: <https://doi.org/10.1016/j.ecss.2006.05.011>.

Miloslavich P, Klein E, Díaz JM, Hernández CE, Bigatti G, Campos L, et al. Marine biodiversity in the Atlantic and Pacific coasts of South America: Knowledge and gaps. *PLoS ONE.* 2011; 6(1). doi: <https://doi.org/10.1371/journal.pone.0014631>.

Muniz K, Neto BDB, de Macêdo SJ, Pinheiro Filho WC. Hydrological impact of the port complex of Suape on the Ipojuca River (Pernambuco-Brazil). *J Coast Res.* 2005; 21(5):909–14. doi: <https://doi.org/10.2112/03-0070.1>.

Nakayama P, Peret AC, Cardoso OR, Lamour MR, Spach HL. Temporal patterns of fish occurrence of the euryhaline sector of a subtropical estuary, southern Brazil. *Acta Scie.* 2020; 42(August):1–10. doi: <https://doi.org/10.4025/actascibiolsci.v42i1.48871>.

- Nelson JS, Grande TC, Wilson MVH. Fishes of the world. New Jersey: John Wiley & Sons; 2016. doi: <https://doi.org/10.1017/CBO9781107415324.004>.
- Neta RNFC, Castro ACL de. Diversidade das assembleias de peixes estuarinos da ilha dos Caranguejos, Maranhão. *Arq Cien Mar*. 2008; 41(1):48–57.
- Oksanen J, Kindt R, Legendre P, O'Hara B, Simpson GL, Solymos P, et al. Vegan: community ecology package. R package version 1.15-2 2009.
- Oksanen JF, Blanchet G, Friendly M, Kindt R, Legendre P, McGlinn D, et al. Vegan: Community Ecology Package. R package version 2.5-6. 2019.
- Paiva A, Lima M, Souza J, Araújo M. Spatial distribution of the estuarine ichthyofauna of the Rio Formoso (Pernambuco, Brazil), with emphasis on reef fish. *Zoologia*. 2009; 26(2):266–78.
- Paiva ACG de, Araújo ME de. Environmental characterization and spatial distribution of fish fauna in estuaries in the state of Pernambuco, Brasil. *Trop Oceano Online*. 2010; 38(1):1–46.
- Paiva ACG de, Tarso P de, Chaves C, Araújo ME de. Estrutura e organização trófica da ictiofauna de águas rasas em um estuário tropical. *Rev Bras Zool*. 2008; 25(4):647–61.
- Parizzi RA, Machado E da C, Tavares, C. P. dos S., Fernandes LF, de Camargo, M. G., Mafra LL. Producción primaria y dinámica del fitoplancton en un estuario subtropical: Un enfoque múltiple de escalas de tiempo. *Sci Mar*. 2016; 80:291–303.
- Passarone R, Aparecido KC, Eduardo LN, Lira AS, Silva LVS, Justino AKS, et al. Ecological and conservation aspects of bycatch fishes: An evaluation of shrimp fisheries impacts in northeastern Brazil. *Braz J Oceanog*. 2019; 67(Figure 1):1–10. doi: <https://doi.org/10.1590/s1679-87592019029106713>.
- Passos AC dos, Contente RF, Abbatepaulo FV, Spach HL, Vilar CC, Joyeux JC, et al. Analysis of fish assemblages in sectors along a salinity gradient based on species, families and functional groups. *Braz J Oceanog*. 2013; 61(4):251–64. doi: <https://doi.org/10.1590/S1679-87592013000400006>.
- Pelage L, Domalain G, Lira AS, Travassos P, Frédo T. Coastal land use in Northeast Brazil: Mangrove coverage evolution over three decades. *Trop Conser Sci*. 2019; 12:194008291882241. doi: <https://doi.org/10.1177/1940082918822411>.
- Pereira L de JG. Análise Multidisciplinar de uma Pescaria Proibida: Estudo de Caso da Pesca do Mero *Epinephelus itajara* (Lichtenstein, 1822) na costa Amazônica Brasileira.
- Pikitch EK, Santora C, Babcock E a, Bakun A, Bonfil R, Conover DO, et al. Ecosystem-Based Fishery Management. *Science* (1979). 2004; 300(February):2003–2003. doi: <https://doi.org/10.1126/science.1106929>.
- Potter IC, Tweedley JR, Elliott M, Whitfield AK. The ways in which fish use estuaries: a refinement and expansion of the guild approach. *Fish Fish*. 2015; 16(2):230–9. doi: <https://doi.org/10.1111/faf.12050>.
- R Core T. R: A language and environment for statistical computing. 2019.
- Ray GC. Connectivities of estuarine fishes to the coastal realm. *Est Coast Shelf Sci*. 2005; 64(1 SPEC. ISS.):18–32. doi: <https://doi.org/10.1016/j.ecss.2005.02.003>.
- Santana FM, Morize E, Labonne M, Lessa R, Clavier J. Connectivity between the marine coast and estuary for white mullet (*Mugil curema*) in northeastern Brazil revealed by otolith Sr:Ca ratio. *Est Coast Shelf Sci*. 2018; 215:124–31. doi: <https://doi.org/10.1016/J.ECSS.2018.09.032>.
- Silva AL. Sedimentologia do Canal de Santa-Cruz-Ilha de Itamaracá-PE.
- Silva AMC. Composição da meiofauna na ilha de itamaracá e sua relação com a descrição morfoscópica e morfométrica dos grãos, Pernambuco. *Rev Nord Zool*. 2013; 7(2):34–47.
- Silva JB da, Galvêncio JD, Corrêa AC de B, Silva DG da, Machado CC. Classificação geomorfológica dos estuários do estado de Pernambuco (Brasil) com base em imagens do LANDSAT 5/TM. *Rev Bras Geog Fis*. 2011; 06:118–33.

Silva JDB, Barletta M, Lima ARA, Ferreira GVB. Use of resources and microplastic contamination throughout the life cycle of grunts (Haemulidae) in a tropical estuary. *Environ Pollut.* 2018; 242(Pt A):1010–21. doi: <https://doi.org/10.1016/j.envpol.2018.07.038>.

Silva JDB da. Ecologia trófica e distribuição espaço-temporal das espécies *Pomadasys ramosus* e *Haemulopsis corvinaeformis* (Steindachner, 1868) (Haemulidae) ao longo do gradiente de variação no estuário do Rio Goiana (PE/PB).

Silva TDAE, Neumann-leitão S, Schwamborn R, Maria L, Gusmão DO, Nascimento-vieira DA. Diel and seasonal changes in the macrozooplankton community of a tropical estuary in Northeastern Brazil tropical. *Reva Bras Zool.* 2003; 20(3):439–46.

Silva-Falcão EC, Severi W, Rocha AAF da. Dinâmica espacial e temporal de zoeas de Brachyura (Crustacea, Decapoda) no estuário do Rio Jaguaribe, Itamaracá, Pernambuco, Brasil. *Iheringia Ser Zool.* 2007; 97(4):434–40. doi: <https://doi.org/10.1590/s0073-47212007000400012>.

Silva-Júnior CAB, Mérigot B, Lucena-Frédou F, Ferreira BP, Coxey MS, Rezende SM, et al. Functional diversity of fish in tropical estuaries: A traits-based approach of communities in Pernambuco, Brazil. *Est Coast Shelf Sci.* 2017; 198:413–20. doi: <https://doi.org/10.1016/j.ecss.2016.08.030>.

Souza MR, Carnerio MH, Quirino-Duarte G, Servo GJ. Caracterização Da “Mistura” Na Pesca De Arrasto-De-Parelha Desembarcada em Santos Guarujá, São Paulo. *Bol Instit Pesca.* 2007; 33(1):43–51.

Team RC. R: A language and environment for statistical computing. 2020.

Teixeira LJ, Galvincto DJ. Uso de Geotecnologias para a Caracterização Física Espacial da Bacia Hidrográfica do Grupo de Pequenos Rios Litorâneos (GL2) PE. *Reva Bras Geo Fis.* 2010; 2:133–9.

Vasconcelos Filho A de L, Oliveira AME de. Composição e ecologia da ictiofauna do Canal de Santa Cruz (Itamaracá-PE, Brasil). *Trab Oceanograf Univ Fed PE.* 1999; 27(1):101–13.

Vasconcelos RP, Henriques S, França S, Pasquaud S, Cardoso I, Laborde M, et al. Global patterns and predictors of fish species richness in estuaries. *J Anim Ecol.* 2015; 84(5):1331–41. doi: <https://doi.org/10.1111/1365-2656.12372>.

Vendel AL, Chaves PDT. Use of an estuarine environment (Barra do Saí lagoon, Brazil) as nursery by fish. *Rev Bras Zool.* 2006; 23(4):1117–22. doi: <https://doi.org/10.1590/S0101-81752006000400020>.

Vilar CC, Joyeux J-C. Synopsis of the knowledge on the Brazilian estuarine fishes. *PeerJ Preprints.* 2018:37. doi: <https://doi.org/10.7287>.

Vorwerk PD, Whitfield AK, Cowley PD, Paterson AW. The influence of selected environmental variables on fish assemblage structure in a range of southeast African estuaries. *Environmental Biol Fishes.* 2003; 66(3):237–47. doi: <https://doi.org/10.1023/A:1023922521835>.

Wannamaker CM, Rice JA. Effects of hypoxia on movements and behavior of selected estuarine organisms from the southeastern United States. *J Exper Mar Biol Ecol.* 2000; 249(2):145–63. doi: [https://doi.org/10.1016/S0022-0981\(00\)00160-X](https://doi.org/10.1016/S0022-0981(00)00160-X).

Whitfield AK. Ichthyofaunal assemblages in estuaries: A South African case study. *Rev Fish Biol Fish.* 1999; 9(2):151–86. doi: <https://doi.org/10.1023/A:1008994405375>.

Whitfield AK. Abundance of larval and O+ juvenile marine fishes in the lower reaches of 3 southern African estuaries with differing freshwater inputs. *Mar Ecol Progr Ser.* 1994; 105(3):257–68. doi: <https://doi.org/10.3354/meps105257>.

Whitfield AK, Elliott M. Fishes as indicators of environmental and ecological changes within estuaries: A review of progress and some suggestions for the future. *J Fish Biol.* 2002; 61(SUPPL. A):229–50. doi: <https://doi.org/10.1006/jfbi.2002.2079>.

Wickham H. *Ggplot2: Elegant Graphics for Data Analysis* 2016.

ZEE. Zoneamento ecológico e econômico da zona costeira do estado do Ceará. Fortaleza: SEMACE; 2005.

Supplementary Material

Table I: Classification of feeding mode functional group (FMFG): ZB - zoobenthivore; PV - piscivore; ZP - zooplanktivore; HV - herbivore; OV - omnivore; DV - detritivore; OP - opportunistic and estuarine use functional group (EUFMG): MM - marine migrants; MS - marine straggler; ES - estuarine species; FS - freshwater species, for the species observed during the study and references used for each category.

Species	FMFG	Reference FMFG	EUFMG	Reference EUFG
<i>Ablennes hians</i> (Valenciennes, 1846)	PV	(Collette 1995)	MS	(Sasekumar et al. 1992); (Collette 2003)
<i>Acanthurus bahianus</i> Castelnau, 1855	HV	(Pimentel 2012)	MS	(Vasconcelos Filho, Oliveira 1999)
<i>Acanthurus chirurgus</i> (Bloch, 1787)	HV	(Longo et al. 2015)	MS	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus achirus</i> (Linnaeus, 1758)	ZB	(Mourão et al. 2014)	ES	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus declives</i> Chabanaud, 1940	ZB	(Duarte, Andreatta 2001)	ES	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus lineatus</i> (Linnaeus, 1758)	ZB	(Vasconcelos Filho et al. 2003)	ES	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009	-	-	-	-
<i>Ahlia egmontis</i> (Jordan, 1884)	ZB	(Pimentel et al. 2018)	MS	(Ross, Rohde 2003)
<i>Albula vulpes</i> (Linnaeus, 1758)	ZB	(Crabtree et al. 1998)	MM	(Paiva et al. 2009)
<i>Alphestes afer</i> (Bloch, 1793)	ZB	(Randall 1967)	MS	(Medeiros et al. 2009; Marques, Ferreira 2011)
<i>Anchoa filifera</i> (Fowler, 1915)	ZB	(Muto et al. 2014)	ES	(Passos et al. 2013)
<i>Anchoa januaria</i> (Steindachner, 1879)	ZP	(Araújo et al. 2016)	ES	(Passos et al. 2013)
<i>Anchoa lyolepis</i> (Evermann & Marsh, 1902)	ZP	(Froese, Pauly, 2018)	MS	(Lieske, Myers 1994)
<i>Anchoa marinii</i> Hildebrand, 1943	ZP	(Whitehead, 1985)	MS	(Riede, 2004)
<i>Anchoa spinifer</i> (Valenciennes, 1848)	PV	(Nizinski, Munroe, 2002)	MM	(Vasconcelos Filho, Oliveira 1999)
<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)	ZB	(Araújo et al. 2008)	MM	(Araújo et al. 2008)
<i>Anchovia clupeioides</i> (Swainson, 1839)	ZP	(Paiva et al. 2008)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Anchoviella lepidentostole</i> (Fowler, 1911)	ZB	(Planquette et al. 1996)	MM	(Paiva Filho et al. 1986)
<i>Antennarius striatus</i> (Shaw & Nodder, 1794)	ZB	(Nakane et al. 2011)	MS	(Paiva et al. 2009)
<i>Archosargus probatocephalus</i> (Walbaum, 1792)	OV	(Castillo-Rivera et al. 2007)	MS	(Castillo Rivera et al., 2007)
<i>Archosargus rhomboidalis</i> Linnaeus, 1758	ZB	(Yáñez-Arancibia 1986)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Ariosoma balearicum</i> (Delaroche, 1809)	-	-	MS	(Miller, 2002; Ross et al. 2007)
<i>Aspistor luniscutis</i> (Valenciennes, 1840)	OV	(Denadai et al. 2012)	MS	(Denadai et al., 2012)
<i>Aspistor quadriscutis</i> (Valenciennes, 1840)	ZB	(Denadai et al., 2012)	MS	(Denadai et al., 2012)
<i>Astroscopus ygraecum</i> Cuvier, 1829	PV	(Cervigón, 1993)	MS	(Grimes et al. 1982; Peterson et al. 2000; César Felix et al. 2006)
<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1824)	OV	(Paiva et al. 2008)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Atherinella cf. blackburni</i> (Schultz, 1949)	ZP	(Gonzalez, Vaske Júnior, 2017)	MM	(Mattox et al. 2008; Correa- Herrera et al. 2017)
<i>Bairdiella ronchus</i> (Cuvier, 1830)	ZB	(Ferreira et al. 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)

<i>Balistes vetula</i> Linnaeus, 1758	ZB	(Menezes, 1979)	MS	(Schärer et al. 2010; Rivera Hernández et al. 2019)
<i>Bathygobius soporator</i> (Valenciennes, 1837)	PV	(Corrêa et al. 2005)	ES	(Paiva et al. 2009)
<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)	ZB	(Collette, 2010)	MS	(Uyeno et al. 1983)
<i>Bothus ocellatus</i> (Agassiz, 1831)	ZB	(Maurakis et al. 2010)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Bothus robinsi</i> Topp & Hoff, 1972	ZB	(Figueiredo, Menezes, 2000)	MS	(Mendonça, Araújo, 2002; Marancik et al. 2005)
<i>Bryx dunckeri</i> (Metzelaar, 1919)	ZP	(Passos et al. 2013)	ES	(Passos et al. 2013)
<i>Carangoides bartholomaei</i> (Cuvier, 1833)	PV	(Paiva et al. 2008)	MS	(Santos, 2012)
<i>Caranx crysos</i> (Mitchill, 1815)	PV	(Sley et al. 2009)	MS	(Barletta, Blaber, 2007)
<i>Caranx hippos</i> (Linnaeus, 1766)	PV	(Temóteo et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Caranx latus</i> (Agassiz, 1831)	ZB	(Temóteo et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)	ZB	-	ES	(Dantas et al. 2012)
<i>Cathorops spixii</i> (Agassiz, 1829)	ZB	(Medeiros et al. 2010)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Centropomus ensiferus</i> Poey, 1860	PV	(Rueda and Defeo 2003)	-	-
<i>Centropomus mexicanus</i> Bocourt 1868	PV	(Ferreira et al. 2019)	-	-
<i>Centropomus parallelus</i> (Poey, 1860)	PV	(Lira et al. 2017)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Centropomus pectinatus</i> Poey, 1860	PV	(Lira, 2017)	MM	(Jackson, Ockelmann-Lobello, 2006)
<i>Centropomus undecimalis</i> (Bloch, 1792)	PV	(Lira et al. 2017)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Cetengraulis edentulus</i> (Cuvier, 1829)	ZP	(Paiva et al. 2008)	MM	(Vasconcelos Filho, Oliveira, 2000)
<i>Chaetodipterus faber</i> (Broussonet, 1782)	OV	(Vasconcelos Filho et al. 2009)	MM	(Riede, 2004)
<i>Chaetodon ocellatus</i> (Bloch, 1787)	ZB	(Carpenter, 2003b)	MS	(Rocha, 2010)
<i>Chaetodon striatus</i> (Linnaeus, 1758)	ZP	(Liedke et al. 2016)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	-	-	MS	(Paiva et al. 2009)
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)	ZB	(Almeida-Silva et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Chirocentron bleekermanus</i> (Poey 1867)	ZB	(Muto et al. 2014)	MS	(Passos et al. 2013)
<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	ZB	(Silva, Lopes, 2002)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900	ZB	(Araújo et al. 2016)	ES	(Paiva et al. 2009)
<i>Citharichthys macrops</i> Dresel, 1885	ZP	(Castillo-Rivera et al. 2000)	MS	(Passos et al. 2013)
<i>Citharichthys spilopterus</i> (Günther, 1880)	ZB	(Vasconcelos Filho et al. 2010)	ES	(Paiva et al. 2009)
<i>Colomesus asellus</i> (Müller & Troschel, 1848)	IN	(Bartolette et al. 2018)	FS	-
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	ZB	(Lobato et al. 2018)	ES	(Barletta-Bergan et al. 2002; Zacardi et al. 2016)
<i>Conodon nobilis</i> Linnaeus, 1758	ZB	(Lopes et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Cosmocampus elucens</i> (Poey, 1868)	ZB	(Carpenter 2003b)	MS	(Passos et al. 2013)
<i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882)	DV	(Vasconcelos Filho et al. 2009)	ES	(Vasconcelos Filho, Oliveira, 1999)

<i>Ctenogobius saepepallens</i> (Gilbert & Randall, 1968)	ZB	(Randall, 2004)	MS	(Baldwin, Smith, 2003; Ross, Rhode, 2004)
<i>Ctenogobius shufeldti</i> (Jordan & Eigenmann, 1887)	OV	(Contente et al. 2012)	ES	(Wyanski, Targett, 2000)
<i>Ctenogobius smaragdus</i> (Valenciennes, 1837)	DV	(Passos et al. 2013)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Ctenogobius stigmaticus</i> (Poey, 1860)	DV	(Passos et al. 2013)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Cynoscion acoupa</i> (Lacepède, 1800)	PV	(Ferreira et al. 2016)	MM	(Passos et al. 2013)
<i>Cynoscion leiarchus</i> (Cuvier, 1830)	ZB	(Araújo et al. 2016)	MM	(Chao, 1978)
<i>Cynoscion virescens</i> (Cuvier, 1830)	-	-	MM	(Paiva et al. 2009)
<i>Dactylopterus volitans</i> Linnaeus, 1758	ZB	(Guedes et al. 2015)	MS	(Paiva et al. 2009)
<i>Dactyloscopus crossotus</i> Starks, 1913	-	-	MS	(Herrema et al. 1985)
<i>Diapterus auratus</i> (Ranzani, 1840)	ZB	(Temóteo et al. 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Diapterus rhombeus</i> (Cuvier, 1829)	ZP	(Temóteo et al. 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Echeneis neucrates</i> Linnaeus, 1758	DV	(Williams et al. 2003)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Eleotris pisonis</i> (Gmelin, 1789)	PV	(Gonçalves et al. 2018)	ES	(Nordlie, 1981; Winemiller, Ponwith, 1998)
<i>Elops saurus</i> Linnaeus, 1758	PV	(Robins, Ray, 1986)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Engraulis anchoita</i> Hubbs & Marini 1935	ZP	(Vasconcelos et al. 1998)	MS	(Riede, 2004)
<i>Epinephelus adscensionis</i> (Osbeck, 1765)	ZB	(Medeiros et al. 2010)	MS	(Pereira et al. 2015)
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	PV/OP	(Freitas et al. 2015)	MS	(Paiva et al. 2009)
<i>Erotelis smaragdus</i> (Valenciennes, 1837)	-	-	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Etropus crossotus</i> Jordan & Gilbert, 1882	ZB	(Paiva et al. 2008)	MM	(Oliveira, Favaro, 2011)
<i>Etropus longimanus</i> Norman, 1933	ZB	(Figueiredo, Menezes, 2000a)	MS	(Saad, Fagundes Netto, 1992; Derisio et al. 2012)
<i>Eucinostomus argenteus</i> (Baird & Girard, 1854)	ZB	(Leão, 2016)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	ZB	(Zahorcsak et al. 2000a)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Eucinostomus havana</i> (Nichols, 1912)	ZB	(Froese, Pauly, 2018)	MM	(Passos et al. 2013)
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	ZB	(Ramos et al. 2011)	MM	(Chaves, Bouchereau 2000)
<i>Eugerres brasilianus</i> (Cuvier, 1830)	OV	(Vasconcelos Filho et al. 2009)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Evorthodus lyricus</i> (Girard, 1858)	DV	(STRI, 2017)	MS	(Vasconcelos Filho and Oliveira 1999)
<i>Fistularia petimba</i> Lacepède, 1803	PV	(Kalogirou et al. 2007); (Paiva et al. 2009)	ES	(Passos et al. 2013)
<i>Fistularia tabacaria</i> Linnaeus, 1758	PV	(Fritzsche, 1990)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Genyatremus luteus</i> Bloch, 1795	OP	(Almeida et al. 2005)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Gerres cinereus</i> (Walbaum, 1792)	ZB	(Randall, Vergara 1978)	MM	(Halpern, 2004)
<i>Gobioides broussonnetii</i> Lacepède, 1800	DV	(Mata-Cortes et al. 2004)	ES	(Mourão et al. 2014)
<i>Gobionellus oceanicus</i> (Pallas, 1770)	DV	(Vasconcelos Filho et al. 2009)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Gobionellus stomatus</i> Starks, 1913	DV	(Passos et al. 2013)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Guavina guavina</i> (Valenciennes in Cuvier & Valenciennes, 1837)	ZB	(Teixeira, 1994)	ES	(Vasconcelos Filho, Oliveira, 1999)

<i>Gymnothorax funebris</i> Ranzani, 1839	ZB	(Carvalho-Filho, 1999)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Gymnothorax ocellatus</i> Agassiz, 1831	ZB	(Santos, Castro, 2003)	MS	(Lieske, Myers, 1994)
<i>Haemulon aurolineatum</i> Cuvier, 1830	ZB	(Pereira et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Haemulon parra</i> Desmarest, 1823	ZB	(Paiva et al. 2008)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Haemulon steindachneri</i> Jordan & Gilbert, 1882	ZB	(Pereira et al. 2015)	MS	(Daros, 2014)
<i>Haemulopsis corvinaeformis</i> Steindachner, 1868	ZB	(Denadai et al. 2013)	MS	(Paiva et al. 2009)
<i>Harengula clupeola</i> (Cuvier, 1829)	ZP	(Paiva et al. 2008)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Harengula humeralis</i> (Cuvier, 1829)	ZP	(Ortiz et al. 1996)	MM	(Bouchereau et al. 2008)
<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)	HV	(Schwamborn 2004)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Hippocampus erectus</i> Perry, 1810	ZB	(Teixeira, Musick, 2001)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Hippocampus reidi</i> Ginsburg, 1933	ZP	(Castro et al. 2008)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Holocentrus adscensionis</i> (Osbeck, 1765)	ZB	(Fischer et al. 1981)	MS	(Silva Júnior et al. 2015)
<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)	ZB	(Stehmann et al. 1978)	MM	(Fuentes et al. 2019)
<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	ZB	(Anderson et al. 2015)	MS	(Uyeno et al. 1983)
<i>Hyporhamphus roberti</i> (Valenciennes, 1847)	HV/OP	(Medeiros et al. 2017)	MM	(Díaz-Ruiz, 2003)
<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)	OV	(Ferreira, 2018)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)	PV	(Lira et al. 2017)	MM	(Silva Júnior et al. 2015)
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	HV	(Denadai et al. 2012)	MM	(Andrade et al. 2015)
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)	HV	(Sales, 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Lobotes surinamensis</i> (Bloch, 1790)	PV	(Franks et al. 2003)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Lutjanus alexandrei</i> Moura & Lindeman, 2007	ZB	(Moraes, 2012)	MS	(Fernandes et al. 2012)
<i>Lutjanus analis</i> (Cuvier, 1828)	ZB	(Freitas et al. 2011)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	ZB	(Allen, 1985)	MS	(Bastos et al. 2013)
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	ZB	(Monteiro et al. 2009)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Lutjanus synagris</i> (Linnaeus, 1758)	ZB	(Freitas et al. 2011)	MS	(Vasconcelos-Filho, Oliveira, 2000)
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	PV	(Bortoluzzi et al. 2006)	ES	(Mai, Vieira, 2013)
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)	PV	(Figueiredo et al. 2014)	MM	(Militelli, Macchi, 2004)
<i>Megalops atlanticus</i> Valenciennes, 1847	PV	(Cataño, 1994)	MM	(Rickards, 1968)
<i>Menticirrhus americanus</i> (Linnaeus, 1758)	ZB	(Rondineli et al. 2007)	MM	(Turra et al. 2012)
<i>Microgobius meeki</i> Evermann & Marsh, 1900	ZB	(Froese, Pauly, 2018)	MS	(WoRMS, 2020)
<i>Microphis lineatus</i> (Kaup, 1856)	ZP	(Teixeira, Perrone, 1998)	ES	(Miranda-Marure et al. 2004)
<i>Micropogonias furnieri</i> (Desmarest, 1823)	ZB	(Mendoza-Carranza, Vieira 2008)	MM	(Paiva et al. 2009)
<i>Mugil brevirostris</i> (Ribeiro, 1915)	-	-	MM	(Dantas et al. 2015)
<i>Mugil curema</i> Valenciennes, 1836	DV	(Passos et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Mugil curvidens</i> Valenciennes, 1836	DV	(Passos et al. 2013)	-	-
<i>Mugil incilis</i> Hancock, 1830	DV	(Passos et al. 2013)	MM	(Dantas et al. 2015)

<i>Mugil liza</i> Valenciennes, 1836	DV	(Cervigón, 1993)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007	-	-	MM	(Dantas et al. 2015)
<i>Mycteroperca bonaci</i> (Poey, 1860)	PV	(Daros, 2014)	MS	(Daros, 2014)
<i>Myrichthys ocellatus</i> (Lesueur, 1825)	ZB	(Luiz et al. 2008)	MS	(Monteiro-Neto et al. 2013)
<i>Myrophis punctatus</i> Lütken, 1852	ZB	(Vaslet et al. 2011)	MM	(Barletta et al. 2000)
<i>Notarius grandicassis</i> (Valenciennes, 1840)	ZB	(Mendes, Barthem, 2010)	MM	(Marceniuk et al. 2017)
<i>Ocyurus chrysurus</i> (Bloch, 1791)	ZB	(Heck, Weinstein, 1989)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	ZB	(Gibran, Castro, 1999)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Oligoplites palometa</i> (Curvier, 1833)	PV	(Vasconcelos Filho et al. 2010)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Oligoplites saliens</i> (Bloch, 1793)	PV	(Winik et al. 2007)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	PV	(Medeiros et al. 2017)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Ophichthus cylindroideus</i> (Ranzani, 1839)	PV	(Giarrizzo, Krumme, 2007)	ES	(Giarrizzo, Krumme, 2007)
<i>Ophichthus ophis</i> (Linnaeus, 1758)	PV	(Randall, 1967)	MS	(Schneider, 1990)
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925	ZB	(Zahorcsak et al. 2000)	MM	(Spach et al. 2004)
<i>Opisthonema oglinum</i> (Lesueur, 1818)	ZP	(Vasconcelos-Filho, 1979)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Paralichthys brasiliensis</i> (Ranzani, 1842)	ZB	(Froese, Pauly, 2018)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Peprilus paru</i> Linnaeus, 1758	OV	(Passos et al. 2013)	MS	(Passos et al. 2013)
<i>Phaeoptyx pigmentaria</i> (Poey, 1860)	ZP	(Greenfield, Johnson, 1990)	MS	(Greenfield, Johnson, 1990)
<i>Platanichthys platana</i> (Regan, 1917)	ZP	(Aguiaro et al. 2003)	ES	(Aguiaro et al. 2003)
<i>Poecilia vivipara</i> (Bloch & Schneider, 1801)	ZP	(Passos et al. 2013)	FS	(Paiva et al. 2009)
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	ZB	(Lopes et al. 1998)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	ZB	(Hourigan et al. 1989)	MS	(Gasparini et al. 2005)
<i>Pomacanthus paru</i> (Bloch, 1787)	ZB	(Hourigan et al. 1989)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Pomadasys croco</i> (Cuvier, 1830)	ZB	(Froese, Pauly, 2018)	MS	(Riede, 2004)
<i>Pomadasys ramosus</i> Poey, 1860	ZB	(Silva et al. 2018)	MM	(Silva et al. 2018)
<i>Prionotus punctatus</i> (Bloch, 1793)	ZB	(Longo et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Pseudupeneus maculatus</i> (Bloch, 1793)	ZB	(Dantas et al. 2012)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)	ZP	(Clark, Pessanha, 2015)	ES	(Clark, Pessanha, 2015)
<i>Rypticus randalli</i> Courtenay, 1967	PV	(Medeiros et al. 2017)	MS	(Passos et al. 2013)
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)	ZB	(Feitosa et al. 2012)	MS	(Bejarano et al. 2014)
<i>Sciades herzbergii</i> (Bloch, 1794)	ZB	(Medeiros et al. 2017)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Sciades parkeri</i> (Trail, 1982)	ZB	(Vasconcelos Filho et al. 2003)	ES	(Betancur et al. 2008)
<i>Sciades proops</i> (Valenciennes, 1840)	ZB	(Guedes, Vasconcelos Filho, 1980)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Scomberomorus brasiliensis</i> Collete, Russo & Zavala-Camin, 1978	PV	(Collette, Nauen, 1983)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Scomberomorus regalis</i> (Bloch, 1793)	PV	(Collette, Nauen, 1983)	MS	(Riede, 2004)

<i>Scorpaena plumieri</i> Bloch, 1789	ZB	(Heck, Weinstein, 1989)	MS	(Edwards, 1990)
<i>Selene brownii</i> (Curvier, 1816)	ZB	(Bomfim, 2014)	MS	(WoRMS, 2020)
<i>Selene vomer</i> (Linnaeus, 1758)	PV	(Daros, 2014)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Serranus flaviventris</i> (Cuvier, 1829)	OP	(Maia-Nogueira et al. 2009)	MS	(Lieske, Myers, 1994)
<i>Sparisoma amplum</i> (Ranzani, 1842)	HV	(Ferreira, Gonçalves, 2006)	MS	(Moura et al. 2001)
<i>Sparisoma axillare</i> (Steindachner, 1878)	HV	(Ferreira, Gonçalves, 2006)	MS	(Moura et al. 2001)
<i>Sparisoma radians</i> (Valenciennes, 1840)	HV	(Paiva et al. 2008)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Sphoeroides greeleyi</i> Gilbert, 1900	ZB	(Figueiredo, Menezes, 2000)	ES	(Schultz et al. 2002)
<i>Sphoeroides spengleri</i> (Bloch, 1785)	ZB	(Targett, 1978)	ES	(Paiva et al. 2009)
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	ZB	(Vasconcelos Filho et al. 1998)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Sphyraena barracuda</i> (Edwards, 1771)	PV	(Akadje et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Sphyraena guachancho</i> Cuvier, 1829	PV	(Lopes et al. 2012)	MS	(Bonecker et al. 2014)
<i>Stellifer brasiliensis</i> (Schultz, 1945)	ZB	(Passos et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Stellifer microps</i> (Steindachner, 1864)	ZB	(Giarrizzo, Krumme, 2007)	ES	(Barletta, Blaber, 2007)
<i>Stellifer rastrifer</i> (Jordan, 1889)	ZB	(Passos et al. 2013)	MM	(Passos et al. 2013)
<i>Stellifer stellifer</i> (Bloch, 1790)	ZB	(Pombo et al. 2013)	ES	(Dantas et al. 2012)
<i>Strongylura marina</i> (Walbaum, 1792)	PV	(Passos et al. 2013)	MM	(Passos et al. 2013)
<i>Strongylura timucu</i> (Walbaum, 1792)	PV	(Passos et al. 2013)	MM	(Paiva et al. 2009)
<i>Syacium micrurum</i> Ranzani, 1842	ZB	(Soares, 1997)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)	ZB	(Almeida, 2018)	MM	(Jaureguizar et al. 2003)
<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)	ZB	(Vasconcelos Filho et al. 2003)	ES	(Paiva et al. 2009)
<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)	ZB	(Guedes et al. 2004)	ES	(Paiva et al. 2009)
<i>Syngnathus pelagicus</i> Linnaeus, 1758	ZB	(Carpenter, 2003)	MS	(Passos et al. 2013)
<i>Synodus foetens</i> (Linnaeus, 1766)	PV	(Cruz-Escalona et al. 2005)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Thalassophryne nattereri</i> Steindachner, 1876	ZB	(Collette, 1966)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Trachinotus carolinus</i> (Linnaeus, 1766)	ZB	(Stefanoni, 2008)	MM	(Denadai et al. 2013)
<i>Trachinotus falcatus</i> (Linnaeus, 1758)	ZB	(Hofling et al. 1998)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Trichiurus lepturus</i> Linnaeus, 1758	PV	(Vasconcelos Filho et al. 2010)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Trinectes microphthalmus</i> Chabanaud, 1928	ZB	(Contente et al. 2009)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	ZB	(Reis-Filho et al. 2011)	MM	(Passos et al. 2013)
<i>Tylosurus acus acus</i> (Lacepède, 1803)	PV	(Contente et al. 2009)	MS	(WoRMS, 2020)
<i>Tylosurus crocodilus</i> (Péron & Lesueur, 1821)	PV	(Blaber, 1990)	MM	(Favero et al. 2019)
<i>Ulaema lefroyi</i> (Goode, 1874)	ZB	(Silva et al. 2016)	ES	(Passos et al. 2013)
Species	FMFG	Reference FMFG	EUFG	Reference EUFG
<i>Ablennes hians</i> (Valenciennes, 1846)	PV	(Collette 1995)	MS	(Sasekumar et al. 1992); (Collette 2003)

<i>Acanthurus bahianus</i> Castelnau, 1855	HV	(Pimentel 2012)	MS	(Vasconcelos Filho, Oliveira 1999)
<i>Acanthurus chirurgus</i> (Bloch, 1787)	HV	(Longo et al. 2015)	MS	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus achirus</i> (Linnaeus, 1758)	ZB	(Mourão et al. 2014)	ES	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus declives</i> Chabanaud, 1940	ZB	(Duarte, Andreatta 2001)	ES	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus lineatus</i> (Linnaeus, 1758)	ZB	(Vasconcelos Filho et al. 2003)	ES	(Vasconcelos Filho, Oliveira 1999)
<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009	-	-	-	-
<i>Ahlia egmontis</i> (Jordan, 1884)	ZB	(Pimentel et al. 2018)	MS	(Ross, Rohde 2003)
<i>Albula vulpes</i> (Linnaeus, 1758)	ZB	(Crabtree et al. 1998)	MM	(Paiva et al. 2009)
<i>Alphesthes afer</i> (Bloch, 1793)	ZB	(Randall 1967)	MS	(Medeiros et al. 2009; Marques, Ferreira 2011)
<i>Anchoa filifera</i> (Fowler, 1915)	ZB	(Muto et al. 2014)	ES	(Passos et al. 2013)
<i>Anchoa januaria</i> (Steindachner, 1879)	ZP	(Araújo et al. 2016)	ES	(Passos et al. 2013)
<i>Anchoa lyolepis</i> (Evermann & Marsh, 1902)	ZP	(Froese, Pauly, 2018)	MS	(Lieske, Myers 1994)
<i>Anchoa marinii</i> Hildebrand, 1943	ZP	(Whitehead, 1985)	MS	(Riede, 2004)
<i>Anchoa spinifer</i> (Valenciennes, 1848)	PV	(Nizinski, Munroe, 2002)	MM	(Vasconcelos Filho, Oliveira 1999)
<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)	ZB	(Araújo et al. 2008)	MM	(Araújo et al. 2008)
<i>Anchovia clupeioides</i> (Swainson, 1839)	ZP	(Paiva et al. 2008)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Anchoviella lepidentostole</i> (Fowler, 1911)	ZB	(Planquette et al. 1996)	MM	(Paiva Filho et al. 1986)
<i>Antennarius striatus</i> (Shaw & Nodder, 1794)	ZB	(Nakane et al. 2011)	MS	(Paiva et al. 2009)
<i>Archosargus probatocephalus</i> (Walbaum, 1792)	OV	(Castillo-Rivera et al. 2007)	MS	(Castilho Rivera et al., 2007)
<i>Archosargus rhomboidalis</i> Linnaeus, 1758	ZB	(Yáñez-Arancibia 1986)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Ariosoma balearicum</i> (Delaroche, 1809)	-	-	MS	(Miller, 2002; Ross et al. 2007)
<i>Aspistor luniscutis</i> (Valenciennes, 1840)	OV	(Denadai et al. 2012)	MS	(Denadai et al., 2012)
<i>Aspistor quadriscutis</i> (Valenciennes, 1840)	ZB	(Denadai et al., 2012)	MS	(Denadai et al., 2012)
<i>Astroscopus ygraecum</i> Cuvier, 1829	PV	(Cervigón, 1993)	MS	(Grimes et al. 1982; Peterson et al. 2000; César Felix et al. 2006)
<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1824)	OV	(Paiva et al. 2008)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Atherinella cf. blackburni</i> (Schultz, 1949)	ZP	(Gonzalez, Vaske Júnior, 2017)	MM	(Mattox et al. 2008; Correa-Herrera et al. 2017)
<i>Bairdiella ronchus</i> (Cuvier, 1830)	ZB	(Ferreira et al. 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Balistes vetula</i> Linnaeus, 1758	ZB	(Menezes, 1979)	MS	(Schärer et al. 2010; Rivera Hernández et al. 2019)
<i>Bathygobius soporator</i> (Valenciennes, 1837)	PV	(Corrêa et al. 2005)	ES	(Paiva et al. 2009)
<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)	ZB	(Collette, 2010)	MS	(Uyeno et al. 1983)
<i>Bothus ocellatus</i> (Agassiz, 1831)	ZB	(Maurakis et al. 2010)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Bothus robinsi</i> Topp & Hoff, 1972	ZB	(Figueiredo, Menezes, 2000)	MS	(Mendonça, Araújo, 2002; Marancik et al. 2005)
<i>Bryx dunckeri</i> (Metzelaar, 1919)	ZP	(Passos et al. 2013)	ES	(Passos et al. 2013)
<i>Carangoides bartholomaei</i> (Cuvier, 1833)	PV	(Paiva et al. 2008)	MS	(Santos, 2012)
<i>Caranx crysos</i> (Mitchill, 1815)	PV	(Sley et al. 2009)	MS	(Barletta, Blaber, 2007)
<i>Caranx hippos</i> (Linnaeus, 1766)	PV	(Temóteo et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)

<i>Caranx latus</i> (Agassiz, 1831)	ZB	(Temóteo et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)	ZB	-	ES	(Dantas et al. 2012)
<i>Cathorops spixii</i> (Agassiz, 1829)	ZB	(Medeiros et al. 2010)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Centropomus ensiferus</i> Poey, 1860	PV	(Rueda and Defeo 2003)	-	-
<i>Centropomus mexicanus</i> Bocourt 1868	PV	(Ferreira et al. 2019)	-	-
<i>Centropomus parallelus</i> (Poey, 1860)	PV	(Lira et al. 2017)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Centropomus pectinatus</i> Poey, 1860	PV	(Lira, 2017)	MM	(Jackson, Ockelmann-Lobello, 2006)
<i>Centropomus undecimalis</i> (Bloch, 1792)	PV	(Lira et al. 2017)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Cetengraulis edentulus</i> (Cuvier, 1829)	ZP	(Paiva et al. 2008)	MM	(Vasconcelos Filho, Oliveira, 2000)
<i>Chaetodipterus faber</i> (Broussonet, 1782)	OV	(Vasconcelos Filho et al. 2009)	MM	(Riede, 2004)
<i>Chaetodon ocellatus</i> (Bloch, 1787)	ZB	(Carpenter, 2003b)	MS	(Rocha, 2010)
<i>Chaetodon striatus</i> (Linnaeus, 1758)	ZP	(Liedke et al. 2016)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	-	-	MS	(Paiva et al. 2009)
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)	ZB	(Almeida-Silva et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Chirocentron bleekermanus</i> (Poey 1867)	ZB	(Muto et al. 2014)	MS	(Passos et al. 2013)
<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	ZB	(Silva, Lopes, 2002)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900	ZB	(Araújo et al. 2016)	ES	(Paiva et al. 2009)
<i>Citharichthys macrops</i> Dresel, 1885	ZP	(Castillo-Rivera et al. 2000)	MS	(Passos et al. 2013)
<i>Citharichthys spilopterus</i> (Günther, 1880)	ZB	(Vasconcelos Filho et al. 2010)	ES	(Paiva et al. 2009)
<i>Colomesus asellus</i> (Müller & Troschel, 1848)	IN	(Bartolette et al. 2018)	FS	-
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	ZB	(Lobato et al. 2018)	ES	(Barletta-Bergan et al. 2002; Zacardi et al. 2016)
<i>Conodon nobilis</i> Linnaeus, 1758	ZB	(Lopes et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Cosmocampus elucens</i> (Poey, 1868)	ZB	(Carpenter 2003b)	MS	(Passos et al. 2013)
<i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882)	DV	(Vasconcelos Filho et al. 2009)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Ctenogobius saepepallens</i> (Gilbert & Randall, 1968)	ZB	(Randall, 2004)	MS	(Baldwin, Smith, 2003; Ross, Rhode, 2004)
<i>Ctenogobius shufeldti</i> (Jordan & Eigenmann, 1887)	OV	(Contente et al. 2012)	ES	(Wyanski, Targett, 2000)
<i>Ctenogobius smaragdus</i> (Valenciennes, 1837)	DV	(Passos et al. 2013)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Ctenogobius stigmaticus</i> (Poey, 1860)	DV	(Passos et al. 2013)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Cynoscion acoupa</i> (Lacepède, 1800)	PV	(Ferreira et al. 2016)	MM	(Passos et al. 2013)
<i>Cynoscion leiarchus</i> (Cuvier, 1830)	ZB	(Araújo et al. 2016)	MM	(Chao, 1978)
<i>Cynoscion virescens</i> (Cuvier, 1830)	-	-	MM	(Paiva et al. 2009)
<i>Dactylopterus volitans</i> Linnaeus, 1758	ZB	(Guedes et al. 2015)	MS	(Paiva et al. 2009)
<i>Dactyloscopus crossotus</i> Starks, 1913	-	-	MS	(Herrema et al. 1985)
<i>Diapterus auratus</i> (Ranzani, 1840)	ZB	(Temóteo et al. 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)

<i>Diapterus rhombeus</i> (Cuvier, 1829)	ZP	(Temóteo et al. 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Echeneis neucrates</i> Linnaeus, 1758	DV	(Williams et al. 2003)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Eleotris pisonis</i> (Gmelin, 1789)	PV	(Gonçalves et al. 2018)	ES	(Nordlie, 1981; Winemiller, Ponwith, 1998)
<i>Elops saurus</i> Linnaeus, 1758	PV	(Robins, Ray, 1986)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Engraulis anchoita</i> Hubbs & Marini 1935	ZP	(Vasconcelos et al. 1998)	MS	(Riede, 2004)
<i>Epinephelus adscensionis</i> (Osbeck, 1765)	ZB	(Medeiros et al. 2010)	MS	(Pereira et al. 2015)
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	PV/OP	(Freitas et al. 2015)	MS	(Paiva et al. 2009)
<i>Erotelis smaragdus</i> (Valenciennes, 1837)	-	-	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Etropus crossotus</i> Jordan & Gilbert, 1882	ZB	(Paiva et al. 2008)	MM	(Oliveira, Favaro, 2011)
<i>Etropus longimanus</i> Norman, 1933	ZB	(Figueiredo, Menezes, 2000a)	MS	(Saad, Fagundes Netto, 1992; Derisio et al. 2012)
<i>Eucinostomus argenteus</i> (Baird & Girard, 1854)	ZB	(Leão, 2016)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	ZB	(Zahorcsak et al. 2000a)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Eucinostomus havana</i> (Nichols, 1912)	ZB	(Froese, Pauly, 2018)	MM	(Passos et al. 2013)
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	ZB	(Ramos et al. 2011)	MM	(Chaves, Bouchereau 2000)
<i>Eugerres brasilianus</i> (Cuvier, 1830)	OV	(Vasconcelos Filho et al. 2009)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Evorthodus lyricus</i> (Girard, 1858)	DV	(STRI, 2017)	MS	(Vasconcelos Filho and Oliveira 1999)
<i>Fistularia petimba</i> Lacepède, 1803	PV	(Kalogirou et al. 2007); (Paiva et al. 2009)	ES	(Passos et al. 2013)
<i>Fistularia tabacaria</i> Linnaeus, 1758	PV	(Fritzsche, 1990)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Genyatremus luteus</i> Bloch, 1795	OP	(Almeida et al. 2005)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Gerres cinereus</i> (Walbaum, 1792)	ZB	(Randall, Vergara 1978)	MM	(Halpern, 2004)
<i>Gobioides broussonnetii</i> Lacepède, 1800	DV	(Mata-Cortes et al. 2004)	ES	(Mourão et al. 2014)
<i>Gobionellus oceanicus</i> (Pallas, 1770)	DV	(Vasconcelos Filho et al. 2009)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Gobionellus stomatus</i> Starks, 1913	DV	(Passos et al. 2013)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Guavina guavina</i> (Valenciennes in Cuvier & Valenciennes, 1837)	ZB	(Teixeira, 1994)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Gymnothorax funebris</i> Ranzani, 1839	ZB	(Carvalho-Filho, 1999)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Gymnothorax ocellatus</i> Agassiz, 1831	ZB	(Santos, Castro, 2003)	MS	(Lieske, Myers, 1994)
<i>Haemulon aurolineatum</i> Cuvier, 1830	ZB	(Pereira et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Haemulon parra</i> Desmarest, 1823	ZB	(Paiva et al. 2008)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Haemulon steindachneri</i> Jordan & Gilbert, 1882	ZB	(Pereira et al. 2015)	MS	(Daros, 2014)
<i>Haemulopsis corvinaeformis</i> Steindachner, 1868	ZB	(Denadai et al. 2013)	MS	(Paiva et al. 2009)
<i>Harengula clupeola</i> (Cuvier, 1829)	ZP	(Paiva et al. 2008)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Harengula humeralis</i> (Cuvier, 1829)	ZP	(Ortiz et al. 1996)	MM	(Bouchereau et al. 2008)

<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)	HV	(Schwamborn 2004)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Hippocampus erectus</i> Perry, 1810	ZB	(Teixeira, Musick, 2001)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Hippocampus reidi</i> Ginsburg, 1933	ZP	(Castro et al. 2008)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Holocentrus adscensionis</i> (Osbeck, 1765)	ZB	(Fischer et al. 1981)	MS	(Silva Júnior et al. 2015)
<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)	ZB	(Stehmann et al. 1978)	MM	(Fuentes et al. 2019)
<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	ZB	(Anderson et al. 2015)	MS	(Uyeno et al. 1983)
<i>Hyporhamphus roberti</i> (Valenciennes, 1847)	HV/OP	(Medeiros et al. 2017)	MM	(Díaz-Ruiz, 2003)
<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)	OV	(Ferreira, 2018)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)	PV	(Lira et al. 2017)	MM	(Silva Júnior et al. 2015)
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	HV	(Denadai et al. 2012)	MM	(Andrade et al. 2015)
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)	HV	(Sales, 2015)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Lobotes surinamensis</i> (Bloch, 1790)	PV	(Franks et al. 2003)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Lutjanus alexandrei</i> Moura & Lindeman, 2007	ZB	(Moraes, 2012)	MS	(Fernandes et al. 2012)
<i>Lutjanus analis</i> (Cuvier, 1828)	ZB	(Freitas et al. 2011)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	ZB	(Allen, 1985)	MS	(Bastos et al. 2013)
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	ZB	(Monteiro et al. 2009)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Lutjanus synagris</i> (Linnaeus, 1758)	ZB	(Freitas et al. 2011)	MS	(Vasconcelos-Filho, Oliveira, 2000)
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	PV	(Bortoluzzi et al. 2006)	ES	(Mai, Vieira, 2013)
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)	PV	(Figueiredo et al. 2014)	MM	(Militelli, Macchi, 2004)
<i>Megalops atlanticus</i> Valenciennes, 1847	PV	(Cataño, 1994)	MM	(Rickards, 1968)
<i>Menticirrhus americanus</i> (Linnaeus, 1758)	ZB	(Rondineli et al. 2007)	MM	(Turra et al. 2012)
<i>Microgobius meeki</i> Evermann & Marsh, 1900	ZB	(Froese, Pauly, 2018)	MS	(WoRMS, 2020)
<i>Microphis lineatus</i> (Kaup, 1856)	ZP	(Teixeira, Perrone, 1998)	ES	(Miranda-Marure et al. 2004)
<i>Micropogonias furnieri</i> (Desmarest, 1823)	ZB	(Mendoza-Carranza, Vieira 2008)	MM	(Paiva et al. 2009)
<i>Mugil brevirostris</i> (Ribeiro, 1915)	-	-	MM	(Dantas et al. 2015)
<i>Mugil curema</i> Valenciennes, 1836	DV	(Passos et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Mugil curvidens</i> Valenciennes, 1836	DV	(Passos et al. 2013)	-	-
<i>Mugil incilis</i> Hancock, 1830	DV	(Passos et al. 2013)	MM	(Dantas et al. 2015)
<i>Mugil liza</i> Valenciennes, 1836	DV	(Cervigón, 1993)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007	-	-	MM	(Dantas et al. 2015)
<i>Mycteroperca bonaci</i> (Poey, 1860)	PV	(Daros, 2014)	MS	(Daros, 2014)
<i>Myrichthys ocellatus</i> (Lesueur, 1825)	ZB	(Luiz et al. 2008)	MS	(Monteiro-Neto et al. 2013)
<i>Myrophis punctatus</i> Lütken, 1852	ZB	(Vaslet et al. 2011)	MM	(Barletta et al. 2000)
<i>Notarius grandicassis</i> (Valenciennes, 1840)	ZB	(Mendes, Barthem, 2010)	MM	(Marceniuk et al. 2017)
<i>Ocyurus chrysurus</i> (Bloch, 1791)	ZB	(Heck, Weinstein, 1989)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Ogocephalus vespertilio</i> (Linnaeus, 1758)	ZB	(Gibran, Castro, 1999)	MS	(Vasconcelos Filho, Oliveira, 1999)

<i>Oligoplites palometa</i> (Curvier, 1833)	PV	(Vasconcelos Filho et al. 2010)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Oligoplites saliens</i> (Bloch, 1793)	PV	(Winik et al. 2007)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	PV	(Medeiros et al. 2017)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Ophichthus cylindroideus</i> (Ranzani, 1839)	PV	(Giarrizzo, Krumme, 2007)	ES	(Giarrizzo, Krumme, 2007)
<i>Ophichthus ophis</i> (Linnaeus, 1758)	PV	(Randall, 1967)	MS	(Schneider, 1990)
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925	ZB	(Zahorcsak et al. 2000)	MM	(Spach et al. 2004)
<i>Opisthonema oglinum</i> (Lesueur, 1818)	ZP	(Vasconcelos-Filho, 1979)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Paralichthys brasiliensis</i> (Ranzani, 1842)	ZB	(Froese, Pauly, 2018)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Peprilus paru</i> Linnaeus, 1758	OV	(Passos et al. 2013)	MS	(Passos et al. 2013)
<i>Phaeoptyx pigmentaria</i> (Poey, 1860)	ZP	(Greenfield, Johnson, 1990)	MS	(Greenfield, Johnson, 1990)
<i>Platanichthys platana</i> (Regan, 1917)	ZP	(Aguiaro et al. 2003)	ES	(Aguiaro et al. 2003)
<i>Poecilia vivipara</i> (Bloch & Schneider, 1801)	ZP	(Passos et al. 2013)	FS	(Paiva et al. 2009)
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	ZB	(Lopes et al. 1998)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	ZB	(Hourigan et al. 1989)	MS	(Gasparini et al. 2005)
<i>Pomacanthus paru</i> (Bloch, 1787)	ZB	(Hourigan et al. 1989)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Pomadasy croco</i> (Cuvier, 1830)	ZB	(Froese, Pauly, 2018)	MS	(Riede, 2004)
<i>Pomadasy ramosus</i> Poey, 1860	ZB	(Silva et al. 2018)	MM	(Silva et al. 2018)
<i>Prionotus punctatus</i> (Bloch, 1793)	ZB	(Longo et al. 2015)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Pseudupeneus maculatus</i> (Bloch, 1793)	ZB	(Dantas et al. 2012)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)	ZP	(Clark, Pessanha, 2015)	ES	(Clark, Pessanha, 2015)
<i>Rypticus randalli</i> Courtenay, 1967	PV	(Medeiros et al. 2017)	MS	(Passos et al. 2013)
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)	ZB	(Feitosa et al. 2012)	MS	(Bejarano et al. 2014)
<i>Sciades herzbergii</i> (Bloch, 1794)	ZB	(Medeiros et al. 2017)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Sciades parkeri</i> (Trail, 1982)	ZB	(Vasconcelos Filho et al. 2003)	ES	(Betancur et al. 2008)
<i>Sciades proops</i> (Valenciennes, 1840)	ZB	(Guedes, Vasconcelos Filho, 1980)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Scomberomorus brasiliensis</i> Collete, Russo & Zavala-Camin, 1978	PV	(Collette, Nauen, 1983)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Scomberomorus regalis</i> (Bloch, 1793)	PV	(Collette, Nauen, 1983)	MS	(Riede, 2004)
<i>Scorpaena plumieri</i> Bloch, 1789	ZB	(Heck, Weinstein, 1989)	MS	(Edwards, 1990)
<i>Selene brownii</i> (Curvier, 1816)	ZB	(Bomfim, 2014)	MS	(WoRMS, 2020)
<i>Selene vomer</i> (Linnaeus, 1758)	PV	(Daros, 2014)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Serranus flaviventris</i> (Cuvier, 1829)	OP	(Maia-Nogueira et al. 2009)	MS	(Lieske, Myers, 1994)
<i>Sparisoma amplum</i> (Ranzani, 1842)	HV	(Ferreira, Gonçalves, 2006)	MS	(Moura et al. 2001)
<i>Sparisoma axillare</i> (Steindachner, 1878)	HV	(Ferreira, Gonçalves, 2006)	MS	(Moura et al. 2001)
<i>Sparisoma radians</i> (Valenciennes, 1840)	HV	(Paiva et al. 2008)	MS	(Vasconcelos Filho, Oliveira, 1999)

<i>Sphoeroides greeleyi</i> Gilbert, 1900	ZB	(Figueiredo, Menezes, 2000)	ES	(Schultz et al. 2002)
<i>Sphoeroides spengleri</i> (Bloch, 1785)	ZB	(Targett, 1978)	ES	(Paiva et al. 2009)
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	ZB	(Vasconcelos Filho et al. 1998)	ES	(Vasconcelos Filho, Oliveira, 1999)
<i>Sphyraena barracuda</i> (Edwards, 1771)	PV	(Akadje et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Sphyraena guachancho</i> Cuvier, 1829	PV	(Lopes et al. 2012)	MS	(Bonecker et al. 2014)
<i>Stellifer brasiliensis</i> (Schultz, 1945)	ZB	(Passos et al. 2013)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Stellifer microps</i> (Steindachner, 1864)	ZB	(Giarrizzo, Krumme, 2007)	ES	(Barletta, Blaber, 2007)
<i>Stellifer rastrifer</i> (Jordan, 1889)	ZB	(Passos et al. 2013)	MM	(Passos et al. 2013)
<i>Stellifer stellifer</i> (Bloch, 1790)	ZB	(Pombo et al. 2013)	ES	(Dantas et al. 2012)
<i>Strongylura marina</i> (Walbaum, 1792)	PV	(Passos et al. 2013)	MM	(Passos et al. 2013)
<i>Strongylura timucu</i> (Walbaum, 1792)	PV	(Passos et al. 2013)	MM	(Paiva et al. 2009)
<i>Syacium micrurum</i> Ranzani, 1842	ZB	(Soares, 1997)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)	ZB	(Almeida, 2018)	MM	(Jaureguizar et al. 2003)
<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)	ZB	(Vasconcelos Filho et al. 2003)	ES	(Paiva et al. 2009)
<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)	ZB	(Guedes et al. 2004)	ES	(Paiva et al. 2009)
<i>Syngnathus pelagicus</i> Linnaeus, 1758	ZB	(Carpenter, 2003)	MS	(Passos et al. 2013)
<i>Synodus foetens</i> (Linnaeus, 1766)	PV	(Cruz-Escalona et al. 2005)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Thalassophryne nattereri</i> Steindachner, 1876	ZB	(Collette, 1966)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Trachinotus carolinus</i> (Linnaeus, 1766)	ZB	(Stefanoni, 2008)	MM	(Denadai et al. 2013)
<i>Trachinotus falcatus</i> (Linnaeus, 1758)	ZB	(Hofling et al. 1998)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Trichiurus lepturus</i> Linnaeus, 1758	PV	(Vasconcelos Filho et al. 2010)	MS	(Vasconcelos Filho, Oliveira, 1999)
<i>Trinectes microphthalmus</i> Chabanaud, 1928	ZB	(Contente et al. 2009)	MM	(Vasconcelos Filho, Oliveira, 1999)
<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	ZB	(Reis-Filho et al. 2011)	MM	(Passos et al. 2013)
<i>Tylosurus acus acus</i> (Lacepède, 1803)	PV	(Contente et al. 2009)	MS	(WoRMS, 2020)
<i>Tylosurus crocodilus</i> (Péron & Lesueur, 1821)	PV	(Blaber, 1990)	MM	(Favero et al. 2019)
<i>Ulaema lefroyi</i> (Goode, 1874)	ZB	(Silva et al. 2016)	ES	(Passos et al. 2013)

Reference Supplementary Material

Aguiaro T, Branco CWC, Verani JR, Caramaschi EP. Diet of the clupeid fish *Platanichthys platana* (Regan, 1917) in two different Brazilian coastal lagoons. Braz Arch Biol Tech. 2003; 46(2):215–22. doi: <https://doi.org/10.1590/S1516-89132003000200013>.

Akadje C, Diaby M, Leloc’H F, Konan JK, N’Da K. Diet of the barracuda *Sphyraena guachancho* in Côte d’Ivoire (Equatorial Eastern Atlantic Ocean). Cybium. 2013; 37(4):285–93.

Allen GR. FAO Species catalogue Volume 6. Snappers of the world. An annotated and illustrated catalogue of lutjanid species known to date. Rome: FAO Spec. Cat. Fish. Purp; 1985.

Almeida R. Distribuição espacial das guildas tróficas de peixes estuarinos no litoral Amazônico Brasileiro. 2018.

Almeida ZS, Nunes JLS, Alves M. Dieta alimentar de *Genyatremus luteus* (BLOCH, 1790)-(TELEOSTEI, PERCIFORMES: HAEMULIDAE) na Baía de São José, Maranhão, Brasil. *Atlântica*, Rio Grande. 2005; 27(1):39–47.

Almeida-Silva PH, Tubino RA, Zambrano LC, Hunder DA, Garritano SR, Monteiro-Neto C. Trophic ecology and food consumption of fishes in a hypersaline tropical lagoon. *J Fish Biol.* 2015; 86(6):1781–95. doi: <https://doi.org/10.1111/jfb.12689>.

Anderson W, Carpenter KE, Gilmore G, Milagrosa Bustamante G, Robertson R. *Diplectrum formosum*. The IUCN Red List of Threatened Species 2015: e.T16439926A16510272 2015. <http://dx.doi.org/10.2305/IUCN.UK.2015-2.RLTS.T16439926A16510272.en> (accessed October 1, 2018).

de Andrade AC, Santos SR, Verani JR, Vianna M. Guild composition and habitat use by Tetraodontiformes (Teleostei, Acanthopterygii) in a south-western Atlantic tropical estuary. *J Mar Biol Assoc UK.* 2015:1–14. doi: <https://doi.org/10.1017/S0025315415001368>.

Araújo FG, de Azevedo MCC, Guedes APP. Inter-decadal changes in fish communities of a tropical bay in southeastern Brazil. *Reg Stud Mar Sci.* 2016; 3:107–18. doi: <https://doi.org/10.1016/j.rsma.2015.06.001>.

Araújo ME de, Feitosa CV, Mattos SMG de. *Ecologia de peixes recifais em Pernambuco*. Recife: UFPE; 2018.

Baldwin CC, Smith DG. Larval gobiidae (Teleostei: Perciformes) of Carrie Bow Cay, Belize, Central America. *Bull Mar Sci.* 2003; 72(3):639–74.

Barletta M, Blaber SJM. Comparison of fish assemblages and guilds in tropical habitats of the Embley (Indo-West Pacific) and Caeté (Western Atlantic) Estuaries. *Bull Mar Sci.* 2007; 80(3):647–80.

Barletta M, Saint-Paul U, Barletta-Bergan A, Ekau W, Shories D. Spatial and temporal distribution of *Myrophis punctatus* (Ophichthidae) and associated fish fauna in a northern Brazilian intertidal mangrove forest. In: Liebezeit G, Dittman S, Kroncke I, editors. *Life at Interfaces and Under Extreme Conditions*. Wilhelmshaven: Springer Science+Business Media; 2000. p.2011.

Barletta-Bergan A, Barletta M, Saint-Paul U. Structure and seasonal dynamics of larval fish in the Caeté River Estuary in north Brazil. *Estuar Coast Shelf S.* 2002; 54(2):193–206. doi: <https://doi.org/10.1006/ecss.2001.0842>.

Bartolette R, Rosa DCO, Beserra DA, Soares BE, Albrecht MP, Brito MFG. Seasonal and ontogenetic diet patterns of the freshwater pufferfish *Colomesus asellus* (Müller & Troschel, 1849) in the upper-middle Tocantins river. *Acta Scient - Biol Sci.* 2018; 40(1). doi: <https://doi.org/10.4025/actascibiols.v40i1.35282>.

- Bejarano I, Appeldoorn RS, Nemeth M. Fishes associated with mesophotic coral ecosystems in La Parguera, Puerto Rico. *Coral Reefs*. 2014; 33(2):313–28. doi: <https://doi.org/10.1007/s00338-014-1125-6>.
- Betancur-R. R, Marceniuk AP, Béarez P. Taxonomic status and redescription of the Gillbacker sea catfish (Siluriformes: Ariidae: *Sciades parkeri*). *Copeia*. 2008(4):827–34. doi: <https://doi.org/10.1643/CI-07-218>.
- Blaber SJM, Milton DA, Rawlinson NJF, Tiroba G, Nichols P v. Diets of Lagoon Fishes of the Solomon Islands: Predators of Tuna Baitfish and Trophic Effects of Baitfishing on the Subsistence Fishery. *Fish Res*. 1990; 8:263–86.
- Bomfim A da C. Bioecologia da ictiofauna marinha descartada pelo arrasto camaroeiro em praias da Bacia Potiguar, Brasil. 2014.
- Bortoluzzi T, Aschenbrenner A da C, da Silveira C da R, Roos DC, Lepkoski ED, Martins JA, et al. Hábito alimentar da sardinha prata, *Lycengraulis grossidens* (Spix & Agassiz, 1829), (Pisces, Engraulidae), Rio Uruguai médio, sudoeste do Rio Grande do Sul, Brasil. *Biodivers Pampeana*. 2006; 4:11–23.
- Bouchereau J-L, Chaves P de T, Monti D. Factors Structuring the Ichthyofauna Assemblage in a Mangrove Lagoon (Guadeloupe, French West Indies). *J Coast Res*. 2008; 244(244):969–82. doi: <https://doi.org/10.2112/06-0804.1>.
- Carpenter KE. The Living Marine Resources of the Western Central Atlantic Volume 3 Bony fishes part 2 (Opistognathidae to Molidae), sea turtles and marine mammals. vol. 1. Virginia: FAOSpecies Identification Guide for Fishery Purposes and American Society of Ichthyologists and Herpetologists Special Publication No. 5.; 2003.
- Carvalho-Filho A. Peixes: costa Brasileira. São Paulo: Melro; 1999.
- Castillo-Rivera M, Kobelkowsky A, Chávez AM. Feeding biology of the flatfish *Citharichthys spilopterus* (Bothidae) in a tropical estuary of Mexico. *J Appl Ichthyol*. 2000; 16:73–8.
- Castillo-Rivera M, Zárate-Hernández R, Salgado-Ugarte IH. Juvenile and adult food habits of *Archosargus probatocephalus* (Teleostei: Sparidae) in a tropical estuary of Veracruz. *Hidrobiol*. 2007; 17(2):119–26.
- Castro A, Diniz A, Martins I, Vendel A, de Oliveira T, Rosa I. Assessing diet composition of seahorses in the wild using a non destructive method: *Hippocampus reidi* (Teleostei: Syngnathidae) as a study-case. *Neotrop Ichthyol*. 2008; 6(4):637–44.
- Cataño S, Garzón-Ferreira J. Ecología trófica del sábalo *Megalops atlanticus* (Pisces: MegaIopidae) en el área de Ciénaga Grande de Santa Marta, Caribe colombiano. *Rev Bio Trop*. 1994; 42(3):673684.
- Cervigón F. Los peces marinos de Venezuela. Volume 2. Caracas: Fundación Científica Los Roques; 1993.
- Cézar Felix F, Louis Spach H, Werner Hackradt C, Simon Moro P, C. Rocha D. Abundância sazonal e a composição da assembléia de peixes em duas praias estuarinas da Baía de Paranaguá, Paraná. *Rev Bras Zool*. 2006; 8(1):35–47.

Chao L. Sciaenidae. In: Fischer W, editor. FAO Species Identification Guide for Fishery Purposes; American Society of Ichthyologists and Herpetologists Special Publication, vol. 5. Rome: FAO; 1978.

Chaves P, Bouchereau JL. Use of mangrove habitat for reproductive activity by the fish assemblage in the Guaratuba Bay, Brazil. *Ocean Acta*. 2000; 23(3):273–80. doi: [https://doi.org/10.1016/S0399-1784\(00\)00130-4](https://doi.org/10.1016/S0399-1784(00)00130-4).

Clark FJK, Pessanha ALM. Diet and ontogenetic shift in habitat use by *Rhinosardinia bahiensis* in a tropical semi-arid estuary, north-eastern Brazil. *J Mar Biol Assoc UK*. 2015; 95(1):175–83. doi: <https://doi.org/10.1017/S0025315414000939>.

Collette B B. *Batrachoides surinamensis*. IUCN. 2010. <https://dx.doi.org/10.2305/IUCN.UK.2010-4.RLTS.T154930A4670747> (accessed August 19, 2018).

Collette B B. Belonidae. In: Carpenter K E, Niem V, editors. FAO para Identificación de Especies para lo Fines de la Pesca. Rome: FAO; 1995. p.919–26.

Collette BB. Family Belonidae Bonaparte 1832 - needlefishes. California Academy of Sciences Annotated Checklists of Fishes. 2003(16):1–22.

Collette BB. A review of the venomous toadfishes, subfamily Thalassophryinae. *Copeia*. 1966; 4:846–64.

Collette BB, Nauen CE. FAO Species Catalogue: Vol. 2 Scombrids of the World. vol. 2. 1983.

Contente RF, Stefanoni MF, Spach HL. Feeding ecology of the American freshwater goby *Ctenogobius shufeldti* (Gobiidae, Perciformes) in a sub-tropical estuary. *J Fish Biol*. 2012; 80(6):2357–73. doi: <https://doi.org/10.1111/j.1095-8649.2012.03300.x>.

Contente RF, Stefanoni MF, Spach HL. Size-related changes in diet of the slipper sole *Trinectes paulistanus* (Actinopterygii, Achiridae) juveniles in a subtropical Brazilian estuary. *Pan-Amer J Aqu Scie*. 2009; 4(1):63–9.

Correa-Herrera T, Barletta M, Lima ARA, Jiménez-Segura LF, Arango-Sánchez LB. Spatial distribution and seasonality of ichthyoplankton and anthropogenic debris in a river delta in the Caribbean Sea. *J Fish Biol*. 2017; 90(4):1356–87. doi: <https://doi.org/10.1111/jfb.13243>.

Crabtree RE, Stevens C, Snodgrass D, Stengard FJ. Feeding habits of bonefish, *Albula vulpes*, from the waters of the Florida Keys. *Fish Bull*. 1998; 96(4):754–766.

Cristina A, Bonecker T. Ordem Perciformes. 2014.

Cruz-Escalona VH, Peterson MS, Campos-Dávila L, Zetina-Rejón M. Feeding habits and trophic morphology of inshore lizardfish (*Synodus foetens*) on the central continental shelf off Veracruz, Gulf of Mexico. *J Appl Ichthyol*. 2005; 21(6):525–30. doi: <https://doi.org/10.1111/j.1439-0426.2005.00651.x>.

Dantas D v., Barletta M, Costa MF. Feeding ecology and seasonal diet overlap between *Stellifer brasiliensis* and *Stellifer stellifer* in a tropical estuarine ecocline. J Fish Biol. 2015; 86(2):707–33. doi: <https://doi.org/10.1111/jfb.12592>.

Dantas NCF de M, Feitosa CV, Araújo ME de. Composition and assemblage structure of demersal fish from São Cristóvão beach, Areia Branca, RN. Biota Neotrop. 2012; 12(3):108–17. doi: <https://doi.org/10.1590/s1676-06032012000300012>.

Daros FAL de M. Estudo da ictiofauna costeira no litoral do Paraná e adjacências através de censos visuais e microquímica de otólitos. 2014.

Denadai MR, Santos FB, Bessa E, Bernardes LP, Turra A. Population biology and diet of the puffer fish *Lagocephalus laevigatus* (Tetraodontiformes : Tetraodontidae) in Caraguatatuba Bay , south-eastern Brazil. J Mar Biol Assoc UK. 2012; 92(2):407–12. doi: <https://doi.org/10.1017/S0025315411001299>.

Denadai MR, Santos FB, Bessa E, Fernandez WS, Lorca L, Turra A. Population biology and diet of *Pomadasys corvinaeformis* (Perciformes: Pomadasyidae) in Caraguatabuba Bay, Southeastern Brazil. Rev Biol Trop. 2013; 61(4):1947–54. doi: <https://doi.org/10.4172/2324-8661.1000108>.

Derisio C, Betti P, de Astarloa JMD, Machinandiarena L. Desarrollo larval de *Etropus longimanus* (Paralichthyidae) y *Symphurus trewavasae* (Cynoglossidae) en la costa Bonaerense, Argentina. Scien Mar. 2012; 76(1):29–37. doi: <https://doi.org/10.3989/scimar.2012.76n1029>.

Díaz-Ruiz S. Characterization of fish assemblages in a tropical coastal lagoon in the northwest Gulf of Mexico. Cien Mar. 2003; 29(4B):631–44. doi: <https://doi.org/10.7773/cm.v29i42.189>.

Duarte GAS, Andreatta J v. Hábito alimentar das espécies de Achiridae e Cynoglossidae que ocorrem na Baía da Ribeira, Angra dos Reis, Rio de Janeiro, Brasil. Bioikos. 2003; 17:39–48.

Edwards A. Fish and Fisheries of Saint Helena Island. Newcastle: Centre for Tropical Coastal Management Studies; 1990.

Favero F de LT, Araujo IM da S, Severi W. Structure of the fish assemblage and functional guilds in the estuary of Maracaípe, Northeastern coast of Brazil. Bol Instit Pes. 2019; 45(1):417. doi: <https://doi.org/10.20950/1678-2305.2019.45.1.417>.

Feitosa CV, de Carvalho Teixeira Chaves LS, Ferreira BP, de Araujo ME. Recreational fish feeding inside Brazilian MPAs: Impacts on reef fish community structure. J Mar Biol Assoc UK. 2012; 92(7):1525–33. doi: <https://doi.org/10.1017/S0025315412000136>.

Fernandes CAF, de Oliveira PGV, Travassos PEP, Hazin FHV. Reproduction of the Brazilian snapper, *Lutjanus alexandrei* Moura & Lindeman, 2007 (Perciformes: Lutjanidae), off the northern coast of Pernambuco, Brazil. Neotrop Ichthyol. 2012; 10(3):587–92. doi: <https://doi.org/10.1590/S1679-62252012005000022>.

- Ferreira B. Fish assemblages in tropical estuaries of northeast Brazil: A multi-component diversity approach 2016(August). doi: <https://doi.org/10.1016/j.ocecoaman.2016.08.004>.
- Ferreira CEL, Gonçalves JEA. Community structure and diet of roving herbivorous reef fishes in the Abrolhos Archipelago, south-western Atlantic. *J Fish Biol.* 2006; 69(5):1533–51. doi: <https://doi.org/10.1111/j.1095-8649.2006.01220.x>.
- Ferreira V. Estrutura Trófica da Ictiofauna Estuarina e Marinha do Complexo Itapissuma/Itamaracá, Norte de Pernambuco, Brasil. 2018.
- Ferreira V, le Loc'h F, Ménard F, Frédou T, Frédou FL. Composition of the fish fauna in a tropical estuary: the ecological guild approach. *Scie Mar.* 2019; 83(2):133. doi: <https://doi.org/10.3989/scimar.04855.25a>.
- Figueiredo JL, Menezes NA. Manual de Peixes Marinhos do Sudeste do Brasil. São Paulo: Museu de Zoologia- USP; 2000.
- Figueiredo MB, Neta RNFC, Luiz Silva Nunes J, da Silva De Almeida Z. Feeding habits of *Macrodon ancylodon* (Actinopterygii, Sciaenidae) in northeast, Brazil. *Rev Biol Mar Oceanogr.* 2014; 49(3):559–66. doi: <https://doi.org/10.4067/s0718-19572014000300012>.
- Fischer W, Bianchi G, Scott WB. Holocentridae. FAO Species Identification sheets for fishery purposes Eastern Central Atlantic, fishing areas 34, 47 (in part). Ottawa: FAO Spec. Cat. Fish. Purp; 1981. p.pag.var.
- Franks JS, VanderKooy KE, Garber NM. Diet of Tripletail, *Lobotes surinamensis*, from Mississippi Coastal Waters. *Gulf Caribb Res.* 2003; 15(1):27–32. doi: <https://doi.org/10.18785/gcr.1501.05>.
- Freitas MO, Abilhoa V, da Costa Silva GH. Feeding ecology of *Lutjanus analis* (Teleostei: Lutjanidae) from Abrolhos Bank, Eastern Brazil. *Neotrop Ichthyol.* 2011; 9(2):411–8. doi: <https://doi.org/10.1590/S1679-62252011005000022>.
- Freitas MO, Abilhoa V, Giglio VJ, Hostim-Silva M, de Moura RL, Francini-Filho RB, et al. Diet and reproduction of the goliath grouper, *Epinephelus itajara* (Actinopterygii: Perciformes: Serranidae), in Eastern Brazil. *Act Ichthyol Pisc.* 2015; 45(1):1–11. doi: <https://doi.org/10.3750/AIP2015.45.1.01>.
- Fritzsch RA. Check-list of the fishes of the eastern tropical Atlantic (CLOFETA). Lisboa: JNICT; 1990.
- Froese R, Pauly D. Fishbase. FishBase. 2018.
- Fuentes MMPB, Gillis AJ, Ceriani SA, Guttridge TL, van Zinnicq Bergmann MPM, Smukall M, et al. Informing marine protected areas in Bimini, Bahamas by considering hotspots for green turtles (*Chelonia mydas*). *Biod Conserv.* 2019; 28(1):197–211. doi: <https://doi.org/10.1007/s10531-018-1647-2>.
- Gasparini JL, Floeter SR, Sazima I. Marine Ornamental Trade in Brazil. *Biod Conserv.* 2005; 14:2883–99. doi: <https://doi.org/10.1007/s10531-004-0222-1>.

Giarrizzo T, Krumme U. Spatial differences and seasonal cyclicality in the intertidal fish fauna from four mangrove creeks in a salinity zone of the Curuçá estuary, North Brazil. *Bol Mar Sci-Miami* . 2007; 80(3):739–54.

Gibrán FZ, Castro RMC. Activity, feeding behaviour and diet of *Ogcocephalus vespertilio* in southern west Atlantic. *J Fish Biol.* 1999; 55:588–95. doi: <https://doi.org/10.1006/jfbi.1999.1019>.

Gonzalez JG, Vaske Júnior T. Feeding ecology of the beach silverside *Atherinella blackburni* (Atherinopsidae) in a tropical sandy beach, Southeastern Brazil. *Braz J Oceanogr.* 2017; 65(3):346–55. doi: <https://doi.org/10.1590/s1679-87592017131506503>.

Greenfield DW, Johnson RK. Heterogeneity in habitat choice in cardinalfish community structure. *Copeia.* 1990:1107–14.

Grimes CB, Manooch CS, Huntsman GR. Reef and Rock Outcropping Fishes of the Outer Continental Shelf of North Carolina and South Carolina, and Ecological Notes on the Red Porgy and Vermilion Snapper. *Bull Mar Sci.* 1982; 32(1):277–89. doi: <https://doi.org/10.1017/CBO9781107415324.004>.

Guedes A, Araújo F, Azevedo M. Estratégia trófica dos linguados *Citharichthys spilopterus* Gunter e *Symphurus tessellatus* (Quoy & Gaimard) (Actinopterygii, Pleuronectiformes) na Baía de Sepetiba, Rio de Janeiro, Brasil. *Rev Bras Zool.* 2004; 21(4):857–64.

Guedes APP, Araújo FG, Pessanha ALM, Milagre RR. Partitioning of the feeding niche along spatial, seasonal and size dimensions by the fish community in a tropical Bay in Southeastern Brazil. *Mar Ecol.* 2015; 36(1):38–56. doi: <https://doi.org/10.1111/maec.12115>.

Guedes D de S, Vasconcelos Filho A de L. Estudo ecológico da região de Itamaracá, Pernambuco, Brasil. IX informações sobre a alimentação dos bagres branco e amarelo, (Pisces Ariidae). *Trab Oceanograf Univ Fed PE.* 1980; 15:323–30.

Halpern BS. Are mangroves a limiting resource for two coral reef fishes? *Mar Ecol Progr Ser.* 2004; 272:93–8. doi: <https://doi.org/10.3354/meps272093>.

Heck KL, Weinstein MP. Feeding Habits of Juvenile Reef Fishes Associated with Panamanian Seagrass Meadows. *Bull Mar Sci.* 1989; 45(3):629–36.

Herrema DJ, Peery BD, Williams-Walls N, Wilcox JR. Spawning Periods of Common Inshore Fishes on the Florida East Coast. *Northeast Gulf Sci.* 1985; 7(2):2–5. doi: <https://doi.org/10.18785/negs.0702.04>.

Hofling JC, Ferreira LI, Neto FBR, Filho AMP, Lima PAB, Gibin TE. Fish alimentation of the Gerreidae family of the estuarine Lagoon complex in Cananéia, São Paulo, Brazil desde a desembocadura do Rio Ribeira até o Canal. *Rev Bioikos.* 1998; 12(1):7–18.

Hourigan TF, Stanton FG, Motta PJ, Kelley CD, Carlson B. The feeding ecology of three species of Caribbean angelfishes (family Pomacanthidae). *Environ Biol Fish.* 1989; 24(2):105–16. doi: <https://doi.org/10.1007/BF00001281>.

Jackson TL, Ockelmann-Lobello L. Centropomidae: Snooks. In: Richards WJ, editor. In early stages of Atlantic Fishes - An identification guide for Western Central North Atlantic. 2006. p.1197–205.

Jaureguizar AJ, Menni R, Bremec C, Mianzan H, Lasta C. Fish assemblage and environmental patterns in the Río de la Plata estuary. *Estuar Coast Shelf S.* 2003; 56(5–6):921–33. doi: [https://doi.org/10.1016/S0272-7714\(02\)00288-3](https://doi.org/10.1016/S0272-7714(02)00288-3).

Kalogirou S, Corsini M, Kondilatos G, Wennhage H. Diet of the invasive piscivorous fish *Fistularia commersonii* in a recently colonized area of the eastern Mediterranean. *Biol Inv.* 2007; 9(8):887–96. doi: <https://doi.org/10.1007/s10530-006-9088-3>.

Leão G do N. Aspectos da biologia de *Eucinostomus argenteus* Baird e Girard, 1855, Gerreidae, capturado no Canal de Santa Cruz- Pernambuco. 2016.

Liedke AMR, Barneche DR, Ferreira CEL, Segal B, Nunes LT, Burigo AP, et al. Abundance, diet, foraging and nutritional condition of the banded butterflyfish (*Chaetodon striatus*) along the western Atlantic. *Mar Biol.* 2016; 163(1):1–13. doi: <https://doi.org/10.1007/s00227-015-2788-4>.

Lieske E, Myers R. Collins Pocket Guide. Coral reef fishes. Indo-Pacif & Caribbean including the Red Sea. New York: HarperCollins Publishers; 1994.

Lima LG. Ecologia Trófica de *Symphurus tessellatus* (Quoy & Gaimard, 1824) e *Citharichthys macrops* Dresel, 1885 (Actinopterygii, Pleuronectiformes) no sistema estuarino do rio Mamanguape, Paraíba – Brasil. 2012.

Lira AS, Frédou FL, Viana AP, Eduardo LN, Frédou T. Feeding ecology of *Centropomus undecimalis* (Bloch, 1792) and *Centropomus parallelus* (Poey, 1860) in two tropical estuaries in Northeastern Brazil. *Pan-Amer J Aqu Sci.* 2017; 12(2):123–35.

Lobato CMC, Soares BE, Montag LFA. Temporal and spatial variation in the trophic ecology of the banded puffer fish *Colomesus psittacus* (Tetraodontiformes: Tetraodontidae) in the Amazon coastal zone. *Mar Freshw Res.* 2018; 69(11):1724–32. doi: <https://doi.org/10.1071/MF17328>.

Longo GO, Morais RA, Martins CDL, Mendes TC, Aued AW, Cândido D v, et al. Between-habitat variation of benthic cover, reef fish assemblage and feeding pressure on the benthos at the only atoll in South Atlantic: Rocas atoll, NE Brazil. *PLoS ONE.* 2015; 10:1–29. doi: <https://doi.org/10.1371/journal.pone.0127176>.

Lopes PRD, Oliveira-Silva JT de, Ferreira-Melo ASA. Contribuição ao conhecimento da ictiofauna do manguezal de Cacha Pregos, Ilha de Itaparica, Baía de Todos os Santos, Bahia. *Rev Bras Zool.* 1998; 15(2):315–25. doi: <https://doi.org/10.1590/s0101-81751998000200005>.

Lopes PRD, Oliveira-Silva JT, Fernandes IP. Notas sobre a alimentação de *Conodon nobilis* (Linnaeus, 1758) (Actinopterygii: Haemulidae) na praia do malhado, Ilhéus (BahiaA). *Revista Ceciliana.* 2013; 5(2):1–38.

- Luiz OJ, Carvalho-Filho A, Ferreira CEL, Floeter SR, Gasparini JL, Sazima I. The reef fish assemblage of the Laje de Santos Marine State Park, Southwestern Atlantic: Annotated checklist with comments on abundance, distribution, trophic structure, symbiotic associations, and conservation. *Zootaxa*. 2008; 25(1807):1–25. doi: <https://doi.org/10.11646/zootaxa.1807.1.1>.
- Mai ACG, Vieira JP. Revisão e considerações sobre o uso do habitat, distribuição e história de vida de *Lycengraulis grossidens* (Agassiz, 1829) (Actinopterygii, Clupeiformes, Engraulidae). *Bio Neotrop*. 2013; 13(3):121–30. doi: <https://doi.org/10.1590/S1676-06032013000300015>.
- Maia-Nogueira R, de Anchieta Nunes JC, C Coni EO, Ferreira CM, Sampaio CL. The twinspace bass *Serranus flaviventris* (Serranidae) as follower of the goldspotted eel *Myrichthys ocellatus* (Ophichthidae) in north-eastern Brazil, with notes on others serranids. *JMBA - Biod Rec*. 2008:1–3.
- Marancik KE, Clough LM, Hare JA. Cross-shelf and seasonal variation in larval fish assemblages on the southeast United States continental shelf off the coast of Georgia. *Fish Bull*. 2005; 103(1):108–29.
- Marceniuk AP, Caires R, Rotundo MM, Alcântara RAK, Wosiacki WB. The ichthyofauna (Teleostei) of the Rio Caeté estuary, northeast Pará, Brazil, with a species identification key from northern Brazilian coast. *Pan-Amer J Aqu Sci*. 2017; 12(January):31–79.
- Marques S, Ferreira BP. Sexual development and reproductive pattern of the Mutton hamlet, *Alphestes afer* (Teleostei: Epinephelidae): A dyandric, hermaphroditic reef fish. *Neotrop Ichthyol*. 2011; 9(3):547–58. doi: <https://doi.org/10.1590/S1679-62252011005000026>.
- Mata-Cortes S, Martinez-Perez JA, Peterson MS. Feeding Habits and Sexual Dimorphism of the Violet Goby, *Gobioides broussoneti* Lacepede (Pisces: Gobiidae), in the Estuarine System of Tecolutla, Veracruz, Mexico. *Gulf Caribb Res*. 2004; 16(1):89–93. doi: <https://doi.org/10.18785/gcr.1601.15>.
- Mattox G, Gondolo G, Cunningham. *Atherinella blackburni* (Schultz, 1949) at Itamambuca Beach, Ubatuba, SP: ecological characterization and distribution on the Brazilian coast (Teleostei: Atheriniformes: Atherinopsidae). *Braz J Biol*. 2008; 68(2):307–13.
- Maurakis EG, Maurakis GE, Maurakis DE. Functional Feeding Groups , Species Richness and Spatial Distributions of Fishes in Rocky and Sandy Richmond , VA 23220. *Virginia J Sci*. 2010; 61(4):127–50.
- Medeiros APM de, Xavier JH de A, Rosa IM de L. Diet and trophic organization of the fish assemblage from the Mamanguape River Estuary, Brazil. *Latin Amer J Aqu Res*. 2017; 45(5):879–90. doi: <https://doi.org/10.3856/vol45-issue5-fulltext-2>.
- Medeiros DV, Nunes J de ACC, Sampaio CLS. A mutton hamlet *Alphestes afer* (Bloch, 1793) reproductive event in northeast Brazil. *Pan-Amer J Aqu Sci*. 2009; 4(2):212–5.
- Medeiros PR, Gempel RG, Souza AT, Ilarri MI, Rosa RS. Non-random reef use by fishes at two dominant zones in a tropical, algal-dominated coastal reef. *Environ Biol Fish*. 2010; 87(3):237–46. doi: <https://doi.org/10.1007/s10641-010-9593-1>.

- Mendes F, Barthem R. Hábitos alimentares de bagres marinhos (Siluriformes: Ariidae) do estuário amazônico. *Amazônia: Ci & Desenv.* 2010; 5(10):153–66.
- Mendonça P, Araújo FG. Composição das populações de linguados (Osteichthyes, Pleuronectiformes) da Baía de Sepetiba, Rio de Janeiro, Brasil. *Rev Bras Zool.* 2002; 19(2):339–47.
- Mendoza-Carranza M, Vieira J. Whitemouth croaker *Micropogonias furnieri* (Desmarest, 1823) feeding strategies across four southern Brazilian estuaries. *Aqu Ecol.* 2008; 42(1):83–93. doi: <https://doi.org/10.1007/s10452-007-9084-4>.
- Menezes MF de. Aspectos da biologia e biometria do cangulo, *Balistes vetula* Linnaeus, no Nordeste do Brasil. *Arq Ciên Mar.* 1979; 19(1/2):57–68.
- Militelli MI, Macchi GJ. Spawning and fecundity of king weakfish, *Macrodon ancylodon*, in the Río de la Plata estuary, Argentina-Uruguay. *J Mar Biol Assoc UK.* 2004; 84(2):443–7. doi: <https://doi.org/10.1017/S0025315404009427h>.
- Miller MJ. The distribution and ecology of *Ariosoma balearicum* (Congridae) leptocephali in the western North Atlantic. *Environ Biol Fish.* 2002; 63(3):235–52. doi: <https://doi.org/10.1023/A:1014311429809>.
- Miranda-Marure ME, Martínez-Pérezand JA, Brown-Peterson NJ. Reproductive biology of the opossum pipefish, *Microphis brachyurus lineatus*, in Tecolutla Estuary, Veracruz, Mexico. *Gulf Caribb Res.* 2004; 16:101–8.
- Monteiro DP, Giarrizzo T, Isaac V. Feeding ecology of juvenile dog snapper *Lutjanus jocu* (Bloch and Shneider, 1801) (Lutjanidae) in intertidal mangrove creeks in Curuçá estuary (Northern Brazil). *Braz Arch Biol Tech.* 2009; 52(6):1421–30. doi: <https://doi.org/10.1590/S1516-89132009000600014>.
- Monteiro-Neto C, Bertoncini ÁA, Chaves LDCT, Noguchi R, Mendonça-Neto JP, Rangel CA. Checklist of marine fish from coastal islands of Rio de Janeiro, with remarks on marine conservation. *Mar Biod Rec.* 2013; 6(Figure 1):1–13. doi: <https://doi.org/10.1017/S1755267213000973>.
- Moraes RLG de. Pesca, parasitismo e dieta alimentar da baúna *Lutjanus alexandrei* Moura & Lindeman, 2007 nos ambientes costeiros do litoral sul de Pernambuco. 2012.
- Moura RL, Figueiredo JL, Sazima I. A new parrotfish (Scaridae) from Brazil, and revalidation of *Sparisoma amplum* (Ranzani, 1842), *Sparisoma frondosum* (Agassiz, 1831), *Sparisoma axillare* (Steindachner, 1878) and *Scarus trispinosus* Valenciennes, 1840. *Bull Mar Sci.* 2001; 68(3):505–24.
- Mourão KRM, Ferreira V, Lucena-Frédou F. Composition of functional ecological guilds of the fish fauna of the internal sector of the amazon estuary, Pará, Brazil. *An Acad Bras Cien.* 2014; 86(4):1783–800. doi: <https://doi.org/10.1590/0001-3765201420130503>.
- Muto EY, Corbisier TN, Coelho LI, Arantes LPL, Chalom A, Soares LSH. Trophic groups of demersal fish of Santos Bay and adjacent continental shelf, São Paulo State, Brazil: Temporal and spatial comparisons. *Braz J Oceanogr.* 2014; 62(2):89–102. doi: <https://doi.org/10.1590/S1679-87592014045906202>.

Nakane Y, Suda Y, Sano M. Food habits of fishes on an exposed sandy beach at Fukiagehama, South-West Kyushu Island, Japan. *Helgo Mar Res.* 2011; 65(2):123–31. doi: <https://doi.org/10.1007/s10152-010-0208-1>.

Nizinski MS, Munroe T. Engraulidae. *Living Marine Resources of the Western Central Atlantic.* Rome: FAO; 2002. p.764–94.

Nordlie FG. Feeding and reproductive biology of eleotrid fishes in a tropical estuary. *J Fish Biol.* 1981; 18:97–110.

Oliveira EC de, Favaro LF. Reproductive biology of the flatfish *Etropus crossotus* (Pleuronectiformes: Paralichthyidae) in the Paranaguá Estuarine Complex, Paraná State, subtropical region of Brazil. *Neotrop Ichthyol.* 2011; 9(4):795–805.

Ortiz M, Rocha ME, Posada JM. Food habits of the sympatric fishes *Harengula humeralis* and *H. clupeola* (Clupeidae) in the Archipiélago de Los Roques National Park, Venezuela. *J Sci.* 1996; 32(1):26–32.

Paiva A, Lima M, Souza J, Araújo M. Spatial distribution of the estuarine ichthyofauna of the Rio Formoso (Pernambuco, Brazil), with emphasis on reef fish. *Zoologia.* 2009; 26(2):266–78.

Paiva ACG de, Tarso P de, Chaves C, Araújo ME de. Estrutura e organização trófica da ictiofauna de águas rasas em um estuário tropical. *Rev Bras Zool.* 2008; 25(4):647–61.

Paiva Filho AM, Zani-Teixeira M de L, Kihara PK. Contribuição ao conhecimento da biologia da manjuba, *Anchoviella lepidentostole* (Fowler, 1911), no Estuário de São Vicente, SP (Osteichthyes, Engraulidae). *Bol Instit Oceanogr.* 1986; 34(0):71–7. doi: <https://doi.org/10.1590/s0373-55241986000100006>.

Passos AC dos, Contente RF, Abbatepaulo FV, Spach HL, Vilar CC, Joyeux JC, et al. Analysis of fish assemblages in sectors along a salinity gradient based on species, families and functional groups. *Braz J Oceanogr.* 2013; 61(4):251–64. doi: <https://doi.org/10.1590/S1679-87592013000400006>.

Pereira PHC, Barros B, Zemoi R, Ferreira BP. Ontogenetic diet changes and food partitioning of *Haemulon* spp. coral reef fishes, with a review of the genus diet. *Rev Fish Biol Fish.* 2015; 25(1):245–60. doi: <https://doi.org/10.1007/s11160-014-9378-2>.

Peterson MS, Comyns BH, Hendon JR, Bond PJ, Duff GA. Habitat use by early life-history stages of fishes and crustaceans along a changing estuarine landscape: Differences between natural and altered shoreline sites. *Wetl Ecol Manag.* 2000; 8(2–3):209–19. doi: <https://doi.org/10.1023/A:1008452805584>.

Pimentel CR. Organização trófica da comunidade de peixes de poças de maré da Praia dos Castelhanos (ES). 2012.

Pimentel CR, Soares LSH, Macieira RM, Joyeux JC. Trophic relationships in tidepool fish assemblages of the tropical Southwestern Atlantic. *Mar Ecol.* 2018; 39(2):1–11. doi: <https://doi.org/10.1111/maec.12496>.

Planquette P, Keith P, Bail P-Y le. Atlas des Poissons d'Eau Douce de Guyane. vol. 22. Paris: Institut d'Ecologie et de Gestion de la Biodiversité; 1996. doi: <https://doi.org/10.2307/1447869>.

Ramos JAA, Barletta M, Dantas D v., Lima ARA, Costa MF. Influence of moon phase on fish assemblages in estuarine mangrove tidal creeks. *J Fish Biol.* 2011; 78(1):344–54. doi: <https://doi.org/10.1111/j.1095-8649.2010.02851.x>.

Randall JE. Food Habits of Reef Fishes of the West Indies. *Stud Trop Oceanogr.* 2004; 5:665–847.

Randall JE, Vergara RR. Gerreidae. FAO Species Identification sheets for fishery purposes. 1978.

Reis-Filho JA, Nunes J de AC da C, Ferreira A. Estuarine ichthyofauna of the Paraguaçu River, Todos os Santos Bay, Bahia, Brazil. *Biota Neotrop.* 2011; 10(4):301–11. doi: <https://doi.org/10.1590/s1676-06032010000400034>.

Rickards WL. Ecology and Growth of Juvenile Tarpon *Megalops Atlanticus* in a Georgia Salt Marsh. *Bull Mar Sci.* 1968; 18(1):220–39.

Riede K. Global Register of Migratory Species - from Global to Regional Scales. Final Report of the R&D-Project 808 05 081. Federal Agency for Nature Conservation, Bonn, Germany. Bonn: 2004.

Rivera Hernández JM, Peña Alvarado N, Correa Vélez K, Nemeth R, Appeldoorn R, Shervette V. Queen Triggerfish Reproductive Biology in U.S. Caribbean Waters. *Trans Am Fish Soc.* 2019; 148(1):134–47. doi: <https://doi.org/10.1002/tafs.10124>.

Robins CR, Ray GC. A field guide to Atlantic coast fishes of North America. Houghton Mifflin Company. 1986:368.

Rocha LA. *Chaetodon ocellatus*. The IUCN Red List of Threatened Species 2010: e.T165611A6067709. 2010. <https://dx.doi.org/10.2305/IUCN.UK.2010-4.RLTS.T165611A6067709.en> (accessed August 19, 2020).

Rondineli G, Braga F, Tutui S, Bastos G. Dieta de *Menticirrhus americanus* (Linnaeus, 1758) e *Cynoscion jamaicensis* (Vaillant e Bocourt, 1883) (Pisces, Sciaenidae) no sudeste do Brasil estado de São Paulo. *Bull Inst Pesca.* 2007; 33(2):221–8.

Ross S, Casazza T, Quattrini A, Sulak K. Aguilliform larvae collected off North Carolina. *Mar Biol.* 2007; 150(4):681–95.

Ross SW, Rhode FC. The gobioid fishes of North Carolina (Pisces: Gobioidae). *Bull Mar Sci.* 2004; 74(2):287–323.

Ross SW, Rhode FC. Collections of ophichthid eels on the surface at night off North Carolina. *Bull Mar Sci.* 2003; 72(1):241–6.

Rueda M, Defeo O. Spatial structure of fish assemblages in a tropical estuarine lagoon: Combining multivariate and geostatistical techniques. *J Exper Mar Biol Ecol.* 2003; 296(1):93–112. doi: [https://doi.org/10.1016/S0022-0981\(03\)00319-8](https://doi.org/10.1016/S0022-0981(03)00319-8).

Saad AM, Fagundes Netto E. Aspectos da biologia reprodutiva de fêmeas de *Etropus longimanus* Norman, 1933 (Bothidae) da região de Cabo Frio, Rio de Janeiro: 1. tamanho da primeira maturação, tipo e época de desova. Bol Instit Oceanogr. 1992; 40(1–2):01–13. doi: <https://doi.org/10.1590/s0373-55241992000100001>.

Sales N. Influência da salinidade na distribuição e dieta da ictiofauna em um estuário hipersalino. 2015.

Santos DM dos. Compostos butílicos de estanho em tecidos de bagres estuarinos (Siluriformes, Ariidae) da costa sul e sudeste brasileira: monitoramento e toxicidade.

Santos FB, Castro RMC. Activity, habitat utilization, feeding behaviour, and diet of the sand moray *Gymnothorax ocellatus* (Anguilliformes, Muraenidae) in the south western Atlantic. Biota Neotrop. 2003; 3(1):1–7. doi: <https://doi.org/10.1590/s1676-06032003000100003>.

Sasekumar A, Chong VC, Leh MU, D’Cruz. Mangroves as a habitat for fish and prawns. In: Jaccarini V, Marters E, editors. The Ecology of Mangrove and Related Ecosystems. Mombasa: Springer Science+Business Media; 1992. p.265. doi: <https://doi.org/10.1017/CBO9781107415324.004>.

Schärer MT, Nemeth MI, Appeldoorn RS. Protecting a Multi-species Spawning Aggregation at Mona Island, Puerto Rico. Gulf and Caribbean Fisheries Institute Proceedings. 2010:252–9.

Schneider W. FAO species identification sheets for fishery purposes. Field guide to the commercial marine resources of the Gulf of Guinea. Prepared and published with the support of the FAO Regional Office for Africa. FAO, Rome. Rome: FAO; 1990.

Schultz YD, Favaro LF, Spach HL. Aspectos reprodutivos de *Sphoeroides greeleyi* (Gilbert), Pisces, Osteichthyes, Tetraodontidae, da gamboa do Bagaçu, Baía De Paranaguá, Paraná, Brasil. Rev Bras Zool. 2002; 19(1):65–76. doi: <https://doi.org/10.1590/s0101-81752002000100004>.

Schwamborn SHL. Dinâmica e organização trófica de assembléias de peixes associadas aos prados de capim marinho (*Halodule wrightii*) de Itamaracá, Pernambuco.

Silva JDB, Barletta M, Lima ARA, Ferreira GVB. Use of resources and microplastic contamination throughout the life cycle of grunts (Haemulidae) in a tropical estuary. Environ Pollut. 2018; 242(Pt A):1010–21. doi: <https://doi.org/10.1016/j.envpol.2018.07.038>.

Silva JTO, Lopes PRD. Notas sobre a alimentação e morfologia do aparelho digestivo de *Chloroscombrus chrysurus* (Linnaeus, 1766) Actinopterygii, Carangidae) na Praia de Ponta da Ilha (Ilha de Itaparica, Bahia). Rev Bras Zool. 2002; 4(1517–6770):179–92.

Silva Júnior CAB da, Viana AP, Frédou FL, Frédou T. Aspects of the reproductive biology and characterization of Sciaenidae captured as bycatch in the prawn trawling in the northeastern Brazil. Act Scient Biol Sci. 2015; 37(1):1–8. doi: <https://doi.org/10.4025/actascibiolsci.v37i1.24962>.

Silva RS, Carvalho KD, Pessanha ALM. Distribution and feeding ecology of three juvenile mojarra in a hypersaline tropical estuary in Northeastern Brazil. *Mar Ecol.* 2016; 37(6):1266–81. doi: <https://doi.org/10.1111/maec.12316>.

Sley A, Jarboui O, Ghorbel M, Bouain A. Food and feeding habits of *Caranx crysos* from the Gulf of Gabs (Tunisia). *J Mar Biol Ass UK.* 2009; 89(7):1375–80. doi: <https://doi.org/10.1017/S0025315409000265>.

Soares L. Trofodinâmica dos peixes Pleuronectiformes do Canal de São Sebastião. 1997.

Spach HL, Santos C, Godefroid RS, Nardi M, Cunha F. A study of the fish community structure in a tidal creek. *Braz J Biol.* 2004; 64(2):337–51. doi: <https://doi.org/10.1590/s1519-69842004000200020>.

Stefanoni MF. Ictiofauna e ecologia trófica de peixes em ambientes praias da Ilha das Peças, complexo estuarino de Paranaguá, Paraná.

Stehmann M, McEachran JD, Vergara RR. Dasyatidae. FAO species identification sheets for fishery purposes. Western Central Atlantic (Fishing Area 31). Rome: FAO Spec. Cat. Fish. Purp; 1978. p.pag.var.

STRI. *Evorthodus lyricus* 2017. biogeodb.stri.si.edu/caribbean/en/thefishes/species/4166 (accessed August 19, 2020).

Targett TE. Food Resource Partitioning by the Pufferfishes *Sphoeroides spengleri* and *S. testudineus* from Biscayne Bay, Florida. *Mar Biol.* 1978; 49:83–91.

Teixeira RL. Abundance, reproductive period, and feeding habits of eleotrid fishes in estuarine habitats of north-east Brazil. *J Fish Biol.* 1994; 45:749–61. doi: <https://doi.org/10.1111/j.1095-8649.1994.tb00941.x>.

Teixeira RL, Musick. Reproduction and food habits of the lined seahorse, *Hippocampus erectus* (Teleostei: Syngnathidae) of Chesapeake Bay, Virginia. *Rev Brasil Biol.* 2001; 61(1):79–90.

Teixeira RL, Perrone EC. Reproductive and feeding biology of freshwater 2 and estuarine populations of the opossum pipefish, *Oostethus lineatus* from 3 southeastern Brazil. *Bolet Mus Biol Mel Leit.* 1998; 8:3–12.

Temóteo TAA, Pina J, Lira AS, Sarmiento GC, Viana AP, Lucena Frédou F, et al. Alimentação de duas espécies do gênero *Caranx* capturadas no litoral de Pernambuco. 2015.

Turra A, Santos FB, Bessa E, Fernandez WS, Bernadochi LC, Denadai MR. Population Biology and Diet of the Southern Kingcroaker *Menticirrhus americanus* (LINNAEUS, 1758) (PERCIFORMES: SCIAENIDAE) in Caraguatatuba Bay, Southeastern Brazil. *Braz J Oceanogr.* 2012; 60(3):343–52.

Uyeno T, Matsuura K, Fujii E. Fishes trawled off Suriname and French Guiana. Tokyo: Japan Marine Fish Rese Res Cent; 1983.

Vasconcellos MC, Freire KF, Castello JP. Distribution patterns and feeding success of anchovy, *Engraulis anchoita*, larvae off southern Brazil. *Scientia Marina*. 1998; 62(4):385–92. doi: <https://doi.org/10.3989/scimar.1998.62n4385>.

Vasconcelos Filho A de L, Oliveira AME de. Composição e ecologia da ictiofauna do Canal de Santa Cruz (Itamaracá-PE, Brasil). *Trab Oceanograf Univ Fed PE*. 1999; 27(1):101–13.

Vasconcelos Filho ADL, Silva KC da, Acioli FD. Hábitos alimentares de *Sphoeroides testudineus* (Linnaeus, 1758) (Teleostei: Tetraodontidae), no Canal de Santa Cruz, Itamaracá-PE. *Trop Oceanogr*. 1998; 26(1):145–57. doi: <https://doi.org/10.5914/tropocean.v26i1.2759>.

Vasconcelos Filho AL, Neumann-Leitão S, Eskinazi-Leça E, Oliveira AME de. Hábitos alimentares de peixes consumidores secundários do Canal de Santa Cruz, Pernambuco, Brasil. *Trop Oceanogr*. 2010; 38(2):122–9. doi: <https://doi.org/10.5914/tropocean.v38i2.5166>.

Vasconcelos Filho AL, Neumann-Leitão S, Eskinazi-Leça E, Schwamborn R, Oliveira AME, Paranaguá MN. Trophic interactions between fish and other compartment communities in a tropical estuary in Brazil as indicator of environmental quality. *Transac Ecol Environ*. 2003; 63.

Vasconcelos-Filho AL. Alimentação da sardinha-bandeira, *Ophistonema oglinum* (LeSueur, 1817) no Canal de Santa Cruz. *Trab Oceanogr*. 1979; 14:105–16.

Vasconcelos-Filho AL, Oliveira AME. Ictiofauna. Gerenciamento participativo de estuários e manguezais. Recife: Editora Universitária; 2000. p.252.

Vaslet A, France C, Phillips DL, Feller IC, Baldwin CC. Stable-isotope analyses reveal the importance of seagrass beds as feeding areas for juveniles of the speckled worm eel *Myrophis punctatus* (Teleostei: Ophichthidae) in Florida. *J Fish Biol*. 2011; 79(3):692–706. doi: <https://doi.org/10.1111/j.1095-8649.2011.03052.x>.

Whitehead PJP. Vol. 7. Clupeoid fishes of the world (Suborder Clupeioidi). An annotated and illustrated catalogue of the herrings, sardines, pilchards, sprats, shads, anchovies and wolf-herrings. Rome: FAO Fish. Synop.; 1985.

Williams EH, Mignucci-Giannoni AA, Bunkley-Williams L, Bonde RK, Self-Sullivan C, Preen A, et al. Echeneid-sirenian associations, with information on sharksucker diet. *J Fish Biol*. 2003; 63(5):1176–83. doi: <https://doi.org/10.1046/j.1095-8649.2003.00236.x>.

Winemiller KO, Ponwith BJ. Comparative of ecology of eleotrid fishes in Central American coastal streams. *Environ Biol Fish*. 1998; 53:373–84.

Winik S, Carneiro MH, Mendonca JT. Alimentação da guaivira *Oligoplites saliens* (Bloch, 1793) (Perciformes: Carangidae) proveniente da pesca na região de Cananéia-SP.

WoRMS EB. World Register of Marine Species 2020.

Wyanski DM, Targett TE. Development of transformation larvae and juveniles of *Ctenogobius boleosoma*, *Ctenogobius shufeldti*, and *Gobionellus oceanicus* (Pisces: Gobiidae) from Western North Atlantic estuaries, with notes on early life history. *Bull Mar Sci.* 2000; 67(2):709–28.

Yáñez-Arancibia A. Ecología de la zona costera. vol. Yáñez-Arancibia A. Mexico: AGT; 1986.

Zacardi DM, Bittencourt SCS, Nakayama L. O Ictioplâncton e Sua Relação com a Variação Diária e os Ciclos de Marés no Estuário Amazônico. *Biot Amaz.* 2016; 6(2):32–40. doi: <https://doi.org/10.18561/2179-5746/biotaamazonia.v6n2p32-40>.

Zahorcsak P, M SRA, Sazima I. Feeding biology of a guild of benthivorous fishes in a sandy shore on south-eastern Brazilian coast. *Rev Bras Biol.* 2000; 60(3):511–8. doi: <https://doi.org/10.1590/s0034-71082000000300016>.

Table II: Fishing gears used throughout the sampling sites.

Estuaries	Fishing Gears
Santa Cruz	Block Net and Seine
Jaguaribe	Beach Seine
Suape	Block Net and Beach Seine
Sirinhaém	Beach Seine and Block Net
Maracaípe	Beach Seine
Rio Formoso	Block Net

**4.1. Estuarine fishes of Northeast Brazil with notes on species ecology,
conservation, and updated checklist**

Estuarine fishes of Northeast Brazil with notes on species ecology, conservation, and updated checklist

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Abstract

Estuaries are dynamic physical-chemical unique environments, being among the most productive ecosystems on Earth as well as an important ecotone. The feeding mode (FMFG), estuarine use functional groups (EUFG) and a conservation status, according to the categories used in the International Union for Conservation of Nature Red List of threatened species and Chico Mendes Institute for Biodiversity Conservation were assigned to each fish species reported in estuaries along the coast of Northeast Brazil, to better understand the structure and functioning of the studied areas. A total of 339 species, distributed in 37 orders and 89 families were identified in the studied estuaries. Marine stragglers and migrants were 77% of the total and zoobenthivorous and piscivorous fishes represented 72% of the total number of species. Higher similarities were found between the states of Maranhão, Ceará, Paraíba, Pernambuco and Bahia, and a second group that

englobed the estuaries of Rio Grande do Norte based on the multivariate analysis. The present study provides an updated checklist with records on habitat and conservation status that might support and increase the knowledge in fisheries management, development, and conservation of marine aquatic resources in coastal areas of Northeast Brazil.

Key words

Guilds, IUCN, ICMbio, ichthyofauna, biodiversity, sustainability

Introduction

Estuaries are dynamic physical-chemical unique environments, being among the most productive ecosystems on Earth (Mateus et al. 2008; Costanza et al. 2014; Vasconcelos et al. 2015; Loureiro et al. 2016). They also perform crucial ecological functions such as pollutant filtration, flood control, biodiversity preservation, and nutrient recycling (Harrison and Whitfield 2021; Roshni et al. 2021a). According to Zapata et al. (2018a), tropical estuaries hold an important role on Earth, given their great rates of primary and secondary production and microhabitats' s diversity. As a consequence of the growth of anthropogenic actions such as urbanization, expansion of industries, and aquaculture, the structure and functioning of these ecosystems are changing (e.g. modification in the sediment's transport caused by alterations in estuary's morphology) (Zapata et al. 2018b; Lessa 2020; Roshni et al. 2021b), threatening the ecological function of these ecosystems which is directly important to the well-being of the society (Blaber 2000), giving their multiples ecosystem services.

The Brazilian coastline has different characteristics in its interface zone from the continent to the ocean (Bernardes et al. 2012). The northeast region of Brazil comprehends nine coastal states with an extension of almost 3400 km (Pinheiro et al. 2008), being the longest coastline of the country, encompassing a high diversity of oceanographic and geomorphologic characteristics and fish communities (Pinheiro et al. 2008; Barletta and Lima 2019), which tend to fluctuate throughout the year due to the interannual climate variability, temperature, currents, and salinity. Studies aiming at characterizing the structure of fish communities in estuaries are extremely important to understand their functioning and influence in the ecosystem (Potter et al. 2011; Bruno et al. 2013; Lessa 2020), but are still fragmented and punctual.

In this study, we provide an estimation of the fish biodiversity along 45 estuaries in the northeast coast of Brazil based on an extensive, but not exhaustive, literature review involving research from the 70s until nowadays. Compiling this information is important for a better comprehension of the importance of estuaries for biodiversity and maintenance of daily lives of traditional populations along with the aesthetic point of view on fish diversity in the Northeast estuaries of Brazil.

Material and Methods

The study area comprehends 45 estuaries located along the nine states of the Northeast coastal region of Brazil (Fig. 1). A literature review based on articles, books, monographs, dissertations, and theses was used to compile a checklist.

According to Lins Oliveira et al. (1999), the studied estuaries comprehends the São Marcos Bay in the state of Maranhão to Todos os Santos Bay, in the state of Bahia. It can be divided into two main subregions. The first one is the semi-arid coast located in the northwest of Calcanhar Cape and the second is known as the oriental Northeastern Coast, located from Calcanhar Cape to Todos os Santos Bay. The semi-arid coast has mobile and fixed dunes, natural beach trays, and sandstones. This region presents strong winds that are responsible for shaping the Lençóis Maranhenses, a developed eolian dune fields (Lins Oliveira et al. 1999; Suguio et al. 2011). The geological characteristics of the oriental Northeastern Coast are basically within the same scope as the first area, however with the domination of beach-rick reefs (Suguio et al. 2011).

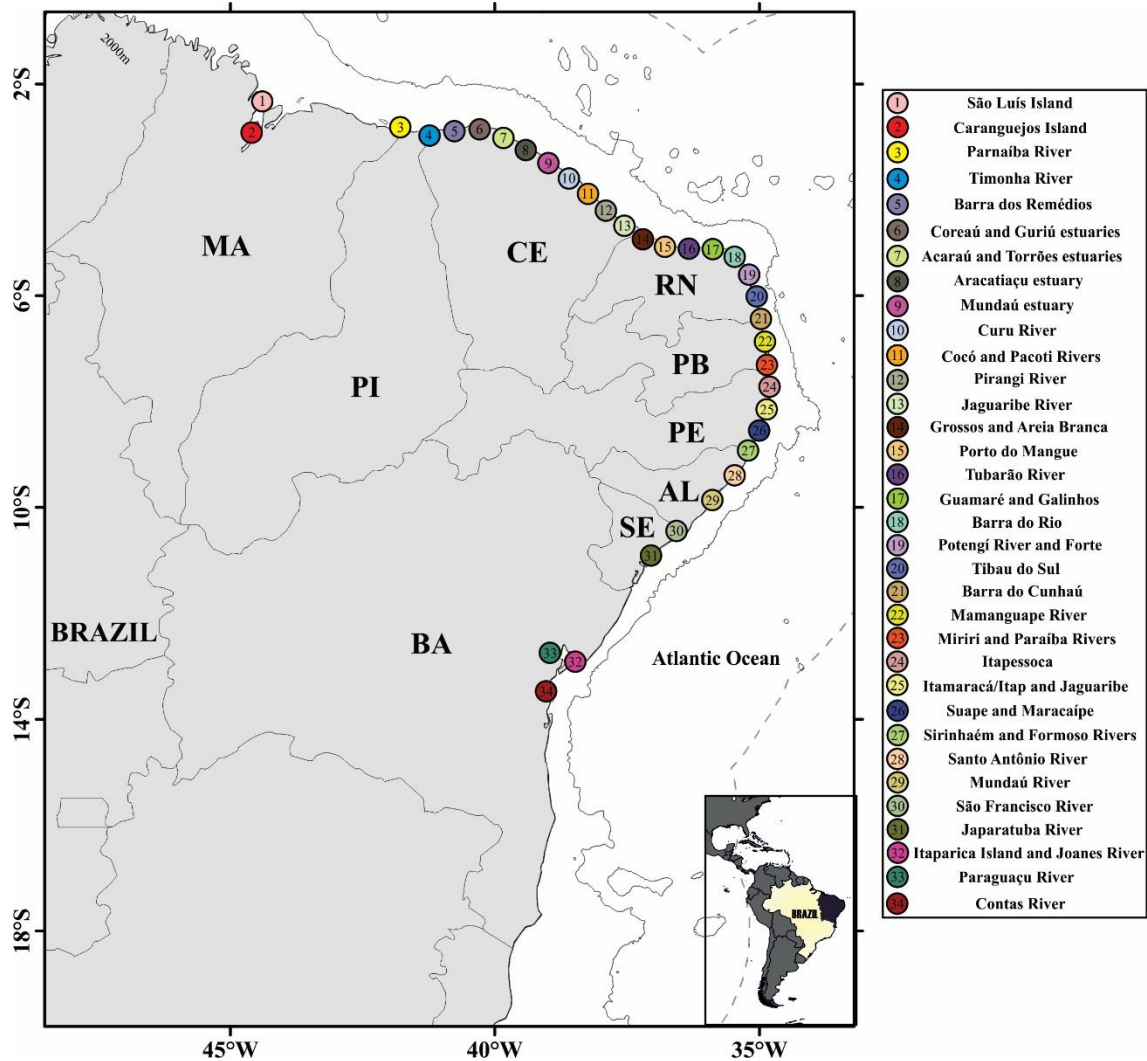


Fig. 1 Estuaries of São Luís (1) and Caranguejos Islands (2)- MA, Parnaíba (3) and Timonha Rivers (4) - PI, Barra dos Remédios (5); Coreaú and Guriú (6); Acaraú and Torrões (7); Aracatiaçu (8); Mundaú (9); Curu River (10); Cocó and Pacoti Rivers (11); Pirangi River (12); Jaguaribe River (13) – CE, Grossos and Areia Branca (14); Porto do Mangue (15); Tubarão River (16); Guamaré and Galinhos (17); Barra do Rio (18); Potengí River and Forte (19); Tibau do Sul (20); Barra do Cunhaú (21) – RN, Mamanguape River (22); Miriri and Paraíba Rivers (23) – PB, Itapessoca (24); Itamaracá/Itapissuma and Jaguaribe Rivers (25); Suape and Maracaípe (26); Sirinhaém and Formoso Rivers (27) – PE, Santo Antônio River (28); Mundaú River (29) – AL, São Francisco River (30); Japarutuba River (31) – SE, Itaparica Island and Joanes River (32); Paraguaçu River (33) and Contas River (34) – BA located in Northeast Brazil.

According to the systematic organization proposed by Nelson et al. (2016), species richness is presented in three levels: order, family, and species. The nomenclature

of species was revised through the Eschmeyer Fish Catalogue (Fricke, R., Eschmeyer, W. N. & Van der Laan 2020) and the World Register of Marine Species (WoRMS 2020). In addition, the original name of the species found within the literature is included (Supplementary Material Table II).

Fish species were analyzed as presence/absence data and classified to their use of estuaries based on the functional guild approach (Elliott et al. 2007) (Supplementary Material Table I): the Estuarine Use Functional Group (EUFG) and the Feeding Mode Functional Group (FMFG) (Elliott et al. 2007). EUFG allows the understanding of the use of estuaries by fishes and how important these environments are. Concerning this guild, species were categorized as marine stragglers (MS), marine migrants (MM), estuarine species (ES), freshwater species (FS), freshwater migrants (FM), amphidromous (AM), anadromous (AN) and catadromous species (CA) (Elliott et al. 2007). FMFG characterizes the feeding habit of a species, also based on Elliott et al. (2007), such as zoobenthivore (ZB), piscivore (PV), zooplanktivore (ZP), herbivore (HV), omnivore (OV), detritivore (DV) and opportunistic (OP). An extensive literature review, totalizing more than 230 references, was used to define the functional guilds (Supplementary Material Table I) (see Elliott et al. 2007 for a detailed description of each trophic category).

According to the categories based in the International Union for Conservation of Nature (IUCN) Red List criteria of threatened species, as well as those categorized at Brazilian level (ICMbio 2018), fish species in this study were assigned to 10 categories: extinct (EX), regionally extinct (RE), extinct in the wild (EW), critically endangered (CR), endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE). A circle barplot was made for a better visualization of how these categories were divided, with the highest and lowest observations being displayed across the groups.

A heatmap displaying estuaries x guild proportion along with an Agglomerative Hierarchical Cluster (AHC) were performed using the Ward's minimum variance method based on a Euclidian distance matrix. Furthermore, pursuing a better understanding between similarities among the fish assemblages and estuaries, a Non-metric Multidimensional Scaling (nMDS) using a Sorensen similarity matrix from species presence/absence data, and a cluster analysis using the average linkage method were

carried out. Finally, a permutational multivariate analysis of variance (PERMANOVA) was performed to show differences of richness amid estuaries (Clarke and Warwick 2001). All statistical analyses were performed using the *vegan* (Oksanen et al. 2019), *ggplot2* (Wickham 2016), *viridis* (Garnier et al. 2021) and *stats* (R Core 2019) packages from Team (2020).

Results

A total of 339 fish species, distributed in 37 orders and 89 families were identified in the studied estuaries (Table I, II, III, IV and V). The richest family was Sciaenidae comprising 25 species (7 % of total), followed by Carangidae and Gobiidae (16 species), Engraulidae (14 species), Haemulidae and Serranidae (13 species), Ariidae (12 species) and Paralichthyidae (11 species). The remaining 82 families had less than 10 species each. The state of Pernambuco was the most diverse in terms of species, comprehending 237, followed by Ceará (163 species), Bahia (156 species), Rio Grande do Norte (154 species), Paraíba (147 species), Maranhão (124 species), Alagoas (58 species), Piauí (53 species) and Sergipe (43 species).

Most of species were classified as Least Concern according to the IUCN (74.7%) (Fig. 2) and ICMbio (72.6%) (Fig. 3) conservation status. A total of 21 species (IUCN) and 20 species (ICMbio) were identified under some degree of threat (Fig. 2 and 3): seven as EN (*Sphyrna tiburo* (IUCN), *Pseudobatos percellens* (IUCN), *Hypanus marianae* (IUCN), *Aetobatus narinari* (IUCN) and *Mobula hypostoma* (IUCN), *Hyporthodus nigrutus* (ICMbio) and *Pogonias cromis* (ICMbio); seven as CR (*Carcharhinus porosus* (IUCN and ICMbio), *Isogomphodon oxyrinchus* (IUCN and ICMbio), *Sphyrna lewini* (IUCN and ICMbio), *Sphyrna tudes* (IUCN and ICMbio), *Pristis pristis* (IUCN and ICMbio), *Fontitrygon geijskesi* (IUCN), *Epinephelus itajara* (ICMbio); and 15 as VU (*Ginglymostoma cirratum* (IUCN and ICMbio), *M. hypostoma* (ICMbio), *Rhizoprionodon lalandii* (IUCN), *Rhizoprionodon porosus* (IUCN), *Rhinoptera bonasus* (IUCN), *Megalops atlanticus* (IUCN and ICMbio), *Sciades parkeri* (IUCN and ICMbio), *Achirus mucuri* (ICMbio), *Hippocampus erectus* (IUCN and ICMbio), *Hippocampus reidi* (ICMbio), *Sparisoma axillare* (ICMbio), *E. itajara* (IUCN), *Epinephelus marginatus* (IUCN and ICMbio), *Mycteroperca bonaci* (ICMbio) and *Lutjanus cyanopterus* (IUCN and ICMbio). In addition, 25 species were classified as NT:

Carcharhinus leucas (IUCN and ICMbio), *Carcharhinus limbatus* (IUCN and ICMbio), *R. lalandii* (ICMbio), *Narcine brasiliensis* (IUCN), *Hypanus americanus* (IUCN), *Hypanus guttatus* (IUCN), *Hypanus say* (IUCN), *Gymnura micrura* (IUCN and ICMbio), *Bagre bagre* (ICMbio), *Dormitator maculatus* (ICMbio), *Mugil liza* (ICMbio), *Hyporhamphus unifasciatus* (ICMbio), *Albula vulpes* (IUCN), *H. reidi* (IUCN), *H. nigrinus* (IUCN), *Sparisoma amplum* (ICMbio), *M. bonaci* (IUCN), *Mycteroperca venenosa* (IUCN), *Lutjanus analis* (IUCN and ICMbio), *Lutjanus jocu* (ICMbio), *Lutjanus synagris* (IUCN and ICMbio), *Ocyurus chrysurus* (ICMbio), *Cynoscion acoupa* (ICMbio), *Balistes vetula* (IUCN and ICMbio) and *Aluterus monocerus* (ICMbio). It was also observed 40 (ICMbio) and 10 (IUCN) species were recognized as DD (Table I, II, III, IV and V).

Assessing by estuary, four showed a higher percentage than the others within the endangered species categories; the estuary of São Luís in the state of Maranhão (12% - IUCN and 10% - ICMbio of the total number of species in the estuary); the Guamaré estuary in the state of Rio Grande do Norte (7% - ICMbio of the total number of species in the estuary); Pirangí in the state of Ceará and Porto do Mangue in Rio Grande do Norte, both presented a percentage of 6% of the total number of species in each estuary (ICMbio). São Luís had 13% (ICMbio) and 3% (IUCN) of its species classified as DD, while the estuary of Mamanguape had 12% of its species categorized as DD (ICMbio).

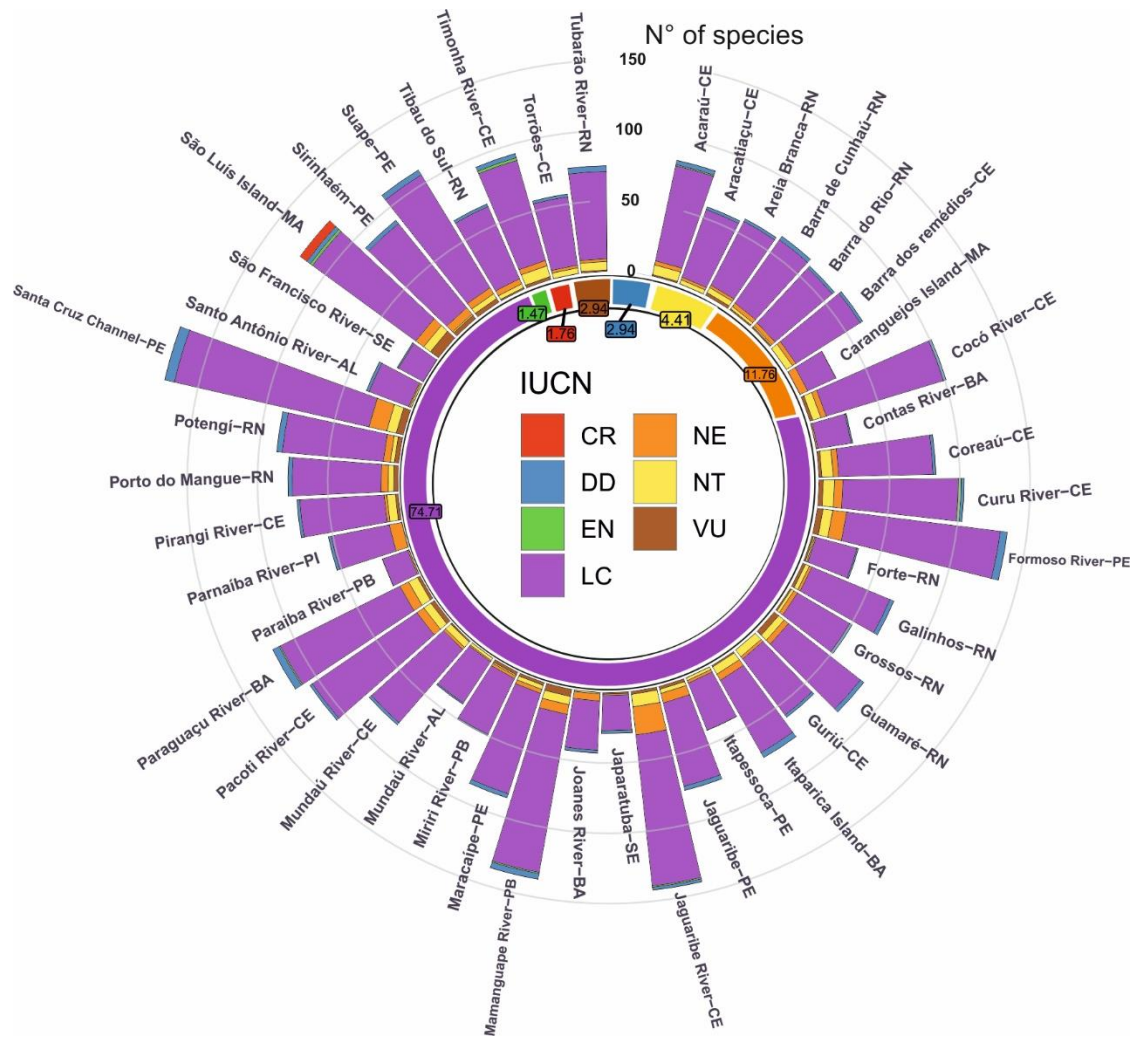


Fig. 2 Number of fish species from each IUCN categories for the studied estuaries along the northeastern coast of Brazil. Endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

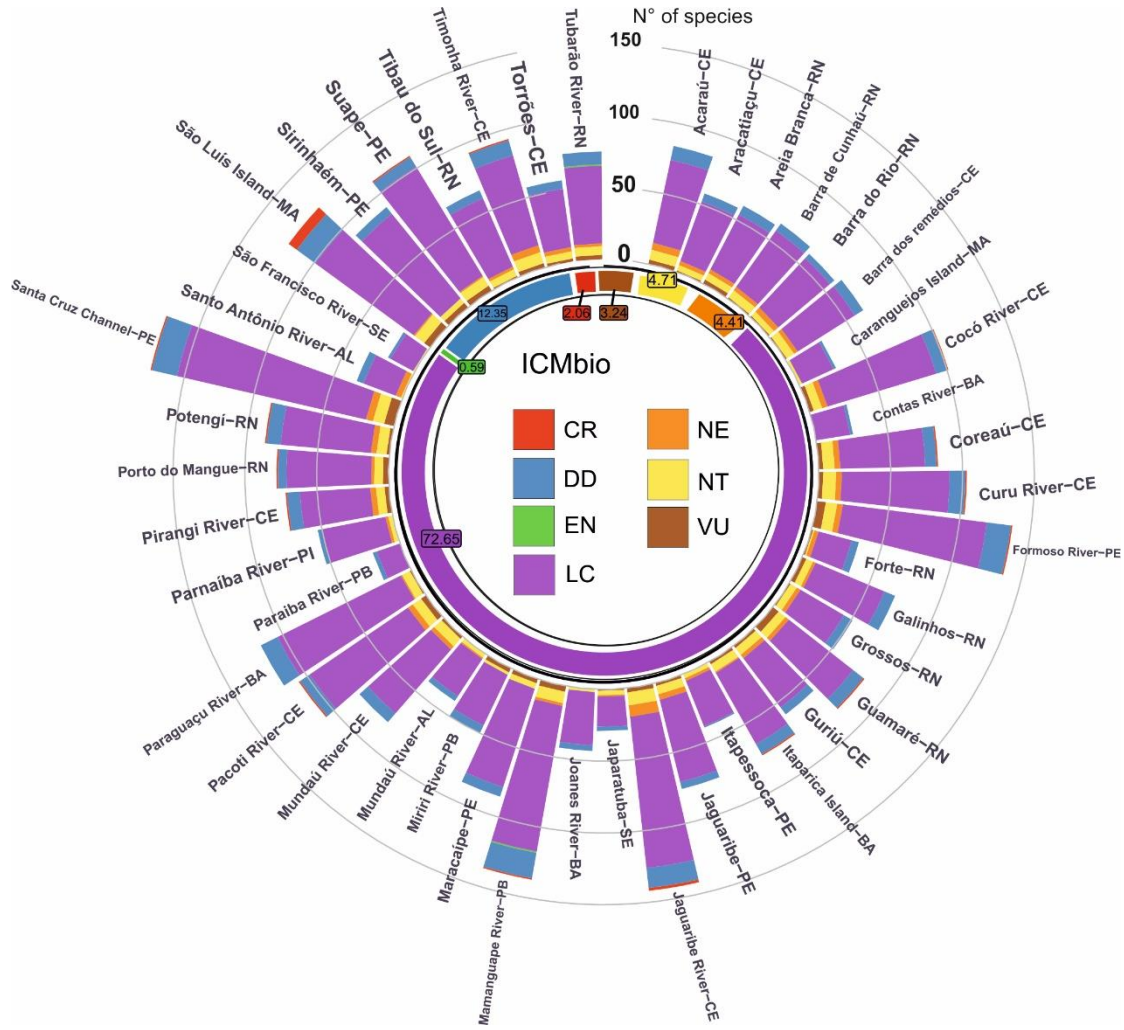


Fig. 3 Number of fish species for each ICMbio categories for the studied estuaries along the northeastern coast of Brazil. Endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

Considering the Estuarine Use Functional Group (EUFG), most of the species belonged to the marine ecosystem: marine stragglers (48% of the total number of species) and marine migrants (29.1% of the total number of species) (Supplementary Material Table I, Fig. 4). Estuarine species were 14% (47 species), while freshwater species was only 7.5% (26 species) and freshwater migrants were only 2% (8 species). It was also found two amphidromous species (*Awaous tajasica* present only on RN) and *Dormitator maculatus* (present in three of the studied states: CE, RN and PE), and one anadromous species (*Microphis brachyurus brachyurus* present on two of the studied states: CE and PB), one catadromous species (*Mugil liza* present on seven of the studied states: MA, CE, RN, PE, AL, SE and BA). When evaluating each estuary alone, the same result was found

when considering the EUFG. Most of the species were classified as marine stragglers and migrants varying from 18% (Caranguejos estuary/MA) to 56% (Mundaú estuary/AL) of the total number of species within the estuary itself.

Considering the Feeding Mode Functional Group (FMFG), most of the species were classified as zoobenthivorous (44%), followed by piscivorous (28%) and zooplantivorous fishes (9%) (Supplementary Material Table I). When analyzing by estuary, this same pattern of feeding habit was also found (Fig. 5). Due to the lack of information, a total of 17 species were not assigned to any functional group in either estuarine habitat use pattern or feeding habit.

Two main groups were formed in the dendrograms' groupings from the EUFG and FMFG. In the first one, for the EUFG, it was included all the estuaries from the state of Ceará, as well as one estuary from Rio Grande do Norte (Potengi); two estuaries from Pernambuco (Santa Cruz and Formoso River); both estuaries from Maranhão; the only estuary from Piauí and one estuary from Bahia (Joanes River). The second group was formed by all of estuaries from the state of Paraíba and Alagoas; the remaining estuaries of Pernambuco (Itapessoca, Suape, Sirinhaém, Maracaípe and Jaguaribe), Paraíba (Forte, Galinhos, Barra do Rio, Tubarão, Guamaré, Galinhos, Grossos, Areia Branca, Cunhaú and Tibaú do Sul) and Bahia (Paraguaçu, Contas and Itaparica), and one estuary from Sergipe (São Francisco). Observing the groupings from the FMFG, the first group was formed by three estuaries from Ceará (Guriú, Torrões and Aracatiaçu); two estuaries from Paraíba (Paraíba and Miriri), Alagoas (Santo Antônio and Mundaú), Bahia (Contas and Joanes), Sergipe (São Francisco and Japarutuba), and Pernambuco (Itapessoca and Jaguaribe), the only estuary from Piauí and one estuary from Maranhão (Caranguejos). The second group had the remaining estuaries from six states (Maranhão, Ceará, Rio Grande do Norte, Paraíba, Pernambuco and Bahia).

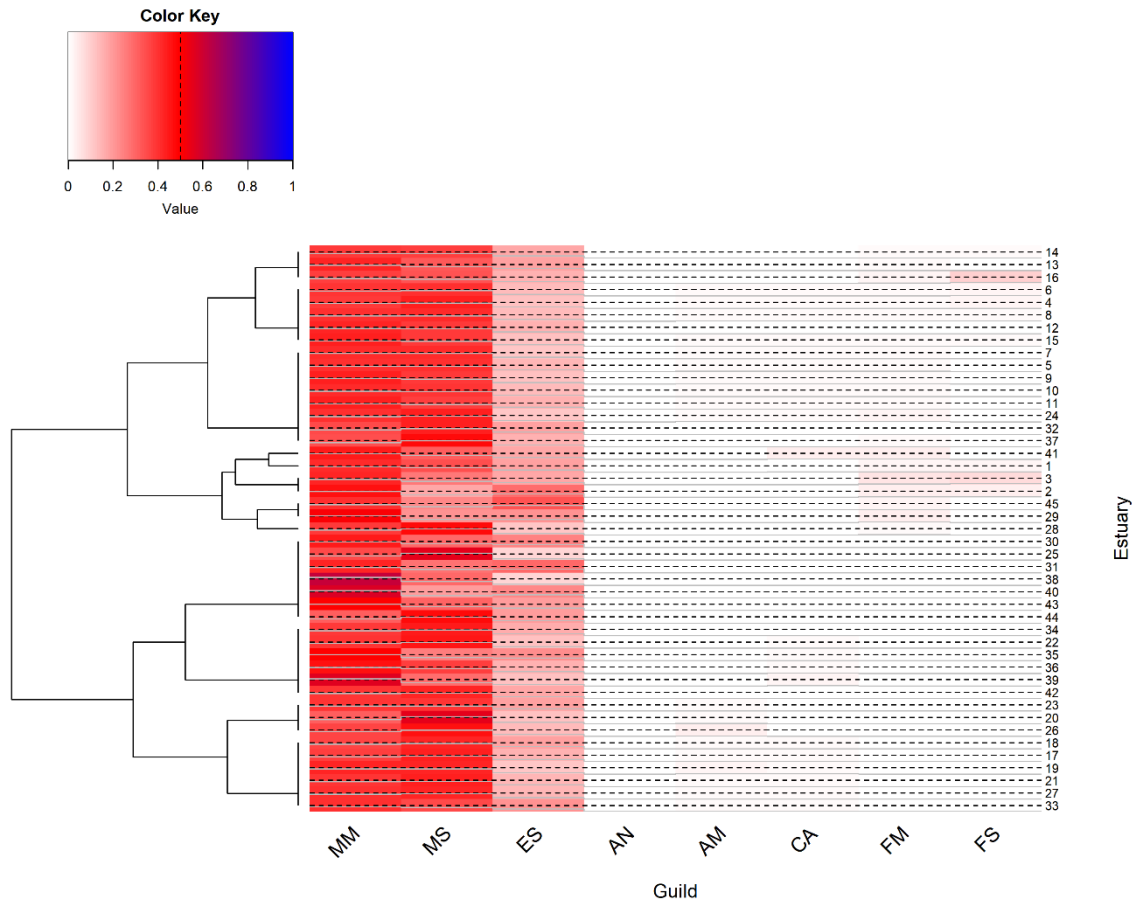


Fig. 4 Percentage of each EUFG categories marine migrant (MM), marine stragglers (MS), estuarine fishes (ES), anadromous species (AN), amphidromous (AM), catadromous (CA), freshwater migrants (FM) and freshwater species (FS) for the estuaries of São Luís Island (1), Caranguejos Island (2), Parnaíba River (3), Timonha River (4), Barro dos Remédios (5), Coreaú (6), Guriú (7), Acaraú (8), Torrões (9), Aracatiaçu (10), Mundaú River- CE (11), Curu River (12), Cocó River (13), Pacoti River (14), Pirangi River (15), Jaguaribe River – CE (16), Grossos (17), Areia Branca (18), Porto do Mangue (19), Tubarão River (20), Guamaré (21), Galinhos (22), Barra do Rio (23), Potengí (24), Forte (25), Tibau do Sul (26), Barra de Cunhaú (27), Mamanguape River (28), Miriri River (29), Paraíba River (30), Itapessoca (31), Santa Cruz (32), Jaguaribe- PE (33), Suape (34), Sirinhaém (35), Maracaípe (36), Formoso River (37), Santo Antônio River (38), Mundaú River – AL (39), São Francisco River (40), Japarutuba (41), Paraguaçu River (42), Contas River (43), Itaparica Island (44) and Joanes River (45).

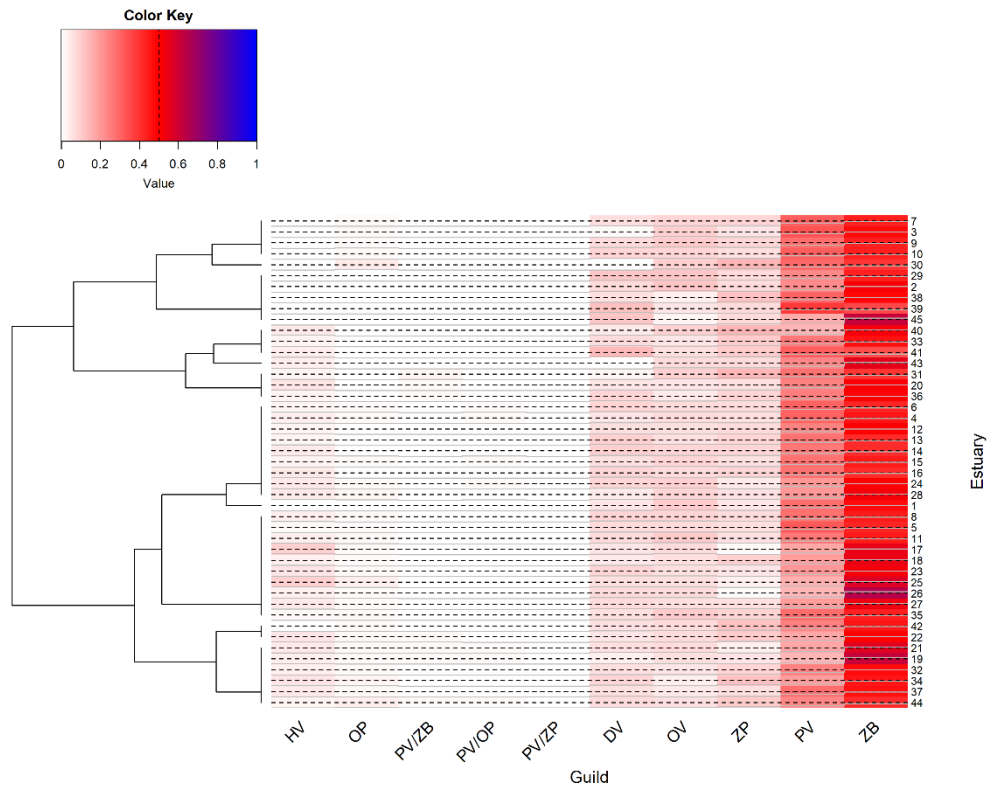


Fig. 5 Percentage of each FMFG categories zoobenthivore (ZB), piscivore (PV), zooplanktivore (ZP), herbivore (HV), omnivore (OV), detritivore (DV) and opportunistic (OP) for the estuaries of São Luís Island (1), Caranguejos Island (2), Parnaíba River (3), Timonha River (4), Barro dos Remédios (5), Coreau (6), Guriú (7), Acaraú (8), Torrões (9), Aracatiaçu (10), Mundaú River- CE (11), Curu River (12), Cocó River (13), Pacoti River (14), Pirangi River (15), Jaguaribe River – CE (16), Grossos (17), Areia Branca (18), Porto do Mangue (19), Tubarão River (20), Guamaré (21), Galinhos (22), Barra do Rio (23), Potengi (24), Forte (25), Tibau do Sul (26), Barra de Cunhaú (27), Mamanguape River (28), Miriri River (29), Paraíba River (30), Itapessoca (31), Santa Cruz (32), Jaguaribe- PE (33), Suape (34), Sirinhaém (35), Maracaípe (36), Formoso River (37), Santo Antônio River (38), Mundaú River – AL (39), São Francisco River (40), Japarutuba (41), Paraguaçu River (42), Contas River (43), Itaparica Island (44) and Joanes River (45).

The nMDS and Cluster analysis based on the taxonomic composition of each estuary showed similar results between groupings. The cluster analysis, with the resemblance level of 40%, showed twelve groups, however, it is possible to see two wider

ones: one formed by the estuaries of Rio Grande do Norte and the second one by the estuarine regions of Maranhão, Ceará, Paraíba, Pernambuco and Bahia states (Figure 6). Even though the nMDS did not clearly separate the wider groups observed in the cluster analysis, it is still possible to see the proximity between the groupings within the estuaries located in the same state and the ones between the estuaries of Ceará, Pernambuco and Rio Grande do Norte (Figure 7).

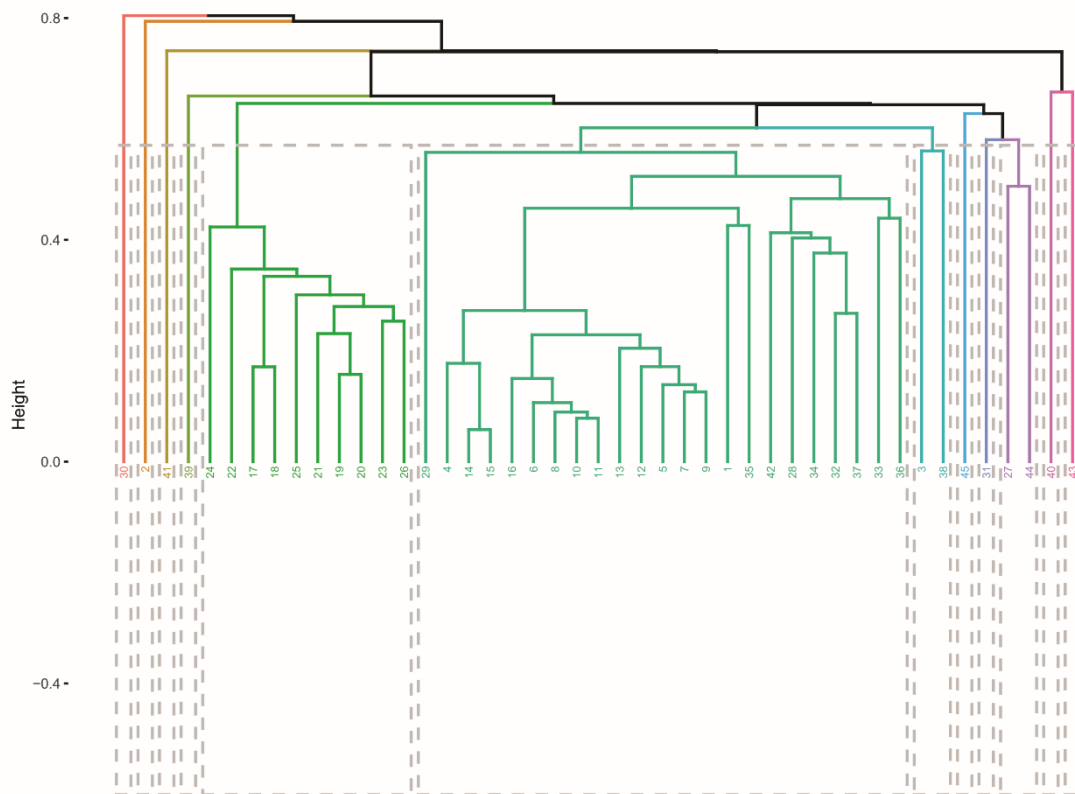


Fig. 6 Cluster analysis based on species records by estuaries. São Luís Island (1), Caranguejos Island (2), Paraíba River (3), Timonha River (4), Barro dos Remédios (5), Coreaú (6), Guriú (7), Acaraú (8), Torrões (9), Aracatiaçu (10), Mundaú River- CE (11), Curu River (12), Cocó River (13), Pacoti River (14), Pirangi River (15), Jaguaribe River – CE (16), Grossos (17), Areia Branca (18), Porto do Mangue (19), Tubarão River (20), Guamaré (21), Galinhos (22), Barra do Rio (23), Potengi (24), Forte (25), Tibau do Sul (26), Barra de Cunhaú (27), Mamanguape River (28), Miriri River (29), Paraíba River (30), Itapessoca (31), Santa Cruz (32), Jaguaribe- PE (33), Suape (34), Sirinhaém (35), Maracápe (36), Formoso River (37), Santo Antônio River (38), Mundaú River – AL (39),

São Francisco River (40), Japaratuba (41), Paraguaçu River (42), Contas River (43), Itaparica Island (44) and Joanes River (45).

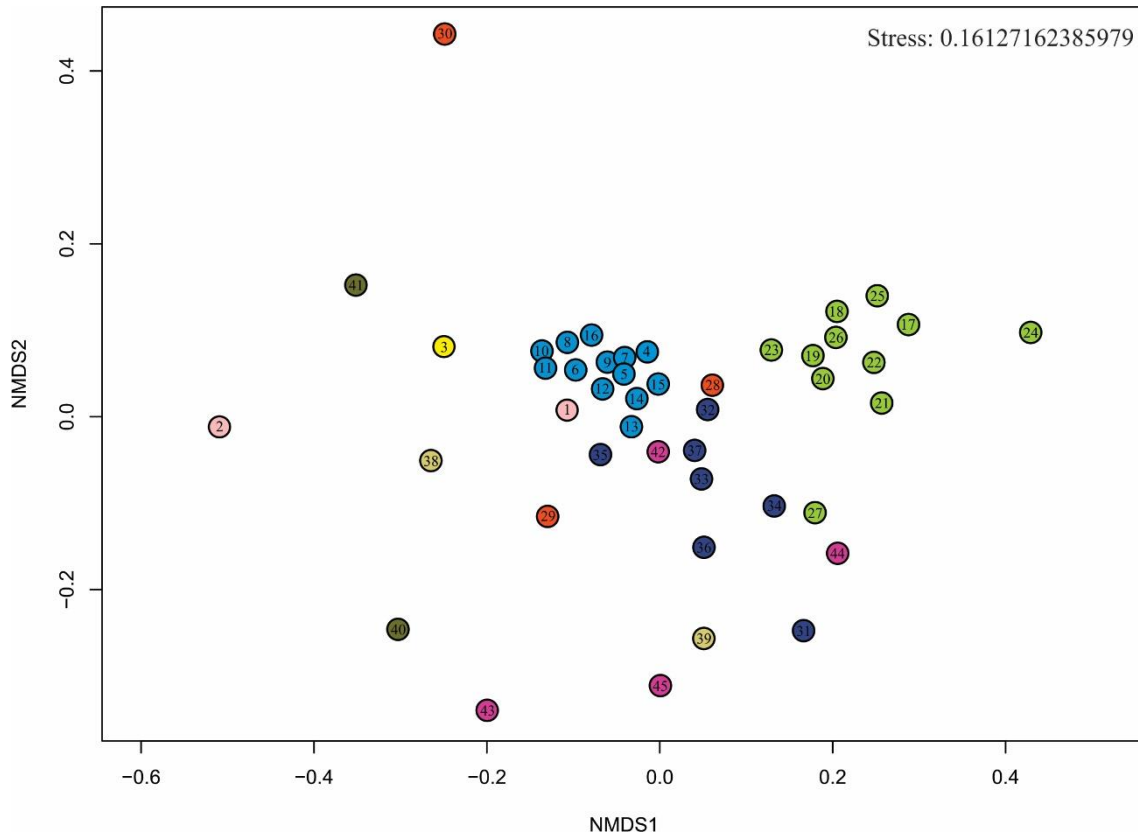


Fig. 7 Non-metric Multidimensional Scaling (nMDS) based on species records by estuaries. São Luís Island (1), Caranguejos Island (2), Parnaíba River (3), Timonha River (4), Barro dos Remédios (5), Coreaú (6), Guriú (7), Acaraú (8), Torrões (9), Aracatiaçu (10), Mundaú River- CE (11), Curu River (12), Cocó River (13), Pacoti River (14), Pirangi River (15), Jaguaribe River – CE (16), Grossos (17), Areia Branca (18), Porto do Mangue (19), Tubarão River (20), Guamaré (21), Galinhos (22), Barra do Rio (23), Potengí (24), Forte (25), Tibau do Sul (26), Barra de Cunhaú (27), Mamanguape River (28), Miriri River (29), Paraíba River (30), Itapessoca (31), Santa Cruz (32), Jaguaribe-PE (33), Suape (34), Sirinhaém (35), Maracaípe (36), Formoso River (37), Santo Antônio River (38), Mundaú River – AL (39), São Francisco River (40), Japaratuba (41), Paraguaçu River (42), Contas River (43), Itaparica Island (44) and Joanes River (45). States divided by colors: light blue: Ceará; green: Rio Grande do Norte; pink: Bahia; light

pink: Maranhão; yellow: Piauí; dark yellow: Alagoas; purple: Pernambuco; orange: Paraíba; and moss: Sergipe.

Discussion

Estuaries are important not only for their role as feeding, breeding, spawning, and sheltering grounds, but are also among the world's most dynamic, productive, and ecologically notable ecosystems (Barletta et al. 2003; Vendel and Chaves 2006; NOAA 2019; Melo et al. 2021).

Our results identified a total of 339 fish species within the nine states located in northeastern Brazil evidencing a diverse and rich ichthyofauna even when compared with other studies including the whole Brazilian coast, such as Vilar and Joyeux (2018b) that found 796 estuarine fishes and 451 reported by Paiva et al. (2013). The number of fish species can be considered relatively low when compared with those studies, however, this reaffirms that the Northeast estuaries are understudied, as well as have a lower sampling effort which can be one of the reasons for this difference. Croakers are used for human consumption, in aquariums, and as part of the diet from other fish species. Jacks are used in aquariums and as part of human consumption. Gobies are mainly used in aquariums due to their different coloration and body format. Anchovies are largely used for human consumption and as animal food for other fish species. Haemulids and serranids are used in aquariums as well as for human consumption. Catfishes are mostly used as human consumption, and flounders are used as animal feed and human consumption (Sampaio De Souza and Júnior; Chao et al. 2015; Spier et al. 2018; Vicente et al. 2020). That result was also found in estuaries along the southeast and south of Brazil (Catelani et al. 2014; Santos et al. 2015; Spier et al. 2018), however, with a higher incidence of mullets (Mugilidae) and mojarras (Gerreidae) in an estuary in Santa Catarina (Branco et al. 2011), and mojarras and croakers in an estuary in São Paulo (Santos et al. 2015). It is also noteworthy to mention that these are the most common and important families found in estuarine and coastal systems (Passos et al. 2013a; Rotundo et al. 2020).

In the present study, *Sphoeroides testudineus* was found in 40 estuaries, *Centropomus undecimalis* and *Diapterus rhombeus* were found in 39, *Mugil curema* in 38, *Eucinostomus melanopterus* and *Lutjanus jocu* in 37, *Atherinella brasiliensis*,

Bathygobius soporator and *Pomadasys corvinaeformis* were found in 34, *Menticirrhus americanus* was found in 33, *Caranx latus* and *Micropogonias furnieri* were found in 31, and *Chaetodipterus faber*, *Cynoscion acoupa*, *Diapterus auratus*, *Eucinostomus argenteus*, *Mugil curvidens*, *Mugil liza*, *Oligoplites saurus* and *Opisthonema oglinum* were found in 30 estuaries. All of these species were present in either all states or at least seven of them. These results demonstrate the wide range in the spatial distribution of the estuarine ichthyofauna. On the other hand, the slender suckerfish (*Phtheirichthys lineatus*) was only found in one of the studied estuaries (Rio Formoso – PE). This can be possibly explained by the fact that this species inhabits coral reefs and that this region is part of the largest environmental protection area (APA Costa dos Corais) which has dense and diverse coastal reefs (Ferreira and Maida 2006).

Guilds have been extensively used to simplify the understanding of how aquatic environments function and their structure (Elliott et al. 2007a; Passos et al. 2013a; Passarone et al. 2019). According to our results, most of species were marine stragglers and migrants emphasizing the high tolerance that these fishes have, being able to endure wide salinity and depth variations (Franco et al. 2008; Passos et al. 2013a; Camara et al. 2020). Species that are more dependent on estuarine areas are more likely to resist high ranges of environmental characteristics having a vast distribution when compared with other groups (e.g., freshwater that are more confined to areas with riverine input) (Reis et al. 2016; Camara et al. 2020).

Estuaries tend to have a diverse number of trophic levels, as observed in the present study, but zoobenthivorous and piscivorous species prevailed among them, as previously reported by Blaber (2000). This may be explained by the prevalence of rich nutrients in the sediment along with the high primary production carried through the organic material transport from adjacent rivers to the ocean (Medeiros 2013b; Mourão et al. 2014b; Affe et al. 2021), that gives support to the estuarine trophic levels boosting the energy quantity to higher trophic levels (Yee et al. 2007; Medeiros 2013b).

The fish community was divided into twelve clusters with two larger ones separating the state of Rio Grande do Norte into one group and the states of Maranhão, Ceará, Paraíba, Pernambuco and Bahia into another one. This separation may be due to numerous reasons, the main one being the lack of available data for many of the studied estuaries or to low sampling effort which can underestimate the number of species in

some cases. Some states have accessible information for only one or two estuaries as those in the state of Alagoas and may even be restricted to one reference in some cases having a low sample size like the state of Piauí. This situation evidences the importance of compiling ‘grey literature’ data since a lack of information, not only in a wider but also local gradient, was shown in the present study.

The majority of species reported in the studied estuaries was categorized as Least Concern according to the IUCN and ICMbio conservation status. However, some of the species with important commercial value in the region were classified in some level of threat or as near threat. These species must be observed more carefully, especially *E. itajara*, currently with forbidden catches and commercialization (PEREIRA 2018). Also, attention is needed to species classified as NT and DD because it leads to an inhibition of a full grasp of their extinction risk.

The ecological importance of estuaries is indisputable, however, there is still a shortage of knowledge on their fish species composition knowledge on a larger scale, what might lead to an underestimation of their biodiversity (Barletta et al. 2010; Rotundo et al. 2020). The existing and available material literature presented in this study indicated a weakness of scientific studies on estuarine fishes for some of the states of Northeast Brazil, along with insufficient studies aiming at the compilation of these types of data. Our results contribute not only to an effective integration of different information on fish data, but also for providing a wider view on fish diversity in the Northeast estuaries of Brazil. It is evident, however, that in some estuaries, such those in Pernambuco, more information are available and abiotic data are easily found within grey literature. The opposite is observed in the estuaries in the state of Rio Grande do Norte for example. Henceforth, further research is still needed in taxonomic diversity and abiotic characterization of estuaries for better understanding the distribution patterns of coastal fish species and applying such knowledge on appropriate conservation policies.

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Author contributions

RSL and ASL performed the statistical analysis; RSL, ASL and LNE prepared the figures and tables; and RSL, LNE, ALS, FLF, TF and WS wrote the manuscript.

Conflict of interest

The authors declare that they have no conflict of interest.

Ethical approval

All applicable international and national guidelines for the care and use of animals were followed by the authors.

Sampling and field studies

Permits for sampling field studies are not applicable.

Data availability

All data generated and related with this specific study are included in this article or supplementary information.

Table I: List of fish species recorded in 45 estuaries in north-eastern Brazil, along with their conservation status (ICMbio and IUCN list). Estuaries: São Luís Island, Caranguejos Island, Parnaíba River, Timonha River, Barro dos Remédios, Coreaú, Guriú, Acaraú and Torrões. ICMbio and IUCN classification: endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

Species	São Luís Island-MA	Caranguejos Island-MA	Parnaíba River-PI	Timonha River-CE	Barra dos remédios- CE	Coreaú- CE	Guriú-CE	Acaraú-CE	Torrões- CE	IUCN	ICMBIO
ORECTOLOBIFORMES											
Ginglymostomatidae											
<i>Ginglymostoma cirratum</i> (Bonnaterre, 1788)	x									VU	VU
CARCHARHINIFORMES											
Carcharhinidae											
<i>Carcharhinus leucas</i> (Muller & Henle, 1839)				x		x		x		NT	NT
<i>Carcharhinus limbatus</i> (Muller & Henle, 1839)										NT	NT
<i>Carcharhinus porosus</i> (Ranzani, 1839)	x									CR	CR
<i>Isogomphodon oxyrinchus</i> (Muller & Henle, 1839)	x									CR	CR
<i>Rhizoprionodon lalandii</i> (Muller & Henle, 1839)	x									VU	NT
<i>Rhizoprionodon porosus</i> (Poey, 1861)	x									VU	DD
Sphyrnidae											
<i>Sphyrna lewini</i> (Griffith & Smith, 1834)	x									CR	CR
<i>Sphyrna tiburo</i> (Linnaeus, 1758)	x									EN	CR
<i>Sphyrna tudes</i> (Valenciennes, 1822)	x									CR	CR
TORPEDINIFORMES											
Narcinidae											
<i>Narcine brasiliensis</i> (Olfers, 1831)	x									NT	DD
PRISTIFORMES											
Pristidae											
<i>Pristis pristis</i> Muller & Henle, 1841	x									CR	CR

	Rhinobatidae																						
	<i>Pseudobatos percellens</i> (Walbaum, 1792)													x						EN	DD		
MYLIOBATIFORMES																							
	Dasyatidae																						
	<i>Fontitrygon geijskesi</i> (Boeseman, 1948)	x																			CR	DD	
	<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)																				NT	DD	
	<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	x												x	x	x	x	x	x		NT	LC	
	<i>Hypanus marianae</i> (Gomes, Rosa & Gadig, 2000)																				EN	DD	
	<i>Hypanus say</i> (Lesueur, 1817)																				NT	DD	
	Gymnuridae																						
	<i>Gymnura micrura</i> (Bloch & Scheneider, 1801)	x												x							NT	NT	
	Myliobatidae																						
	<i>Aetobatus narinari</i> (Euphrasen, 1790)																					EN	DD
	<i>Mobula hypostoma</i> (Bancroft, 1831)																					EN	VU
	<i>Rhinoptera bonasus</i> (Mitchill, 1815)																					VU	DD
ELOPIFORMES																							
	Elopidae																						
	<i>Elops saurus</i> Linnaeus, 1766	x												x	x	x	x	x	x		LC	NE	
	Megalopidae																						
	<i>Megalops atlanticus</i> Valenciennes, 1847	x												x								VU	VU
ALBULIFORMES																							
	Albulidae																						
	<i>Albula vulpes</i> (Linnaeus, 1758)	x												x	x	x	x	x	x		NT	DD	
ANGUILLIFORMES																							
	Muraenidae																						
	<i>Gymnothorax funebris</i> Ranzani, 1839	x																				LC	DD
	<i>Gymnothorax moringa</i> (Cuvier, 1829)																					LC	DD
	<i>Gymnothorax ocellatus</i> Agassiz, 1831																					LC	DD

										LC	DD		
	<i>Gymnothorax vicinus</i> (Castelnau, 1855)												
	Ophichthidae												
	<i>Ahlia egmontis</i> (Jordan, 1884)									LC	LC		
	<i>Myrichthys breviceps</i> (Richardson, 1848)									LC	LC		
	<i>Myrichthys ocellatus</i> (Lesueur, 1825)					x				LC	LC		
	<i>Myrophis punctatus</i> Lütken, 1852									LC	LC		
	<i>Ophichthus cylindroideus</i> (Ranzani, 1839)								x	LC	LC		
	<i>Ophichthus ophis</i> (Linnaeus, 1758)									LC	LC		
	Muraenesocidae												
	<i>Cynoponticus savanna</i> (Bancroft, 1831)									LC	LC		
	Congridae												
	<i>Ariosoma balearicum</i> (Delaroche, 1809)									LC	LC		
CLUPEIFORMES													
	Pristigasteridae												
	<i>Chirocentrodon bleekermanus</i> (Poey, 1867)					x	x	x	x	x	x	LC	LC
	<i>Odontognathus mucronatus</i> Lacepède, 1800											LC	LC
	<i>Pellona flavipinnis</i> (Valenciennes, 1837)										x	LC	LC
	<i>Pellona harroweri</i> (Fowler, 1917)											LC	LC
	Engraulidae												
	<i>Anchoa filifera</i> (Fowler, 1915)											LC	LC
	<i>Anchoa hepsetus</i> (Linnaeus, 1758)					x	x	x	x	x	x	LC	NE
	<i>Anchoa januaria</i> (Steindachner, 1879)											LC	LC
	<i>Anchoa lyolepis</i> (Evermann & Marsh, 1900)											LC	LC
	<i>Anchoa parva</i> (Meek & Hildebrand, 1923)											LC	NE
	<i>Anchoa spinifer</i> (Valenciennes, 1848)					x	x	x	x	x	x	LC	LC
	<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)											LC	LC
	<i>Anchovia clupeioides</i> (Swainson, 1839)					x	x	x	x	x	x	LC	LC
	<i>Anchoviella lepidentostole</i> (Fowler, 1911)											LC	LC

<i>Cetengraulis edentulus</i> (Cuvier, 1829)	x		x							LC	LC
<i>Engraulis anchoita</i> Hubbs & Marini 1935										LC	LC
<i>Lycengraulis batesii</i> (Gunther, 1868)				x	x	x	x	x	x	LC	LC
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	x				x	x	x	x	x	LC	LC
<i>Pterengraulis atherinoides</i> (Linnaeus, 1766)	x			x						LC	LC
Clupeidae											
<i>Harengula clupeola</i> (Cuvier, 1829)										LC	LC
<i>Harengula jaguana</i> Poey, 1865										LC	LC
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)	x									LC	LC
<i>Opisthonema oglinum</i> (Lesueur, 1818)	x			x	x	x	x	x	x	LC	LC
<i>Platanichthys platana</i> (Regan, 1917)										LC	LC
<i>Rhinosardinia amazonica</i> (Steindachner, 1879)	x									LC	LC
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)										LC	LC
CHARACIFORMES											
Erythrinidae											
<i>Hoplias malabaricus</i> (Bloch, 1794)				x	x		x		x	LC	LC
<i>Prochilodus argenteus</i> Spix & Agassiz, 1829										NE	LC
Serrasalminidae											
<i>Pygocentrus nattereri</i> Kner, 1858	x									NE	LC
<i>Serrasalmus rhombeus</i> (Linnaeus, 1766)				x						NE	LC
<i>Serrasalmus spilopleura</i> Kner, 1858										NE	LC
Anostomidae											
<i>Schizodon fasciatus</i> Spix & Agassiz, 1829										NE	LC
<i>Leporinus friderici</i> (Bloch, 1794)										NE	LC
Curimatidae											
<i>Steindachnerina elegans</i> (Steindachner, 1875)										NE	LC
Characidae											
<i>Astyanax bimaculatus</i> (Linnaeus, 1758)										NE	LC

SILURIFORMES										NE	LC
	<i>Hemigrammus marginatus</i> Ellis, 1911										
	Loricariidae										
	<i>Hypostomus plecostomus</i> (Linnaeus, 1758)									NE	NE
	<i>Loricaria parnahybae</i> Steindachner, 1907									NE	LC
	Ariidae										
	<i>Aspistor luniscutis</i> (Valenciennes, 1840)				x					NE	LC
	<i>Aspistor quadriscutis</i> (Valenciennes, 1840)	x								LC	LC
	<i>Bagre bagre</i> (Linnaeus, 1766)	x	x		x					LC	NT
	<i>Bagre marinus</i> (Mitchill, 1815)									LC	DD
	<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)			x						NE	LC
	<i>Cathorops spixii</i> (Agassiz, 1829)	x	x		x	x		x		NE	LC
	<i>Genidens genidens</i> (Cuvier, 1829)									LC	LC
	<i>Notarius grandicassis</i> (Valenciennes, 1840)	x								LC	LC
	<i>Sciades couma</i> (Valenciennes, 1840)	x								LC	DD
	<i>Sciades herzbergii</i> (Bloch, 1794)	x	x		x				x	LC	LC
	<i>Sciades parkeri</i> (Traill, 1832)	x								VU	VU
	<i>Sciades proops</i> (Valenciennes, 1840)	x	x							NE	DD
	Aspredinidae										
	<i>Aspredo aspredo</i> (Linnaeus, 1758)	x			x					LC	LC
	<i>Aspredinichthys tибicen</i> (Valenciennes, 1840)	x	x							LC	LC
	Auchenipteridae										
	<i>Ageneiosus inermis</i> (Linnaeus, 1766)				x					NE	LC
	<i>Pseudauchenipterus nodosus</i> (Bloch, 1794)	x	x		x					NE	LC
	<i>Trachelyopterus galeatus</i> (Linnaeus, 1766)									NE	LC
	Heptapteridae										
	<i>Pimelodella cristata</i> (Muller & Troschel, 1849)									LC	LC
	<i>Pimelodella lateristriga</i> (Lichtenstein, 1823)									NE	DD

	Pimelodidae																	
	<i>Brachyplatystoma vaillantii</i> (Valenciennes, 1840)										x	NE	LC					
	<i>Pseudoplatystoma fasciatum</i> (Linnaeus, 1766)										x	NE	NE					
	<i>Sorubim lima</i> (Bloch & Schneider, 1801)										x	NE	LC					
AULOPIFORMES																		
	Synodontidae																	
	<i>Synodus foetens</i> (Linnaeus, 1766)										x	x	x	x	LC	LC		
	<i>Synodus poeyi</i> Jordan, 1887														LC	LC		
	<i>Trachinocephalus myops</i> (Forster, 1801)														LC	LC		
HOLOCENTRIFORMES																		
	Holocentridae																	
	<i>Holocentrus adscensionis</i> (Osbeck, 1765)														LC	LC		
BATRACHOIDIFORMES																		
	Batrachoididae																	
	<i>Amphichthys cryptocentrus</i> (Valenciennes, 1837)														LC	LC		
	<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)										x	x	x	x	x	x	LC	LC
	<i>Thalassophryne nattereri</i> Steindachner, 1876										x		x	x	x	x	LC	LC
	<i>Thalassophryne punctata</i> Steindachner, 1876																NE	LC
KURTIFORMES																		
	Apogonidae																	
	<i>Phaeoptyx pigmentaria</i> (Poey, 1860)														LC	LC		
GOBIIFORMES																		
	Eleotridae																	
	<i>Dormitator maculatus</i> (Bloch, 1792)											x	x	x	x	x	LC	NT
	<i>Eleotris pisonis</i> (Gmelin, 1789)																LC	LC
	<i>Erotelis smaragdus</i> (Valenciennes, 1837)																LC	LC
	<i>Guavina guavina</i> (Valenciennes, 1837)										x						LC	LC
	Gobiidae																	

	<i>Mugil liza</i> Valenciennes, 1836	x		x	x	x	x	x	x	DD	NT
	<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007									LC	DD
CICHLIFORMES											
	Cichlidae										
	<i>Cichla ocellaris</i> Bloch & Schneider, 1801		x			x				NE	LC
	<i>Oreochromis niloticus</i> (Linnaeus, 1758)		x			x			x	LC	NE
BLENNIIFORMES											
	Dactyloscopidae										
	<i>Dactyloscopus crossotus</i> Starks, 1913									LC	LC
	Blenniidae										
	<i>Scartella cristata</i> (Linnaeus, 1758)									LC	LC
	Labrisomidae										
	<i>Labrisomus nuchipinnis</i> (Quoy & Gaimard, 1824)									LC	LC
ATHERINIFORMES											
	Atherinidae										
	<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)	x		x	x	x	x	x	x	LC	LC
	<i>Atherinella cf. blackburni</i> (Schultz, 1949)									LC	LC
	<i>Odontesthes bonariensis</i> (Valenciennes, 1835)									NE	DD
BELONIFORMES											
	Exocoetidae										
	<i>Cheilopogon melanurus</i> (Valenciennes, 1847)	x								LC	LC
	<i>Hirundichthys affinis</i> (Gunther, 1866)									LC	LC
	Hemiramphidae										
	<i>Hemiramphus balao</i> Lesueur, 1821									LC	DD
	<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)									LC	LC
	<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)									LC	LC
	<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)								x	LC	NT
	Belonidae										

<i>Ablennes hians</i> (Valenciennes, 1846)									LC	LC	
<i>Strongylura marina</i> (Walbaum, 1792)	x	x		x	x		x		x	LC	LC
<i>Strongylura timucu</i> (Walbaum, 1792)	x				x	x			x	LC	LC
<i>Tylosurus acus acus</i> (Lacepède, 1803)										LC	LC
<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)										LC	LC
CYPRINODONTIFORMES											
Poeciliidae											
<i>Poecilia vivipara</i> Bloch & Schneider, 1801										NE	LC
CARANGIFORMES											
Rachycentridae											
<i>Rachycentron canadum</i> (Linnaeus, 1766)										LC	LC
Echeneidae											
<i>Echeneis naucrates</i> Linnaeus, 1758	x			x		x			x	LC	LC
<i>Phtheichthys lineatus</i> (Menziés, 1791)										LC	LC
<i>Remora remora</i> (Linnaeus, 1758)										LC	LC
Carangidae											
<i>Carangoides bartholomaei</i> (Cuvier, 1833)										LC	LC
<i>Caranx crysos</i> (Mitchill, 1815)	x					x			x	LC	LC
<i>Caranx hippos</i> (Linnaeus, 1766)	x		x	x	x	x	x	x	x	LC	LC
<i>Caranx latus</i> Agassiz, 1831	x		x	x	x	x	x	x	x	LC	LC
<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	x		x	x	x	x	x	x	x	LC	LC
<i>Hemicaranx amblyrhynchus</i> (Cuvier, 1833)	x									LC	LC
<i>Oligoplites palometa</i> (Cuvier, 1832)	x		x	x	x	x	x	x	x	LC	LC
<i>Oligoplites saliens</i> (Bloch, 1793)			x	x	x	x	x	x	x	LC	LC
<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	x	x		x	x	x	x	x	x	LC	LC
<i>Selene brownii</i> (Curvier, 1816)										LC	LC
<i>Selene setapinnis</i> (Mitchill, 1815)				x		x				LC	LC
<i>Selene vomer</i> (Linnaeus, 1758)	x		x	x	x	x			x	LC	LC

	<i>Trachinotus carolinus</i> (Linnaeus, 1766)	x								LC	LC
	<i>Trachinotus cayennensis</i> Cuvier, 1832	x								LC	DD
	<i>Trachinotus falcatus</i> (Linnaeus, 1758)	x		x	x	x	x	x	x	LC	LC
	<i>Trachinotus goodei</i> Jordan & Evermann, 1896		x	x	x	x	x	x	x	LC	LC
ISTIOPHORIFORMES											
	Sphyraenidae										
	<i>Sphyraena barracuda</i> (Edwards, 1771)			x			x			LC	LC
	<i>Sphyraena guachancho</i> Cuvier, 1829									LC	LC
PLEURONECTIFORMES											
	Paralichthyidae										
	<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900									LC	LC
	<i>Citharichthys cornutus</i> (Günther, 1880)									LC	LC
	<i>Citharichthys macrops</i> Dresel, 1885									LC	LC
	<i>Citharichthys spilopterus</i> Günther, 1862	x		x	x			x	x	LC	LC
	<i>Etropus crossotus</i> Jordan & Gilbert, 1882	x			x			x	x	LC	LC
	<i>Etropus longimanus</i> Norman, 1933									NE	LC
	<i>Paralichthys brasiliensis</i> (Ranzani, 1842)		x							NE	LC
	<i>Paralichthys orbignyanus</i> (Valenciennes, 1839)									DD	DD
	<i>Paralichthys tropicus</i> Ginsburg, 1933									DD	NE
	<i>Syacium micrurum</i> Ranzani, 1842									LC	LC
	<i>Syacium papillosum</i> (Linnaeus, 1758)									LC	LC
	Bothidae										
	<i>Bothus lunatus</i> (Linnaeus, 1758)									LC	LC
	<i>Bothus ocellatus</i> (Agassiz, 1831)									LC	LC
	<i>Bothus robinsi</i> Topp & Hoff, 1972									LC	LC
	Achiridae										
	<i>Achirus achirus</i> (Linnaeus, 1758)	x	x	x	x				x	LC	LC
	<i>Achirus declivis</i> Chabanaud, 1940			x	x	x	x			LC	LC

	<i>Achirus lineatus</i> (Linnaeus, 1758)		x		x		x		x	LC	LC	
	<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009									NE	VU	
	<i>Trinectes microphthalmus</i> (Chabanaud, 1928)									LC	LC	
	<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	x				x		x		LC	LC	
	Cynoglossidae											
	<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)									LC	LC	
	<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)	x								LC	LC	
	<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)									LC	LC	
SYNGNATHIFORMES												
	Syngnathidae											
	<i>Bryx dunckeri</i> (Metzelaar, 1919)									LC	LC	
	<i>Cosmocampus elucens</i> (Poey, 1868)									LC	LC	
	<i>Hippocampus erectus</i> Perry, 1810									VU	VU	
	<i>Hippocampus reidi</i> Ginsburg, 1933		x			x		x		x	NT	VU
	<i>Microphis brachyurus lineatus</i> (Kaup, 1856)									NE	NE	
	<i>Microphis brachyurus brachyurus</i> (Bleeker, 1854)									LC	LC	
	Fistulariidae											
	<i>Fistularia petimba</i> Lacepède, 1803									LC	LC	
	<i>Fistularia tabacaria</i> Linnaeus, 1758									LC	LC	
	<i>Syngnathus pelagicus</i> Linnaeus, 1758									LC	LC	
	Dactylopteridae											
	<i>Dactylopterus volitans</i> (Linnaeus, 1758)									LC	LC	
SCOMBRIFORMES												
	Trichiuridae											
	<i>Trichiurus lepturus</i> Linnaeus, 1758	x				x		x		x	LC	LC
	Scombridae											
	<i>Euthynnus alletteratus</i> (Rafinesque, 1810)									LC	LC	

	<i>Scomberomorus brasiliensis</i> Collette, Russo & Zavala-Camin, 1978									x		LC	LC	
	<i>Scomberomorus cavalla</i> (Cuvier, 1829)											LC	LC	
	<i>Scomberomorus maculatus</i> (Mitchill, 1815)											LC	NE	
	<i>Scomberomorus regalis</i> (Bloch, 1793)											LC	LC	
	Stromateidae													
	<i>Peprilus paru</i> (Linnaeus, 1758)	x				x				x		LC	LC	
TRACHINIFORMES														
	Uranoscopidae													
	<i>Astroscopus y-graecum</i> Cuvier, 1829											LC	LC	
LABRIFORMES														
	Labridae													
	<i>Halichoeres poeyi</i> (Steindachner, 1867)											LC	LC	
	Scaridae													
	<i>Nicholsina usta</i> (Valenciennes, 1840)											LC	LC	
	<i>Sparisoma amplum</i> (Ranzani, 1842)											LC	NT	
	<i>Sparisoma axillare</i> (Steindachner, 1878)											DD	VU	
	<i>Sparisoma radians</i> (Valenciennes, 1840)											LC	LC	
PERCIFORMES														
	Centropomidae													
	<i>Centropomus ensiferus</i> Poey, 1860											x	LC	LC
	<i>Centropomus mexicanus</i> Bocourt, 1868												LC	LC
	<i>Centropomus parallelus</i> Poey, 1860	x	x	x	x	x	x	x	x	x	x	LC	LC	
	<i>Centropomus pectinatus</i> Poey, 1860											LC	LC	
	<i>Centropomus undecimalis</i> (Bloch, 1792)	x		x	x	x	x	x	x	x	x	LC	LC	
	Gerreidae													
	<i>Diapterus auratus</i> Ranzani, 1842	x		x	x	x	x	x	x	x	x	LC	LC	
	<i>Diapterus rhombeus</i> (Cuvier, 1829)	x	x	x	x		x		x			LC	LC	

<i>Eucinostomus argenteus</i> Baird & Girard, 1855	x				x					LC	LC
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	x									LC	LC
<i>Eucinostomus havana</i> (Nichols, 1912)										LC	LC
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	x		x		x		x		x	LC	LC
<i>Eugerres brasilianus</i> (Cuvier, 1830)	x		x						x	LC	LC
<i>Gerres cinereus</i> (Walbaum, 1792)			x		x		x		x	LC	LC
<i>Ulaema lefroyi</i> (Goode, 1874)										LC	LC
Mullidae											
<i>Pseudupeneus maculatus</i> (Bloch, 1793)										LC	LC
Serranidae											
<i>Alphestes afer</i> (Bloch, 1793)										LC	DD
<i>Diplectrum radiale</i> (Quoy & Gaimard, 1824)	x		x		x					LC	LC
<i>Epinephelus adscensionis</i> (Osbeck, 1765)										LC	DD
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	x		x				x			VU	CR
<i>Hyporthodus nigritus</i> (Holbrook, 1855)										NT	EN
<i>Mycteroperca bonaci</i> (Poey, 1860)	x		x		x		x		x	NT	VU
<i>Mycteroperca tigris</i> (Valenciennes, 1833)										DD	DD
<i>Mycteroperca venenosa</i> (Linnaeus, 1758)										NT	DD
<i>Rypticus randalli</i> Courtenay, 1967	x		x		x		x		x	LC	LC
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)										LC	LC
<i>Serranus flaviventris</i> (Cuvier, 1829)										LC	LC
<i>Serranus phoebe</i> Poey, 1851										LC	LC
Pomacanthidae											
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)										LC	DD
<i>Pomacanthus paru</i> (Bloch, 1787)	x									LC	DD
Chaetodontidae											
<i>Chaetodon ocellatus</i> (Bloch, 1787)										LC	DD
<i>Chaetodon striatus</i> Linnaeus, 1758										LC	LC

Haemulidae											
<i>Anisotremus surinamensis</i> (Bloch, 1791)								x		DD	DD
<i>Anisotremus virginicus</i> (Linnaeus, 1758)								x	x	LC	LC
<i>Conodon nobilis</i> (Linnaeus, 1758)	x			x	x				x	LC	LC
<i>Genyatremus luteus</i> (Bloch, 1790)	x	x		x	x			x	x	NE	LC
<i>Haemulon aurolineatum</i> Cuvier, 1830										LC	LC
<i>Haemulon parra</i> (Desmarest, 1823)									x	LC	LC
<i>Haemulon plumierii</i> (Lacepède, 1801)										LC	DD
<i>Haemulon squamipinna</i> Rocha & Rosa, 1999										NE	LC
<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)										LC	LC
<i>Orthopristis ruber</i> (Cuvier, 1830)	x									LC	LC
<i>Pomadasys corvinaeformis</i> (Steindachner, 1868)	x			x	x			x	x	LC	LC
<i>Pomadasys crocro</i> (Cuvier, 1830)										DD	LC
<i>Pomadasys ramosus</i> Poey, 1860										NE	LC
Lutjanidae											
<i>Lutjanus alexandrei</i> Moura & Linderman, 2007										NE	LC
<i>Lutjanus analis</i> (Cuvier, 1828)								x	x	NT	NT
<i>Lutjanus apodus</i> (Walbaum, 1792)								x	x	LC	NE
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)										VU	VU
<i>Lutjanus griseus</i> (Linnaeus, 1758)									x	LC	NE
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	x			x	x			x	x	DD	NT
<i>Lutjanus synagris</i> (Linnaeus, 1758)	x			x	x			x	x	NT	NT
<i>Ocyurus chrysurus</i> (Bloch, 1791)										DD	NT
Polynemidae											
<i>Polydactylus oligodon</i> (Gunther, 1860)	x									LC	LC
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	x			x						LC	LC
SCORPAENIFORMES											
Scorpaenidae											

	<i>Scorpaena isthmensis</i> Meek & Hilderbrand, 1928									LC	LC
	<i>Scorpaena plumieri</i> Bloch, 1789	x								LC	LC
	Triglidae										
	<i>Prionotus punctatus</i> (Bloch, 1793)									LC	LC
MORONIFORMES											
	Ephippidae										
	<i>Chaetodipterus faber</i> (Broussonet, 1782)	x	x	x	x	x	x	x	x	LC	LC
ACANTHURIFORMES											
	Sciaenidae										
	<i>Bairdiella ronchus</i> (Cuvier, 1830)	x	x	x	x	x	x	x	x	LC	LC
	<i>Cynoscion acoupa</i> (Lacepède, 1801)	x	x	x	x	x	x	x	x	LC	NT
	<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1833)									LC	LC
	<i>Cynoscion leiarchus</i> (Cuvier, 1830)	x	x	x	x	x	x	x	x	LC	LC
	<i>Cynoscion microlepidotus</i> (Cuvier, 1830)	x		x	x	x	x	x	x	LC	LC
	<i>Cynoscion steindachmeri</i> (Jordan, 1889)	x	x							LC	LC
	<i>Cynoscion virescens</i> (Cuvier, 1830)			x				x		LC	LC
	<i>Isopisthus parvipinnis</i> (Cuvier, 1830)	x		x						LC	LC
	<i>Larimus breviceps</i> Cuvier, 1830									LC	LC
	<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)	x	x	x						LC	LC
	<i>Menticirrhus americanus</i> (Linnaeus, 1758)	x		x	x	x	x	x	x	LC	DD
	<i>Menticirrhus littoralis</i> (Holbrook, 1847)			x	x	x	x	x	x	LC	DD
	<i>Micropogonias furnieri</i> (Desmarest, 1823)	x		x	x	x	x	x	x	LC	LC
	<i>Nebris microps</i> Cuvier, 1830	x		x						LC	LC
	<i>Odontoscion dentex</i> (Cuvier, 1830)									LC	LC
	<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925	x				x		x		LC	DD
	<i>Paralonchurus brasiliensis</i> (Steindachner, 1875)									LC	LC
	<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)									LC	DD
	<i>Pogonias cromis</i> (Linnaeus, 1766)									LC	EN

	<i>Stellifer brasiliensis</i> (Schultz, 1945)	x								NE	LC	
	<i>Stellifer microps</i> (Steindachner, 1864)									LC	LC	
	<i>Stellifer naso</i> (Jordan, 1889)	x	x							LC	LC	
	<i>Stellifer rastrifer</i> (Jordan, 1889)	x	x	x						LC	LC	
	<i>Stellifer stellifer</i> (Bloch, 1790)	x		x						DD	LC	
	<i>Umbrina coroides</i> Cuvier, 1830									LC	LC	
	Acanthuridae											
	<i>Acanthurus bahianus</i> Castelnau, 1855			x		x			x		LC	LC
	<i>Acanthurus chirurgus</i> (Bloch, 1787)									LC	LC	
	<i>Acanthurus coeruleus</i> Bloch & Schneider, 1801			x		x			x	LC	LC	
SPARIFORMES												
	Lobotidae											
	<i>Lobotes surinamensis</i> (Bloch, 1790)	x		x	x	x	x	x	x	LC	LC	
	Sparidae											
	<i>Archosargus probatocephalus</i> (Walbaum, 1792)	x		x		x			x	LC	DD	
	<i>Archosargus rhomboidalis</i> (Linnaeus, 1758)			x		x				LC	LC	
LOPHIIFORMES												
	Antennariidae											
	<i>Antennarius multiocellatus</i> (Valenciennes, 1837)									LC	DD	
	<i>Antennarius striatus</i> (Shaw, 1794)			x	x					LC	DD	
	Ogcocephalidae											
	<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	x		x	x	x	x	x	x	NE	LC	
TETRAODONTIFORMES												
	Ostraciidae											
	<i>Acanthostracion quadricornis</i> (Linnaeus, 1758)									LC	LC	
	<i>Lactophrys trigonus</i> (Linnaeus, 1758)									LC	LC	
	<i>Lactophrys triqueter</i> (Linnaeus, 1758)									LC	NE	
	Balistidae											

<i>Balistes vetula</i> Linnaeus, 1758										NT	NT
<i>Melichthys niger</i> (Bloch, 1786)										LC	LC
Monacanthidae											
<i>Aluterus heudelotii</i> Hollard, 1855										LC	LC
<i>Aluterus monoceros</i> (Linnaeus, 1758)	x									LC	NT
<i>Aluterus schoepfii</i> (Walbaum, 1792)										LC	LC
<i>Cantherhines pullus</i> (Ranzani, 1842)										LC	LC
<i>Monacanthus ciliatus</i> (Mitchill, 1818)										LC	LC
<i>Stephanolepis hispidus</i> (Linnaeus, 1766)										LC	LC
<i>Stephanolepis setifer</i> (Bennett, 1831)										LC	LC
Tetraodontidae											
<i>Canthigaster figueiredoi</i> Moura & Castro, 2002										LC	LC
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	x	x	x	x	x	x	x	x	x	LC	LC
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	x			x	x				x	LC	LC
<i>Sphoeroides greeleyi</i> Gilbert, 1900	x									LC	LC
<i>Sphoeroides nephelus</i> (Goode & Bean, 1882)										LC	NE
<i>Sphoeroides spengleri</i> (Bloch, 1785)										LC	LC
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	x		x	x	x	x	x	x	x	LC	DD
<i>Sphoeroides tyleri</i> Shipp, 1972										LC	LC
Diodontidae											
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	x									LC	LC
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)				x				x		LC	LC
<i>Diodon holocanthus</i> Linnaeus, 1758										LC	LC
<i>Diodon hystrix</i> Linnaeus, 1758										LC	LC

Table II: List of fish species recorded in 45 estuaries in north-eastern Brazil, along with their conservation status (ICMbio and IUCN list). Estuaries: Aracatiaçu, Mundaú River- CE, Curu River, Cocó River, Pacoti River, Pirangi River, Jaguaribe River – CE, Grossos and Areia Branca. ICMbio and IUCN classification: endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

Species	Aracatiaçu- CE	Mundaú River-CE	Curu River-CE	Cocó River-CE	Pacoti River-CE	Pirangi River-CE	Jaguaribe River-CE	Grossos- RN	Areia Branca-RN	IUCN	ICMbio
ORECTOLOBIFORMES											
Ginglymostomatidae											
<i>Ginglymostoma cirratum</i> (Bonnaterre, 1788)										VU	VU
CARCHARHINIFORMES											
Carcharhinidae											
<i>Carcharhinus leucas</i> (Muller & Henle, 1839)				x			x			NT	NT
<i>Carcharhinus limbatus</i> (Muller & Henle, 1839)			x							NT	NT
<i>Carcharhinus porosus</i> (Ranzani, 1839)										CR	CR
<i>Isogomphodon oxyrinchus</i> (Muller & Henle, 1839)										CR	CR
<i>Rhizoprionodon lalandii</i> (Muller & Henle, 1839)										VU	NT
<i>Rhizoprionodon porosus</i> (Poey, 1861)			x							VU	DD
Sphyrnidae											
<i>Sphyrna lewini</i> (Griffith & Smith, 1834)										CR	CR
<i>Sphyrna tiburo</i> (Linnaeus, 1758)							x			EN	CR
<i>Sphyrna tudes</i> (Valenciennes, 1822)										CR	CR
TORPEDINIFORMES											
Narcinidae											
<i>Narcine brasiliensis</i> (Olfers, 1831)									x	NT	DD
PRISTIFORMES											
Pristidae											
<i>Pristis pristis</i> Muller & Henle, 1841										CR	CR
Rhinobatidae											

											EN	DD
MYLIOBATIFORMES	<i>Pseudobatos percellens</i> (Walbaum, 1792)											
	Dasyatidae											
	<i>Fontitrygon geijskesi</i> (Boeseman, 1948)										CR	DD
	<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)										NT	DD
	<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	x	x	x	x	x	x	x			NT	LC
	<i>Hypanus marianae</i> (Gomes, Rosa & Gadig, 2000)			x							EN	DD
	<i>Hypanus say</i> (Lesueur, 1817)								x		NT	DD
	Gymnuridae											
	<i>Gymnura micrura</i> (Bloch & Scheneider, 1801)		x	x					x		NT	NT
	Myliobatidae											
	<i>Aetobatus narinari</i> (Euphrasen, 1790)			x							EN	DD
	<i>Mobula hypostoma</i> (Bancroft, 1831)										EN	VU
	<i>Rhinoptera bonasus</i> (Mitchill, 1815)										VU	DD
ELOPIFORMES												
	Elopidae											
	<i>Elops saurus</i> Linnaeus, 1766	x	x	x	x	x	x	x	x	x	LC	NE
	Megalopidae											
	<i>Megalops atlanticus</i> Valenciennes, 1847		x	x	x	x	x	x		x	VU	VU
ALBULIFORMES												
	Albulidae											
	<i>Albula vulpes</i> (Linnaeus, 1758)	x	x	x	x	x	x	x			NT	DD
ANGUILLIFORMES												
	Muraenidae											
	<i>Gymnothorax funebris</i> Ranzani, 1839										LC	DD
	<i>Gymnothorax moringa</i> (Cuvier, 1829)										LC	DD
	<i>Gymnothorax ocellatus</i> Agassiz, 1831										LC	DD
	<i>Gymnothorax vicinus</i> (Castelnau, 1855)										LC	DD

Ophichthidae												
											LC	LC
											LC	LC
			x	x						x	LC	LC
						x	x			x	LC	LC
											LC	LC
											LC	LC
Muraenesocidae												
											LC	LC
Congridae												
											LC	LC
CLUPEIFORMES												
Pristigasteridae												
		x	x	x	x	x	x	x		x	LC	LC
										x	LC	LC
											LC	LC
										x	LC	LC
Engraulidae												
										x	LC	LC
		x	x	x	x	x	x	x			LC	NE
											LC	LC
											LC	LC
											LC	NE
		x	x	x	x	x	x	x			LC	LC
					x						LC	LC
		x	x	x	x	x	x	x			LC	LC
											LC	LC
					x						LC	LC

	<i>Engraulis anchoita</i> Hubbs & Marini 1935									LC	LC
	<i>Lycengraulis batesii</i> (Gunther, 1868)	x	x	x	x	x	x	x		LC	LC
	<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	x	x	x	x	x	x	x	x	LC	LC
	<i>Pterengraulis atherinoides</i> (Linnaeus, 1766)									LC	LC
	Clupeidae										
	<i>Harengula clupeola</i> (Cuvier, 1829)			x						LC	LC
	<i>Harengula jaguana</i> Poey, 1865									LC	LC
	<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)								x	LC	LC
	<i>Opisthonema oglinum</i> (Lesueur, 1818)	x	x	x	x	x	x	x	x	LC	LC
	<i>Platanichthys platana</i> (Regan, 1917)								x	LC	LC
	<i>Rhinosardinia amazonica</i> (Steindachner, 1879)				x	x		x		LC	LC
	<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)				x	x		x	x	LC	LC
CHARACIFORMES											
	Erythrinidae										
	<i>Hoplias malabaricus</i> (Bloch, 1794)			x	x	x	x	x		LC	LC
	<i>Prochilodus argenteus</i> Spix & Agassiz, 1829									NE	LC
	Serrasalminidae										
	<i>Pygocentrus nattereri</i> Kner, 1858							x		NE	LC
	<i>Serrasalmus rhombeus</i> (Linnaeus, 1766)							x		NE	LC
	<i>Serrasalmus spilopleura</i> Kner, 1858							x		NE	LC
	Anostomidae										
	<i>Schizodon fasciatus</i> Spix & Agassiz, 1829					x		x		NE	LC
	<i>Leporinus friderici</i> (Bloch, 1794)							x		NE	LC
	Curimatidae										
	<i>Steindachnerina elegans</i> (Steindachner, 1875)							x		NE	LC
	Characidae										
	<i>Astyanax bimaculatus</i> (Linnaeus, 1758)							x		NE	LC
	<i>Hemigrammus marginatus</i> Ellis, 1911							x		NE	LC

SILURIFORMES

Loricariidae										
<i>Hypostomus plecostomus</i> (Linnaeus, 1758)								x	NE	NE
<i>Loricaria parmahybae</i> Steindachner, 1907								x	NE	LC
Ariidae										
<i>Aspistor luniscutis</i> (Valenciennes, 1840)									NE	LC
<i>Aspistor quadriscutis</i> (Valenciennes, 1840)									LC	LC
<i>Bagre bagre</i> (Linnaeus, 1766)									LC	NT
<i>Bagre marinus</i> (Mitchill, 1815)								x	x	x
<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)									NE	LC
<i>Cathorops spixii</i> (Agassiz, 1829)	x	x	x					x	x	x
<i>Genidens genidens</i> (Cuvier, 1829)									LC	LC
<i>Notarius grandicassis</i> (Valenciennes, 1840)									LC	LC
<i>Sciades couma</i> (Valenciennes, 1840)									LC	DD
<i>Sciades herzbergii</i> (Bloch, 1794)	x	x	x	x				x	LC	LC
<i>Sciades parkeri</i> (Traill, 1832)									VU	VU
<i>Sciades proops</i> (Valenciennes, 1840)								x	NE	DD
Aspredinidae										
<i>Aspredo aspredo</i> (Linnaeus, 1758)									LC	LC
<i>Aspredinichthys tibicen</i> (Valenciennes, 1840)									LC	LC
Auchenipteridae										
<i>Ageneiosus inermis</i> (Linnaeus, 1766)									NE	LC
<i>Pseudauchenipterus nodosus</i> (Bloch, 1794)									NE	LC
<i>Trachelyopterus galeatus</i> (Linnaeus, 1766)				x				x	NE	LC
Heptapteridae										
<i>Pimelodella cristata</i> (Muller & Troschel, 1849)								x	LC	LC
<i>Pimelodella lateristriga</i> (Lichtenstein, 1823)								x	NE	DD
Pimelodidae										

	<i>Brachyplatystoma vaillantii</i> (Valenciennes, 1840)									NE	LC
	<i>Pseudoplatystoma fasciatum</i> (Linnaeus, 1766)									NE	NE
	<i>Sorubim lima</i> (Bloch & Schneider, 1801)									NE	LC
AULOPIFORMES											
	Synodontidae										
	<i>Synodus foetens</i> (Linnaeus, 1766)	x		x	x	x	x	x		LC	LC
	<i>Synodus poeyi</i> Jordan, 1887									LC	LC
	<i>Trachinocephalus myops</i> (Forster, 1801)									LC	LC
HOLOCENTRIFORMES											
	Holocentridae										
	<i>Holocentrus adscensionis</i> (Osbeck, 1765)									LC	LC
BATRACHOIDIFORMES											
	Batrachoididae										
	<i>Amphichthys cryptocentrus</i> (Valenciennes, 1837)								x	LC	LC
	<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)	x	x	x	x	x	x	x		LC	LC
	<i>Thalassophryne nattereri</i> Steindachner, 1876	x	x	x	x	x	x	x	x	LC	LC
	<i>Thalassophryne punctata</i> Steindachner, 1876									NE	LC
KURTIFORMES											
	Apogonidae										
	<i>Phaeoptyx pigmentaria</i> (Poey, 1860)									LC	LC
GOBIIFORMES											
	Eleotridae										
	<i>Dormitator maculatus</i> (Bloch, 1792)	x	x	x	x	x	x	x	x	LC	NT
	<i>Eleotris pisonis</i> (Gmelin, 1789)								x	LC	LC
	<i>Erotelis smaragdus</i> (Valenciennes, 1837)									LC	LC
	<i>Guavina guavina</i> (Valenciennes, 1837)								x	LC	LC
	Gobiidae										
	<i>Awaous tajasica</i> (Lichtenstein, 1822)									LC	LC

	<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007										LC	DD		
CICHLIFORMES														
	Cichlidae													
	<i>Cichla ocellaris</i> Bloch & Schneider, 1801										x	NE LC		
	<i>Oreochromis niloticus</i> (Linnaeus, 1758)			x		x		x		x		LC NE		
BLENNIIFORMES														
	Dactyloscopidae													
	<i>Dactyloscopus crossotus</i> Starks, 1913											LC LC		
	Blenniidae													
	<i>Scartella cristata</i> (Linnaeus, 1758)											LC LC		
	Labrisomidae													
	<i>Labrisomus nuchipinnis</i> (Quoy & Gaimard, 1824)											LC LC		
ATHERINIFORMES														
	Atherinidae													
	<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)	x		x		x		x		x		x	LC LC	
	<i>Atherinella cf. blackburni</i> (Schultz, 1949)												LC LC	
	<i>Odontesthes bonariensis</i> (Valenciennes, 1835)												NE DD	
BELONIFORMES														
	Exocoetidae													
	<i>Cheilopogon melanurus</i> (Valenciennes, 1847)												LC LC	
	<i>Hirundichthys affinis</i> (Gunther, 1866)												LC LC	
	Hemiramphidae													
	<i>Hemiramphus balao</i> Lesueur, 1821									x		x	LC DD	
	<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)												LC LC	
	<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)												LC LC	
	<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)			x		x						x	LC NT	
	Belonidae													
	<i>Ablennes hians</i> (Valenciennes, 1846)											x	x	LC LC

	<i>Strongylura marina</i> (Walbaum, 1792)	x		x	x	x	x	x			LC	LC
	<i>Strongylura timucu</i> (Walbaum, 1792)	x	x		x	x		x	x	x	LC	LC
	<i>Tylosurus acus acus</i> (Lacepède, 1803)										LC	LC
	<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)							x			LC	LC
CYPRINODONTIFORMES												
	Poeciliidae											
	<i>Poecilia vivipara</i> Bloch & Schneider, 1801								x		NE	LC
CARANGIFORMES												
	Rachycentridae											
	<i>Rachycentron canadum</i> (Linnaeus, 1766)										LC	LC
	Echeneidae											
	<i>Echeneis naucrates</i> Linnaeus, 1758			x				x			LC	LC
	<i>Phtheichthys lineatus</i> (Menziés, 1791)										LC	LC
	<i>Remora remora</i> (Linnaeus, 1758)							x			LC	LC
	Carangidae											
	<i>Carangoides bartholomaei</i> (Cuvier, 1833)										LC	LC
	<i>Caranx crysos</i> (Mitchill, 1815)							x			LC	LC
	<i>Caranx hippos</i> (Linnaeus, 1766)	x	x	x	x	x	x	x			LC	LC
	<i>Caranx latus</i> Agassiz, 1831	x	x	x	x	x	x	x			LC	LC
	<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	x	x	x	x	x	x	x			LC	LC
	<i>Hemicaranx amblyrhynchus</i> (Cuvier, 1833)										LC	LC
	<i>Oligoplites palometa</i> (Cuvier, 1832)	x	x	x	x	x	x	x			LC	LC
	<i>Oligoplites saliens</i> (Bloch, 1793)	x	x	x	x	x	x	x			LC	LC
	<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	x	x	x	x	x	x	x			LC	LC
	<i>Selene brownii</i> (Curvier, 1816)										LC	LC
	<i>Selene setapinnis</i> (Mitchill, 1815)					x		x	x		LC	LC
	<i>Selene vomer</i> (Linnaeus, 1758)		x	x	x	x		x			LC	LC
	<i>Trachinotus carolinus</i> (Linnaeus, 1766)			x				x			LC	LC

										LC	DD	
										LC	LC	
									x	LC	LC	
ISTIOPHORIFORMES												
	Sphyraenidae											
	<i>Sphyraena barracuda</i> (Edwards, 1771)		x		x	x			x	x	LC	LC
	<i>Sphyraena guachancho</i> Cuvier, 1829										LC	LC
PLEURONECTIFORMES												
	Paralichthyidae											
	<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900										LC	LC
	<i>Citharichthys cornutus</i> (Günther, 1880)										LC	LC
	<i>Citharichthys macrops</i> Dresel, 1885										LC	LC
	<i>Citharichthys spilopterus</i> Günther, 1862		x	x	x	x	x				LC	LC
	<i>Etropus crossotus</i> Jordan & Gilbert, 1882	x		x	x	x	x				LC	LC
	<i>Etropus longimanus</i> Norman, 1933										NE	LC
	<i>Paralichthys brasiliensis</i> (Ranzani, 1842)								x		NE	LC
	<i>Paralichthys orbignyanus</i> (Valenciennes, 1839)										DD	DD
	<i>Paralichthys tropicus</i> Ginsburg, 1933										DD	NE
	<i>Syacium micrurum</i> Ranzani, 1842				x	x				x	LC	LC
	<i>Syacium papillosum</i> (Linnaeus, 1758)										LC	LC
	Bothidae											
	<i>Bothus lunatus</i> (Linnaeus, 1758)										LC	LC
	<i>Bothus ocellatus</i> (Agassiz, 1831)								x	x	LC	LC
	<i>Bothus robinsi</i> Topp & Hoff, 1972										LC	LC
	Achiridae											
	<i>Achirus achirus</i> (Linnaeus, 1758)			x	x	x				x	LC	LC
	<i>Achirus declivis</i> Chabanaud, 1940	x	x	x	x	x	x			x	LC	LC
	<i>Achirus lineatus</i> (Linnaeus, 1758)		x	x	x	x				x	LC	LC

	<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009								NE	VU
	<i>Trinectes microphthalmus</i> (Chabanaud, 1928)								LC	LC
	<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	x	x	x	x	x	x		LC	LC
	Cynoglossidae									
	<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)								LC	LC
	<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)			x	x			x	LC	LC
	<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)		x						LC	LC
SYNGNATHIFORMES										
	Syngnathidae									
	<i>Bryx dunckeri</i> (Metzelaar, 1919)								LC	LC
	<i>Cosmocampus elucens</i> (Poey, 1868)								LC	LC
	<i>Hippocampus erectus</i> Perry, 1810								VU	VU
	<i>Hippocampus reidi</i> Ginsburg, 1933		x	x	x	x	x	x	NT	VU
	<i>Microphis brachyurus lineatus</i> (Kaup, 1856)							x	NE	NE
	<i>Microphis brachyurus brachyurus</i> (Bleeker, 1854)			x	x			x	LC	LC
	Fistulariidae									
	<i>Fistularia petimba</i> Lacepède, 1803								LC	LC
	<i>Fistularia tabacaria</i> Linnaeus, 1758								LC	LC
	<i>Syngnathus pelagicus</i> Linnaeus, 1758			x	x			x	LC	LC
	Dactylopteridae									
	<i>Dactylopterus volitans</i> (Linnaeus, 1758)								LC	LC
SCOMBRIFORMES										
	Trichiuridae									
	<i>Trichiurus lepturus</i> Linnaeus, 1758		x		x	x		x	LC	LC
	Scombridae									
	<i>Euthymus alletteratus</i> (Rafinesque, 1810)								LC	LC
	<i>Scomberomorus brasiliensis</i> Collette, Russo & Zavala-Camin, 1978		x	x	x			x	LC	LC

	<i>Scomberomorus cavalla</i> (Cuvier, 1829)								LC	LC	
	<i>Scomberomorus maculatus</i> (Mitchill, 1815)							x	LC	NE	
	<i>Scomberomorus regalis</i> (Bloch, 1793)								LC	LC	
	Stromateidae										
	<i>Peprilus paru</i> (Linnaeus, 1758)		x			x		x	LC	LC	
TRACHINIFORMES											
	Uranoscopidae										
	<i>Astroscopus y-graecum</i> Cuvier, 1829								LC	LC	
LABRIFORMES											
	Labridae										
	<i>Halichoeres poeyi</i> (Steindachner, 1867)								LC	LC	
	Scaridae										
	<i>Nicholsina usta</i> (Valenciennes, 1840)								LC	LC	
	<i>Sparisoma amplum</i> (Ranzani, 1842)								LC	NT	
	<i>Sparisoma axillare</i> (Steindachner, 1878)								DD	VU	
	<i>Sparisoma radians</i> (Valenciennes, 1840)							x	LC	LC	
PERCIFORMES											
	Centropomidae										
	<i>Centropomus ensiferus</i> Poey, 1860	x	x	x	x	x	x	x	LC	LC	
	<i>Centropomus mexicanus</i> Bocourt, 1868								LC	LC	
	<i>Centropomus parallelus</i> Poey, 1860	x	x	x				x	LC	LC	
	<i>Centropomus pectinatus</i> Poey, 1860				x				LC	LC	
	<i>Centropomus undecimalis</i> (Bloch, 1792)	x	x	x	x	x	x	x	LC	LC	
	Gerreidae										
	<i>Diapterus auratus</i> Ranzani, 1842	x	x	x	x	x	x	x	LC	LC	
	<i>Diapterus rhombeus</i> (Cuvier, 1829)		x	x	x	x		x	x	LC	LC
	<i>Eucinostomus argenteus</i> Baird & Girard, 1855			x	x			x	x	LC	LC
	<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)			x	x	x		x		LC	LC

<i>Eucinostomus havana</i> (Nichols, 1912)				x	x		x			LC	LC	
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	x	x	x	x	x	x	x	x	x	LC	LC	
<i>Eugerres brasilianus</i> (Cuvier, 1830)		x	x	x	x	x	x			LC	LC	
<i>Gerres cinereus</i> (Walbaum, 1792)	x	x	x	x	x	x	x			LC	LC	
<i>Ulaema lefroyi</i> (Goode, 1874)										LC	LC	
Mullidae												
<i>Pseudupeneus maculatus</i> (Bloch, 1793)									x	x	LC	LC
Serranidae												
<i>Alphestes afer</i> (Bloch, 1793)											LC	DD
<i>Diplectrum radiale</i> (Quoy & Gaimard, 1824)		x	x	x	x	x	x				LC	LC
<i>Epinephelus adscensionis</i> (Osbeck, 1765)											LC	DD
<i>Epinephelus itajara</i> (Lichtenstein, 1822)			x	x	x	x	x				VU	CR
<i>Hyporthodus nigritus</i> (Holbrook, 1855)											NT	EN
<i>Mycteroperca bonaci</i> (Poey, 1860)	x	x	x	x	x	x	x				NT	VU
<i>Mycteroperca tigris</i> (Valenciennes, 1833)											DD	DD
<i>Mycteroperca venenosa</i> (Linnaeus, 1758)											NT	DD
<i>Rypticus randalli</i> Courtenay, 1967	x	x	x	x	x	x	x				LC	LC
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)											LC	LC
<i>Serranus flaviventris</i> (Cuvier, 1829)											LC	LC
<i>Serranus phoebe</i> Poey, 1851											LC	LC
Pomacanthidae												
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)											LC	DD
<i>Pomacanthus paru</i> (Bloch, 1787)									x	x	LC	DD
Chaetodontidae												
<i>Chaetodon ocellatus</i> (Bloch, 1787)											LC	DD
<i>Chaetodon striatus</i> Linnaeus, 1758											LC	LC
Haemulidae												
<i>Anisotremus surinamensis</i> (Bloch, 1791)											DD	DD

<i>Anisotremus virginicus</i> (Linnaeus, 1758)	x	x	x	x	x	x	x	x	x	LC	LC
<i>Conodon nobilis</i> (Linnaeus, 1758)		x								LC	LC
<i>Genyatremus luteus</i> (Bloch, 1790)	x	x	x	x	x	x	x			NE	LC
<i>Haemulon aurolineatum</i> Cuvier, 1830								x	x	LC	LC
<i>Haemulon parra</i> (Desmarest, 1823)			x		x			x	x	LC	LC
<i>Haemulon plumierii</i> (Lacepède, 1801)								x	x	LC	DD
<i>Haemulon squamipinna</i> Rocha & Rosa, 1999										NE	LC
<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)								x	x	LC	LC
<i>Orthopristis ruber</i> (Cuvier, 1830)								x	x	LC	LC
<i>Pomadasys corvinaeformis</i> (Steindachner, 1868)	x	x	x	x	x	x	x	x	x	LC	LC
<i>Pomadasys crocro</i> (Cuvier, 1830)										DD	LC
<i>Pomadasys ramosus</i> Poey, 1860										NE	LC
Lutjanidae											
<i>Lutjanus alexandrei</i> Moura & Linderman, 2007			x		x					NE	LC
<i>Lutjanus analis</i> (Cuvier, 1828)		x	x	x	x	x	x	x	x	NT	NT
<i>Lutjanus apodus</i> (Walbaum, 1792)	x	x	x	x	x	x	x	x	x	LC	NE
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)					x			x	x	VU	VU
<i>Lutjanus griseus</i> (Linnaeus, 1758)							x	x	x	LC	NE
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	x	x	x	x	x	x	x			DD	NT
<i>Lutjanus synagris</i> (Linnaeus, 1758)			x		x	x	x	x	x	NT	NT
<i>Ocyurus chrysurus</i> (Bloch, 1791)										DD	NT
Polynemidae											
<i>Polydactylus oligodon</i> (Gunther, 1860)										LC	LC
<i>Polydactylus virginicus</i> (Linnaeus, 1758)				x	x		x	x	x	LC	LC
SCORPAENIFORMES											
Scorpaenidae											
<i>Scorpaena isthmensis</i> Meek & Hilderbrand, 1928										LC	LC
<i>Scorpaena plumieri</i> Bloch, 1789				x	x		x			LC	LC

Triglidae											
<i>Prionotus punctatus</i> (Bloch, 1793)											
									x	LC	LC
MORONIFORMES											
Ephippidae											
<i>Chaetodipterus faber</i> (Broussonet, 1782)											
x	x	x	x	x	x	x	x			LC	LC
ACANTHURIFORMES											
Sciaenidae											
<i>Bairdiella ronchus</i> (Cuvier, 1830)											
x	x	x	x	x	x	x	x			LC	LC
<i>Cynoscion acoupa</i> (Lacepède, 1801)											
x	x	x	x	x	x	x	x	x	x	LC	NT
<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1833)											
										LC	LC
<i>Cynoscion leiarchus</i> (Cuvier, 1830)											
x	x	x	x	x	x	x	x		x	LC	LC
<i>Cynoscion microlepidotus</i> (Cuvier, 1830)											
x	x	x	x	x	x	x	x			LC	LC
<i>Cynoscion steindachneri</i> (Jordan, 1889)											
										LC	LC
<i>Cynoscion virescens</i> (Cuvier, 1830)											
	x					x	x			LC	LC
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)											
										LC	LC
<i>Larimus breviceps</i> Cuvier, 1830											
								x	x	LC	LC
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)											
										LC	LC
<i>Menticirrhus americanus</i> (Linnaeus, 1758)											
x	x	x	x	x	x	x	x	x	x	LC	DD
<i>Menticirrhus littoralis</i> (Holbrook, 1847)											
x	x	x	x	x	x	x	x			LC	DD
<i>Micropogonias furnieri</i> (Desmarest, 1823)											
x	x	x	x	x	x	x	x	x	x	LC	LC
<i>Nebris microps</i> Cuvier, 1830											
										LC	LC
<i>Odontoscion dentex</i> (Cuvier, 1830)											
										LC	LC
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925											
							x			LC	DD
<i>Paralonchurus brasiliensis</i> (Steindachner, 1875)											
								x	x	LC	LC
<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)											
										LC	DD
<i>Pogonias cromis</i> (Linnaeus, 1766)											
										LC	EN
<i>Stellifer brasiliensis</i> (Schultz, 1945)											
										NE	LC
<i>Stellifer microps</i> (Steindachner, 1864)											
										LC	LC

										LC	LC		
										LC	LC		
										DD	LC		
										LC	LC		
	Acanthuridae												
	<i>Acanthurus bahianus</i> Castelnau, 1855		x	x	x	x				x	LC	LC	
	<i>Acanthurus chirurgus</i> (Bloch, 1787)									x	x	LC	LC
	<i>Acanthurus coeruleus</i> Bloch & Schneider, 1801		x	x	x	x				x	x	LC	LC
SPARIFORMES													
	Lobotidae												
	<i>Lobotes surinamensis</i> (Bloch, 1790)	x	x	x	x	x	x	x				LC	LC
	Sparidae												
	<i>Archosargus probatocephalus</i> (Walbaum, 1792)		x	x	x	x	x	x				LC	DD
	<i>Archosargus rhomboidalis</i> (Linnaeus, 1758)			x						x	x	LC	LC
LOPHIIFORMES													
	Antennariidae												
	<i>Antennarius multiocellatus</i> (Valenciennes, 1837)											LC	DD
	<i>Antennarius striatus</i> (Shaw, 1794)		x	x	x	x	x	x				LC	DD
	Ogcocephalidae												
	<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	x	x	x	x	x	x	x				NE	LC
TETRAODONTIFORMES													
	Ostraciidae												
	<i>Acanthostracion quadricornis</i> (Linnaeus, 1758)											LC	LC
	<i>Lactophrys trigonus</i> (Linnaeus, 1758)											LC	LC
	<i>Lactophrys triqueter</i> (Linnaeus, 1758)											LC	NE
	Balistidae												
	<i>Balistes vetula</i> Linnaeus, 1758											NT	NT
	<i>Melichthys niger</i> (Bloch, 1786)									x		LC	LC

Monacanthidae											
<i>Aluterus heudelotii</i>										LC	LC
<i>Aluterus monoceros</i> (Linnaeus, 1758)										LC	NT
<i>Aluterus schoepfii</i> (Walbaum, 1792)										LC	LC
<i>Cantherhines pullus</i> (Ranzani, 1842)										LC	LC
<i>Monacanthus ciliatus</i> (Mitchill, 1818)										LC	LC
<i>Stephanolepis hispidus</i> (Linnaeus, 1766)										LC	LC
<i>Stephanolepis setifer</i> (Bennett, 1831)										LC	LC
Tetraodontidae											
<i>Canthigaster figueiredoi</i> Moura & Castro, 2002										LC	LC
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	x	x	x	x	x	x	x			LC	LC
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)		x	x	x	x	x	x			LC	LC
<i>Sphoeroides greeleyi</i> Gilbert, 1900			x							LC	LC
<i>Sphoeroides nephelus</i> (Goode & Bean, 1882)										LC	NE
<i>Sphoeroides spengleri</i> (Bloch, 1785)										LC	LC
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	x	x	x	x	x	x	x	x	x	LC	DD
<i>Sphoeroides tyleri</i> Shipp, 1972										LC	LC
Diodontidae											
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897										LC	LC
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)		x	x	x	x				x	LC	LC
<i>Diodon holocanthus</i> Linnaeus, 1758										LC	LC
<i>Diodon hystrix</i> Linnaeus, 1758										LC	LC

Table III: List of fish species recorded in 45 estuaries in north-eastern Brazil, along with their conservation status (ICMbio and IUCN list). Estuaries: Porto do Mangue, Tubarão River, Guamaré, Galinhos, Barra do Rio, Potengí, Forte, Tibau do Sul and Barra de Cunhaú. ICMbio and IUCN classification: endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

Species	Porto do Mangue-RN	Tubarão River-RN	Guamaré-RN	Galinhos-RN	Barra do Rio-RN	Potengí-RN	Forte-RN	Tibau do Sul-RN	Barra de Cunhaú-RN	IUCN	ICMbio
ORECTOLOBIFORMES											
Ginglymostomatidae											
<i>Ginglymostoma cirratum</i> (Bonnaterre, 1788)										VU	VU
CARCHARHINIFORMES											
Carcharhinidae											
<i>Carcharhinus leucas</i> (Muller & Henle, 1839)										NT	NT
<i>Carcharhinus limbatus</i> (Muller & Henle, 1839)										NT	NT
<i>Carcharhinus porosus</i> (Ranzani, 1839)										CR	CR
<i>Isogomphodon oxyrinchus</i> (Muller & Henle, 1839)										CR	CR
<i>Rhizoprionodon lalandii</i> (Muller & Henle, 1839)										VU	NT
<i>Rhizoprionodon porosus</i> (Poey, 1861)										VU	DD
Sphyrnidae											
<i>Sphyrna lewini</i> (Griffith & Smith, 1834)										CR	CR
<i>Sphyrna tiburo</i> (Linnaeus, 1758)										EN	CR
<i>Sphyrna tudes</i> (Valenciennes, 1822)										CR	CR
TORPEDINIFORMES											
Narcinidae											
<i>Narcine brasiliensis</i> (Olfers, 1831)			x					x		NT	DD
PRISTIFORMES											
Pristidae											
<i>Pristis pristis</i> Muller & Henle, 1841										CR	CR

	Rhinobatidae								
	<i>Pseudobatos percellens</i> (Walbaum, 1792)							EN	DD
MYLIOBATIFORMES									
	Dasyatidae								
	<i>Fontitrygon geijskesi</i> (Boeseman, 1948)							CR	DD
	<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)							NT	DD
	<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)							NT	LC
	<i>Hypanus marianae</i> (Gomes, Rosa & Gadig, 2000)							EN	DD
	<i>Hypanus say</i> (Lesueur, 1817)							NT	DD
	Gymnuridae								
	<i>Gymnura micrura</i> (Bloch & Scheneider, 1801)							NT	NT
	Myliobatidae								
	<i>Aetobatus narinari</i> (Euphrasen, 1790)							EN	DD
	<i>Mobula hypostoma</i> (Bancroft, 1831)							EN	VU
	<i>Rhinoptera bonasus</i> (Mitchill, 1815)							VU	DD
ELOPIFORMES									
	Elopidae								
	<i>Elops saurus</i> Linnaeus, 1766	x		x		x		x	LC NE
	Megalopidae								
	<i>Megalops atlanticus</i> Valenciennes, 1847	x				x			VU VU
ALBULIFORMES									
	Albulidae								
	<i>Albula vulpes</i> (Linnaeus, 1758)		x						NT DD
ANGUILLIFORMES									
	Muraenidae								
	<i>Gymnothorax funebris</i> Ranzani, 1839								LC DD
	<i>Gymnothorax moringa</i> (Cuvier, 1829)								LC DD
	<i>Gymnothorax ocellatus</i> Agassiz, 1831								LC DD

	<i>Gymnothorax vicinus</i> (Castelnau, 1855)					LC	DD
	Ophichthidae						
	<i>Ahlia egmontis</i> (Jordan, 1884)					LC	LC
	<i>Myrichthys breviceps</i> (Richardson, 1848)					LC	LC
	<i>Myrichthys ocellatus</i> (Lesueur, 1825)	x		x		LC	LC
	<i>Myrophis punctatus</i> Lütken, 1852					LC	LC
	<i>Ophichthus cylindroideus</i> (Ranzani, 1839)					LC	LC
	<i>Ophichthus ophis</i> (Linnaeus, 1758)					LC	LC
	Muraenesocidae						
	<i>Cynoponticus savanna</i> (Bancroft, 1831)					LC	LC
	Congridae						
	<i>Ariosoma balearicum</i> (Delaroche, 1809)					LC	LC
CLUPEIFORMES							
	Pristigasteridae						
	<i>Chirocentrodon bleekermanus</i> (Poey, 1867)			x		LC	LC
	<i>Odontognathus mucronatus</i> Lacepède, 1800					LC	LC
	<i>Pellona flavipinnis</i> (Valenciennes, 1837)					LC	LC
	<i>Pellona harroweri</i> (Fowler, 1917)	x		x		LC	LC
	Engraulidae						
	<i>Anchoa filifera</i> (Fowler, 1915)					LC	LC
	<i>Anchoa hepsetus</i> (Linnaeus, 1758)					LC	NE
	<i>Anchoa januaria</i> (Steindachner, 1879)					LC	LC
	<i>Anchoa lyolepis</i> (Evermann & Marsh, 1900)					LC	LC
	<i>Anchoa parva</i> (Meek & Hildebrand, 1923)					LC	NE
	<i>Anchoa spinifer</i> (Valenciennes, 1848)					LC	LC
	<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)					LC	LC
	<i>Anchovia clupeioides</i> (Swainson, 1839)					LC	LC
	<i>Anchoviella lepidentostole</i> (Fowler, 1911)					LC	LC

<i>Cetengraulis edentulus</i> (Cuvier, 1829)									LC	LC
<i>Engraulis anchoita</i> Hubbs & Marini 1935									LC	LC
<i>Lycengraulis batesii</i> (Gunther, 1868)									LC	LC
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)									LC	LC
<i>Pterengraulis atherinoides</i> (Linnaeus, 1766)									LC	LC
Clupeidae										
<i>Harengula clupeola</i> (Cuvier, 1829)			x						LC	LC
<i>Harengula jaguana</i> Poey, 1865					x			x	LC	LC
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)					x	x		x	LC	LC
<i>Opisthonema oglinum</i> (Lesueur, 1818)	x	x	x	x				x	LC	LC
<i>Platanichthys platana</i> (Regan, 1917)									LC	LC
<i>Rhinosardinia amazonica</i> (Steindachner, 1879)								x	LC	LC
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)									LC	LC
CHARACIFORMES										
Erythrinidae										
<i>Hoplias malabaricus</i> (Bloch, 1794)									LC	LC
<i>Prochilodus argenteus</i> Spix & Agassiz, 1829									NE	LC
Serrasalminidae										
<i>Pygocentrus nattereri</i> Kner, 1858									NE	LC
<i>Serrasalmus rhombeus</i> (Linnaeus, 1766)									NE	LC
<i>Serrasalmus spilopleura</i> Kner, 1858									NE	LC
Anostomidae										
<i>Schizodon fasciatus</i> Spix & Agassiz, 1829									NE	LC
<i>Leporinus friderici</i> (Bloch, 1794)									NE	LC
Curimatidae										
<i>Steindachnerina elegans</i> (Steindachner, 1875)									NE	LC
Characidae										
<i>Astyanax bimaculatus</i> (Linnaeus, 1758)									NE	LC

									NE	LC
SILURIFORMES	<i>Hemigrammus marginatus</i> Ellis, 1911									
	Loricariidae									
	<i>Hypostomus plecostomus</i> (Linnaeus, 1758)								NE	NE
	<i>Loricaria paraguayae</i> Steindachner, 1907								NE	LC
	Ariidae									
	<i>Aspistor luniscutis</i> (Valenciennes, 1840)								NE	LC
	<i>Aspistor quadriscutis</i> (Valenciennes, 1840)								LC	LC
	<i>Bagre bagre</i> (Linnaeus, 1766)								LC	NT
	<i>Bagre marinus</i> (Mitchill, 1815)	x	x	x			x		LC	DD
	<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)								NE	LC
	<i>Cathorops spixii</i> (Agassiz, 1829)	x	x				x		NE	LC
	<i>Genidens genidens</i> (Cuvier, 1829)								LC	LC
	<i>Notarius grandicassis</i> (Valenciennes, 1840)								LC	LC
	<i>Sciades couma</i> (Valenciennes, 1840)								LC	DD
	<i>Sciades herzbergii</i> (Bloch, 1794)								LC	LC
	<i>Sciades parkeri</i> (Traill, 1832)								VU	VU
	<i>Sciades proops</i> (Valenciennes, 1840)								NE	DD
	Aspredinidae									
	<i>Aspredo aspredo</i> (Linnaeus, 1758)								LC	LC
	<i>Aspredinichthys tiben</i> (Valenciennes, 1840)								LC	LC
	Auchenipteridae									
	<i>Ageneiosus inermis</i> (Linnaeus, 1766)								NE	LC
	<i>Pseudauchenipterus nodosus</i> (Bloch, 1794)								NE	LC
	<i>Trachelyopterus galeatus</i> (Linnaeus, 1766)								NE	LC
	Heptapteridae									
	<i>Pimelodella cristata</i> (Muller & Troschel, 1849)								LC	LC
	<i>Pimelodella lateristriga</i> (Lichtenstein, 1823)								NE	DD

	Pimelodidae									
	<i>Brachyplatystoma vaillantii</i> (Valenciennes, 1840)								NE	LC
	<i>Pseudoplatystoma fasciatum</i> (Linnaeus, 1766)								NE	NE
	<i>Sorubim lima</i> (Bloch & Schneider, 1801)								NE	LC
AULOPIFORMES										
	Synodontidae									
	<i>Synodus foetens</i> (Linnaeus, 1766)								LC	LC
	<i>Synodus poeyi</i> Jordan, 1887								LC	LC
	<i>Trachinocephalus myops</i> (Forster, 1801)								LC	LC
HOLOCENTRIFORMES										
	Holocentridae									
	<i>Holocentrus adscensionis</i> (Osbeck, 1765)								LC	LC
BATRACHOIDIFORMES										
	Batrachoididae									
	<i>Amphichthys cryptocentrus</i> (Valenciennes, 1837)								LC	LC
	<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)								LC	LC
	<i>Thalassophryne nattereri</i> Steindachner, 1876								LC	LC
	<i>Thalassophryne punctata</i> Steindachner, 1876								NE	LC
KURTIFORMES										
	Apogonidae									
	<i>Phaeoptyx pigmentaria</i> (Poey, 1860)								LC	LC
GOBIIFORMES										
	Eleotridae									
	<i>Dormitator maculatus</i> (Bloch, 1792)								LC	NT
	<i>Eleotris pisonis</i> (Gmelin, 1789)								LC	LC
	<i>Erotelis smaragdus</i> (Valenciennes, 1837)								LC	LC
	<i>Guavina guavina</i> (Valenciennes, 1837)								LC	LC
	Gobiidae									

<i>Awaous tajasica</i> (Lichtenstein, 1822)	x							x		LC	LC
<i>Bathygobius mystacium</i> Ginsburg, 1947					x	x	x			LC	NE
<i>Bathygobius soporator</i> (Valenciennes, 1837)	x	x	x	x	x	x	x	x	x	LC	LC
<i>Coryphopterus glaucofraenum</i> Gill, 1863										LC	LC
<i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882)		x	x	x	x	x	x	x	x	LC	LC
<i>Ctenogobius saepepallens</i> (Gilbert & Randall, 1968)										LC	LC
<i>Ctenogobius shufeldti</i> (Jordan & Eigenmann, 1887)										LC	LC
<i>Ctenogobius smaragdus</i> (Valenciennes, 1837)		x						x		LC	LC
<i>Ctenogobius stigmaticus</i> (Poey, 1860)										LC	LC
<i>Evorthodus lyricus</i> (Girard, 1858)					x	x		x		LC	LC
<i>Gobioides broussonneti</i> Lacepède, 1800										LC	LC
<i>Gobionellus oceanicus</i> (Pallas, 1770)	x		x		x	x		x	x	LC	LC
<i>Gobionellus stomatus</i> Starks, 1913								x		NE	LC
<i>Microdesmus bahianus</i> Dawson, 1973										LC	LC
<i>Microdesmus longipinnis</i> (Weymouth, 1910)										LC	LC
<i>Microgobius meeki</i> Evermann & Marsh, 1899					x	x		x		LC	LC
Pomacentridae											
<i>Abudefduf saxatilis</i> (Linnaeus, 1758)	x		x	x	x	x	x	x	x	LC	LC
<i>Stegastes fuscus</i> (Cuvier, 1830)	x		x	x	x	x	x	x	x	LC	LC
<i>Stegastes variabilis</i> (Castelnau, 1855)	x		x	x	x	x	x	x	x	NE	LC
Opistognathidae											
<i>Opistognathus cuvierii</i> Valenciennes, 1836										NE	LC
MUGILIFORMES											
Mugilidae											
<i>Mugil brevirostris</i> (Ribeiro, 1915)										NE	NE
<i>Mugil curema</i> Valenciennes, 1836	x	x	x	x		x	x		x	LC	DD
<i>Mugil curvidens</i> Valenciennes, 1836			x	x	x	x				LC	DD
<i>Mugil incilis</i> Hancock, 1830					x	x			x	LC	LC

	<i>Mugil liza</i> Valenciennes, 1836	x		x		x		x	DD	NT
	<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007								LC	DD
CICHLIFORMES										
	Cichlidae									
	<i>Cichla ocellaris</i> Bloch & Schneider, 1801								NE	LC
	<i>Oreochromis niloticus</i> (Linnaeus, 1758)								LC	NE
BLENNIIFORMES										
	Dactyloscopidae									
	<i>Dactyloscopus crossotus</i> Starks, 1913								LC	LC
	Blenniidae									
	<i>Scartella cristata</i> (Linnaeus, 1758)								LC	LC
	Labrisomidae									
	<i>Labrisomus nuchipinnis</i> (Quoy & Gaimard, 1824)								LC	LC
ATHERINIFORMES										
	Atherinidae									
	<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)			x		x		x	LC	LC
	<i>Atherinella cf. blackburni</i> (Schultz, 1949)								LC	LC
	<i>Odontesthes bonariensis</i> (Valenciennes, 1835)								NE	DD
BELONIFORMES										
	Exocoetidae									
	<i>Cheilopogon melanurus</i> (Valenciennes, 1847)								LC	LC
	<i>Hirundichthys affinis</i> (Gunther, 1866)								LC	LC
	Hemiramphidae									
	<i>Hemiramphus balao</i> Lesueur, 1821					x		x	LC	DD
	<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)					x		x	LC	LC
	<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)	x						x	LC	LC
	<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)		x					x	LC	NT
	Belonidae									

	<i>Ablennes hians</i> (Valenciennes, 1846)		x		x		x		x	LC	LC
	<i>Strongylura marina</i> (Walbaum, 1792)									LC	LC
	<i>Strongylura timucu</i> (Walbaum, 1792)	x		x			x		x	LC	LC
	<i>Tylosurus acus acus</i> (Lacepède, 1803)									LC	LC
	<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)									LC	LC
CYPRINODONTIFORMES											
	Poeciliidae										
	<i>Poecilia vivipara</i> Bloch & Schneider, 1801	x		x		x		x		x	NE LC
CARANGIFORMES											
	Rachycentridae										
	<i>Rachycentron canadum</i> (Linnaeus, 1766)									LC	LC
	Echeneidae										
	<i>Echeneis naucrates</i> Linnaeus, 1758			x						LC	LC
	<i>Phtheichthys lineatus</i> (Menziés, 1791)									LC	LC
	<i>Remora remora</i> (Linnaeus, 1758)									LC	LC
	Carangidae										
	<i>Carangoides bartholomaei</i> (Cuvier, 1833)									LC	LC
	<i>Caranx crysos</i> (Mitchill, 1815)									LC	LC
	<i>Caranx hippos</i> (Linnaeus, 1766)			x						LC	LC
	<i>Caranx latus</i> Agassiz, 1831			x						LC	LC
	<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)									LC	LC
	<i>Hemicaranx amblyrhynchus</i> (Cuvier, 1833)									LC	LC
	<i>Oligoplites palometa</i> (Cuvier, 1832)									LC	LC
	<i>Oligoplites saliens</i> (Bloch, 1793)									LC	LC
	<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	x		x					x	LC	LC
	<i>Selene brownii</i> (Curvier, 1816)									LC	LC
	<i>Selene setapinnis</i> (Mitchill, 1815)	x		x		x			x	LC	LC
	<i>Selene vomer</i> (Linnaeus, 1758)	x		x		x			x	LC	LC

	<i>Trachinotus carolinus</i> (Linnaeus, 1766)	x		x			x		x	x	LC	LC
	<i>Trachinotus cayennensis</i> Cuvier, 1832										LC	DD
	<i>Trachinotus falcatus</i> (Linnaeus, 1758)	x	x	x			x		x	x	LC	LC
	<i>Trachinotus goodei</i> Jordan & Evermann, 1896	x		x			x			x	LC	LC
ISTIOPHORIFORMES												
	Sphyraenidae											
	<i>Sphyraena barracuda</i> (Edwards, 1771)		x	x		x				x	LC	LC
	<i>Sphyraena guachancho</i> Cuvier, 1829										LC	LC
PLEURONECTIFORMES												
	Paralichthyidae											
	<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900										LC	LC
	<i>Citharichthys cornutus</i> (Günther, 1880)		x								LC	LC
	<i>Citharichthys macrops</i> Dresel, 1885		x								LC	LC
	<i>Citharichthys spilopterus</i> Günther, 1862		x								LC	LC
	<i>Etropus crossotus</i> Jordan & Gilbert, 1882										LC	LC
	<i>Etropus longimanus</i> Norman, 1933										NE	LC
	<i>Paralichthys brasiliensis</i> (Ranzani, 1842)	x		x			x		x		NE	LC
	<i>Paralichthys orbignyanus</i> (Valenciennes, 1839)										DD	DD
	<i>Paralichthys tropicus</i> Ginsburg, 1933										DD	NE
	<i>Syacium micrurum</i> Ranzani, 1842	x		x		x	x		x	x	LC	LC
	<i>Syacium papillosum</i> (Linnaeus, 1758)	x	x	x		x	x		x	x	LC	LC
	Bothidae											
	<i>Bothus lunatus</i> (Linnaeus, 1758)	x		x					x		LC	LC
	<i>Bothus ocellatus</i> (Agassiz, 1831)	x	x	x		x	x		x	x	LC	LC
	<i>Bothus robinsi</i> Topp & Hoff, 1972	x				x			x	x	LC	LC
	Achiridae											
	<i>Achirus achirus</i> (Linnaeus, 1758)										LC	LC
	<i>Achirus declivis</i> Chabanaud, 1940										LC	LC

<i>Achirus lineatus</i> (Linnaeus, 1758)	x	x	x		x		x	x	LC	LC
<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009									NE	VU
<i>Trinectes microphthalmus</i> (Chabanaud, 1928)	x		x			x		x	LC	LC
<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	x		x					x	LC	LC
Cynoglossidae										
<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)									LC	LC
<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)									LC	LC
<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)									LC	LC
SYNGNATHIFORMES										
Syngnathidae										
<i>Bryx dunckeri</i> (Metzelaar, 1919)									LC	LC
<i>Cosmocampus elucens</i> (Poey, 1868)			x						LC	LC
<i>Hippocampus erectus</i> Perry, 1810	x		x		x				VU	VU
<i>Hippocampus reidi</i> Ginsburg, 1933	x	x	x		x				NT	VU
<i>Microphis brachyurus lineatus</i> (Kaup, 1856)									NE	NE
<i>Microphis brachyurus brachyurus</i> (Bleeker, 1854)									LC	LC
Fistulariidae										
<i>Fistularia petimba</i> Lacepède, 1803									LC	LC
<i>Fistularia tabacaria</i> Linnaeus, 1758	x				x	x	x		LC	LC
<i>Syngnathus pelagicus</i> Linnaeus, 1758									LC	LC
Dactylopteridae										
<i>Dactylopterus volitans</i> (Linnaeus, 1758)			x						LC	LC
SCOMBRIFORMES										
Trichiuridae										
<i>Trichiurus lepturus</i> Linnaeus, 1758									LC	LC
Scombridae										
<i>Euthynnus alletteratus</i> (Rafinesque, 1810)									LC	LC

	<i>Scomberomorus brasiliensis</i> Collette, Russo & Zavala-Camin, 1978	x								LC	LC
	<i>Scomberomorus cavalla</i> (Cuvier, 1829)									LC	LC
	<i>Scomberomorus maculatus</i> (Mitchill, 1815)									LC	NE
	<i>Scomberomorus regalis</i> (Bloch, 1793)									LC	LC
	Stromateidae										
	<i>Peprilus paru</i> (Linnaeus, 1758)									LC	LC
TRACHINIFORMES											
	Uranoscopidae										
	<i>Astroscopus y-graecum</i> Cuvier, 1829									LC	LC
LABRIFORMES											
	Labridae										
	<i>Halichoeres poeyi</i> (Steindachner, 1867)									LC	LC
	Scaridae										
	<i>Nicholsina usta</i> (Valenciennes, 1840)		x							LC	LC
	<i>Sparisoma amplum</i> (Ranzani, 1842)									LC	NT
	<i>Sparisoma axillare</i> (Steindachner, 1878)									DD	VU
	<i>Sparisoma radians</i> (Valenciennes, 1840)	x		x	x			x		LC	LC
PERCIFORMES											
	Centropomidae										
	<i>Centropomus ensiferus</i> Poey, 1860	x							x	LC	LC
	<i>Centropomus mexicanus</i> Bocourt, 1868							x		LC	LC
	<i>Centropomus parallelus</i> Poey, 1860					x	x			LC	LC
	<i>Centropomus pectinatus</i> Poey, 1860					x				LC	LC
	<i>Centropomus undecimalis</i> (Bloch, 1792)	x	x	x	x	x	x			LC	LC
	Gerreidae										
	<i>Diapterus auratus</i> Ranzani, 1842		x			x	x			LC	LC
	<i>Diapterus rhombeus</i> (Cuvier, 1829)	x	x	x	x	x	x	x	x	LC	LC

<i>Eucinostomus argenteus</i> Baird & Girard, 1855	x	x	x	x	x	x	x	x		LC	LC
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	x		x	x	x	x	x	x	x	LC	LC
<i>Eucinostomus havana</i> (Nichols, 1912)										LC	LC
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	x	x	x	x	x	x		x	x	LC	LC
<i>Eugerres brasiliensis</i> (Cuvier, 1830)		x	x	x	x	x			x	LC	LC
<i>Gerres cinereus</i> (Walbaum, 1792)										LC	LC
<i>Ulaema lefroyi</i> (Goode, 1874)		x	x	x	x	x	x	x	x	LC	LC
Mullidae											
<i>Pseudupeneus maculatus</i> (Bloch, 1793)	x		x	x		x	x	x	x	LC	LC
Serranidae											
<i>Alphestes afer</i> (Bloch, 1793)		x								LC	DD
<i>Diplectrum radiale</i> (Quoy & Gaimard, 1824)		x								LC	LC
<i>Epinephelus adscensionis</i> (Osbeck, 1765)		x								LC	DD
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	x		x			x				VU	CR
<i>Hyporthodus nigritus</i> (Holbrook, 1855)		x								NT	EN
<i>Mycteroperca bonaci</i> (Poey, 1860)	x	x	x							NT	VU
<i>Mycteroperca tigris</i> (Valenciennes, 1833)		x								DD	DD
<i>Mycteroperca venenosa</i> (Linnaeus, 1758)						x				NT	DD
<i>Rypticus randalli</i> Courtenay, 1967						x			x	LC	LC
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)										LC	LC
<i>Serranus flaviventris</i> (Cuvier, 1829)										LC	LC
<i>Serranus phoebe</i> Poey, 1851		x								LC	LC
Pomacanthidae											
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)										LC	DD
<i>Pomacanthus paru</i> (Bloch, 1787)	x		x	x		x	x	x	x	LC	DD
Chaetodontidae											
<i>Chaetodon ocellatus</i> (Bloch, 1787)										LC	DD
<i>Chaetodon striatus</i> Linnaeus, 1758										LC	LC

Haemulidae										
<i>Anisotremus surinamensis</i> (Bloch, 1791)		x							DD	DD
<i>Anisotremus virginicus</i> (Linnaeus, 1758)	x		x	x	x	x	x	x	LC	LC
<i>Conodon nobilis</i> (Linnaeus, 1758)	x		x	x					LC	LC
<i>Genyatremus luteus</i> (Bloch, 1790)	x				x	x		x	NE	LC
<i>Haemulon aurolineatum</i> Cuvier, 1830	x		x	x	x	x	x	x	LC	LC
<i>Haemulon parra</i> (Desmarest, 1823)	x		x	x	x	x		x	LC	LC
<i>Haemulon plumierii</i> (Lacepède, 1801)	x		x	x	x	x	x	x	LC	DD
<i>Haemulon squamipinna</i> Rocha & Rosa, 1999									NE	LC
<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)	x		x	x	x	x	x	x	LC	LC
<i>Orthopristis ruber</i> (Cuvier, 1830)	x	x	x	x	x	x	x	x	LC	LC
<i>Pomadasys corvinaeformis</i> (Steindachner, 1868)	x	x	x	x	x	x	x	x	LC	LC
<i>Pomadasys crocro</i> (Cuvier, 1830)						x			DD	LC
<i>Pomadasys ramosus</i> Poey, 1860									NE	LC
Lutjanidae										
<i>Lutjanus alexandrei</i> Moura & Linderman, 2007		x							NE	LC
<i>Lutjanus analis</i> (Cuvier, 1828)	x	x	x	x	x	x	x	x	NT	NT
<i>Lutjanus apodus</i> (Walbaum, 1792)	x	x	x	x	x	x	x	x	LC	NE
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	x	x	x	x	x	x	x	x	VU	VU
<i>Lutjanus griseus</i> (Linnaeus, 1758)	x	x	x	x	x	x	x	x	LC	NE
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	x	x	x	x	x	x		x	DD	NT
<i>Lutjanus synagris</i> (Linnaeus, 1758)	x	x	x	x		x		x	NT	NT
<i>Ocyurus chrysurus</i> (Bloch, 1791)		x							DD	NT
Polynemidae										
<i>Polydactylus oligodon</i> (Gunther, 1860)						x			LC	LC
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	x		x	x	x	x	x	x	LC	LC
SCORPAENIFORMES										
Scorpaenidae										

<i>Scorpaena isthmensis</i> Meek & Hilderbrand, 1928	x								LC	LC
<i>Scorpaena plumieri</i> Bloch, 1789	x								LC	LC
Triglidae										
<i>Prionotus punctatus</i> (Bloch, 1793)	x								LC	LC
MORONIFORMES										
Ephippidae										
<i>Chaetodipterus faber</i> (Broussonet, 1782)	x								LC	LC
ACANTHURIFORMES										
Sciaenidae										
<i>Bairdiella ronchus</i> (Cuvier, 1830)	x								LC	LC
<i>Cynoscion acoupa</i> (Lacepède, 1801)	x	x	x	x	x		x	x	LC	NT
<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1833)									LC	LC
<i>Cynoscion leiarchus</i> (Cuvier, 1830)	x	x	x		x		x	x	LC	LC
<i>Cynoscion microlepidotus</i> (Cuvier, 1830)							x		LC	LC
<i>Cynoscion steindachneri</i> (Jordan, 1889)									LC	LC
<i>Cynoscion virescens</i> (Cuvier, 1830)									LC	LC
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)									LC	LC
<i>Larimus breviceps</i> Cuvier, 1830	x	x	x		x		x	x	LC	LC
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)									LC	LC
<i>Menticirrhus americanus</i> (Linnaeus, 1758)	x	x	x	x	x	x	x	x	LC	DD
<i>Menticirrhus littoralis</i> (Holbrook, 1847)									LC	DD
<i>Micropogonias furnieri</i> (Desmarest, 1823)	x	x	x		x		x	x	LC	LC
<i>Nebris microps</i> Cuvier, 1830									LC	LC
<i>Odontoscion dentex</i> (Cuvier, 1830)									LC	LC
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925									LC	DD
<i>Paralanchurus brasiliensis</i> (Steindachner, 1875)	x	x	x	x	x		x	x	LC	LC
<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)									LC	DD
<i>Pogonias cromis</i> (Linnaeus, 1766)									LC	EN

										NE	LC
										LC	LC
										LC	LC
										LC	LC
										DD	LC
										LC	LC
	Acanthuridae										
										LC	LC
										LC	LC
										LC	LC
SPARIFORMES											
	Lobotidae										
										LC	LC
	Sparidae										
										LC	DD
										LC	LC
LOPHIIFORMES											
	Antennariidae										
										LC	DD
										LC	DD
	Ogcocephalidae										
										NE	LC
TETRAODONTIFORMES											
	Ostraciidae										
										LC	LC
										LC	LC
										LC	NE
	Balistidae										

<i>Balistes vetula</i> Linnaeus, 1758										NT	NT
<i>Melichthys niger</i> (Bloch, 1786)										LC	LC
Monacanthidae											
<i>Aluterus heudelotii</i> Hollard, 1855										LC	LC
<i>Aluterus monoceros</i> (Linnaeus, 1758)										LC	NT
<i>Aluterus schoepfii</i> (Walbaum, 1792)										LC	LC
<i>Cantherhines pullus</i> (Ranzani, 1842)										LC	LC
<i>Monacanthus ciliatus</i> (Mitchill, 1818)		x								LC	LC
<i>Stephanolepis hispidus</i> (Linnaeus, 1766)		x								LC	LC
<i>Stephanolepis setifer</i> (Bennett, 1831)										LC	LC
Tetraodontidae											
<i>Canthigaster figueiredoi</i> Moura & Castro, 2002										LC	LC
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	x					x		x	x	LC	LC
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	x	x	x		x	x			x	LC	LC
<i>Sphoeroides greeleyi</i> Gilbert, 1900	x	x			x					LC	LC
<i>Sphoeroides nephelus</i> (Goode & Bean, 1882)										LC	NE
<i>Sphoeroides spengleri</i> (Bloch, 1785)	x	x	x		x					LC	LC
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	x	x	x		x	x	x	x	x	LC	DD
<i>Sphoeroides tyleri</i> Shipp, 1972		x								LC	LC
Diodontidae											
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	x		x	x	x	x	x	x	x	LC	LC
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)		x								LC	LC
<i>Diodon holocanthus</i> Linnaeus, 1758	x		x	x					x	LC	LC
<i>Diodon hystrix</i> Linnaeus, 1758										LC	LC

Table IV: List of fish species recorded in 45 estuaries in north-eastern Brazil, along with their conservation status (ICMbio and IUCN list). Estuaries: Mamanguape River, Miriri River, Paraíba River, Itapessoca, Santa Cruz, Jaguaribe- PE, Suape, Sirinhaém and Maracaípe. ICMbio and IUCN classification: endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

Species	Mamanguape River-PB	Miriri River-PB	Paraíba River-PB	Itapessoca- PE	Santa Cruz-PE	Jaguaribe- PE	Suape-PE	Sirinhaém- PE	Maracaípe- PE	IUCN	ICMBIO
ORECTOLOBIFORMES											
Ginglymostomatidae											
<i>Ginglymostoma cirratum</i> (Bonnaterre, 1788)										VU	VU
CARCHARHINIFORMES											
Carcharhinidae											
<i>Carcharhinus leucas</i> (Muller & Henle, 1839)										NT	NT
<i>Carcharhinus limbatus</i> (Muller & Henle, 1839)										NT	NT
<i>Carcharhinus porosus</i> (Ranzani, 1839)										CR	CR
<i>Isogomphodon oxyrinchus</i> (Muller & Henle, 1839)										CR	CR
<i>Rhizoprionodon lalandii</i> (Muller & Henle, 1839)	x									VU	NT
<i>Rhizoprionodon porosus</i> (Poey, 1861)										VU	DD
Sphyrnidae											
<i>Sphyrna lewini</i> (Griffith & Smith, 1834)										CR	CR
<i>Sphyrna tiburo</i> (Linnaeus, 1758)										EN	CR
<i>Sphyrna tudes</i> (Valenciennes, 1822)										CR	CR
TORPEDINIFORMES											
Narcinidae											
<i>Narcine brasiliensis</i> (Olfers, 1831)						x				NT	DD
PRISTIFORMES											
Pristidae											
<i>Pristis pristis</i> Muller & Henle, 1841										CR	CR

	Rhinobatidae																						
	<i>Pseudobatos percellens</i> (Walbaum, 1792)																			EN	DD		
MYLIOBATIFORMES																							
	Dasyatidae																						
	<i>Fontitrygon geijskesi</i> (Boeseman, 1948)																				CR	DD	
	<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)																				NT	DD	
	<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	x					x		x												NT	LC	
	<i>Hypanus marianae</i> (Gomes, Rosa & Gadig, 2000)																				EN	DD	
	<i>Hypanus say</i> (Lesueur, 1817)																				NT	DD	
	Gymnuridae																						
	<i>Gymnura micrura</i> (Bloch & Scheneider, 1801)	x																			NT	NT	
	Myliobatidae																						
	<i>Aetobatus narinari</i> (Euphrasen, 1790)	x																			EN	DD	
	<i>Mobula hypostoma</i> (Bancroft, 1831)																				EN	VU	
	<i>Rhinoptera bonasus</i> (Mitchill, 1815)	x																			VU	DD	
ELOPIFORMES																							
	Elopidae																						
	<i>Elops saurus</i> Linnaeus, 1766	x					x		x												LC	NE	
	Megalopidae																						
	<i>Megalops atlanticus</i> Valenciennes, 1847	x		x					x												VU	VU	
ALBULIFORMES																							
	Albulidae																						
	<i>Albula vulpes</i> (Linnaeus, 1758)	x							x												x	NT	DD
ANGUILLIFORMES																							
	Muraenidae																						
	<i>Gymnothorax funebris</i> Ranzani, 1839	x		x					x												x	LC	DD
	<i>Gymnothorax moringa</i> (Cuvier, 1829)								x												x	LC	DD
	<i>Gymnothorax ocellatus</i> Agassiz, 1831	x							x													LC	DD

	<i>Gymnothorax vicinus</i> (Castelnau, 1855)	x							LC	DD			
	Ophichthidae												
	<i>Ahlia egmontis</i> (Jordan, 1884)							x	LC	LC			
	<i>Myrichthys breviceps</i> (Richardson, 1848)	x							LC	LC			
	<i>Myrichthys ocellatus</i> (Lesueur, 1825)	x		x			x		x	LC	LC		
	<i>Myrophis punctatus</i> Lütken, 1852					x			x	LC	LC		
	<i>Ophichthus cylindroideus</i> (Ranzani, 1839)	x							x	LC	LC		
	<i>Ophichthus ophis</i> (Linnaeus, 1758)	x							x	LC	LC		
	Muraenesocidae												
	<i>Cynoponticus savanna</i> (Bancroft, 1831)					x				LC	LC		
	Congridae												
	<i>Ariosoma balearicum</i> (Delaroche, 1809)								x	LC	LC		
CLUPEIFORMES													
	Pristigasteridae												
	<i>Chirocentrodon bleekermanus</i> (Poey, 1867)					x				LC	LC		
	<i>Odontognathus mucronatus</i> Lacepède, 1800	x								LC	LC		
	<i>Pellona flavipinnis</i> (Valenciennes, 1837)									LC	LC		
	<i>Pellona harroweri</i> (Fowler, 1917)	x								LC	LC		
	Engraulidae												
	<i>Anchoa filifera</i> (Fowler, 1915)					x			x	LC	LC		
	<i>Anchoa hepsetus</i> (Linnaeus, 1758)									LC	NE		
	<i>Anchoa januaria</i> (Steindachner, 1879)	x		x		x			x	x	LC	LC	
	<i>Anchoa lyolepis</i> (Evermann & Marsh, 1900)				x	x					LC	LC	
	<i>Anchoa parva</i> (Meek & Hildebrand, 1923)					x					LC	NE	
	<i>Anchoa spinifer</i> (Valenciennes, 1848)	x		x		x			x	x	x	LC	LC
	<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)	x				x			x	x	x	LC	LC
	<i>Anchovia clupeioides</i> (Swainson, 1839)	x		x		x			x	x	x	LC	LC
	<i>Anchoviella lepidentostole</i> (Fowler, 1911)							x			x	LC	LC

<i>Cetengraulis edentulus</i> (Cuvier, 1829)	x	x			x		x	x	x	LC	LC
<i>Engraulis anchoita</i> Hubbs & Marini 1935					x					LC	LC
<i>Lycengraulis batesii</i> (Gunther, 1868)										LC	LC
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	x	x	x	x	x	x	x	x	x	LC	LC
<i>Pterengraulis atherinoides</i> (Linnaeus, 1766)										LC	LC
Clupeidae											
<i>Harengula clupeola</i> (Cuvier, 1829)	x			x	x	x	x	x	x	LC	LC
<i>Harengula jaguana</i> Poey, 1865								x		LC	LC
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)				x	x	x	x	x	x	LC	LC
<i>Opisthonema oglinum</i> (Lesueur, 1818)	x		x		x	x	x	x		LC	LC
<i>Platanichthys platana</i> (Regan, 1917)					x			x		LC	LC
<i>Rhinosardinia amazonica</i> (Steindachner, 1879)	x	x								LC	LC
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)										LC	LC
CHARACIFORMES											
Erythrinidae											
<i>Hoplias malabaricus</i> (Bloch, 1794)										LC	LC
<i>Prochilodus argenteus</i> Spix & Agassiz, 1829										NE	LC
Serrasalminidae											
<i>Pygocentrus nattereri</i> Kner, 1858										NE	LC
<i>Serrasalmus rhombeus</i> (Linnaeus, 1766)										NE	LC
<i>Serrasalmus spilopleura</i> Kner, 1858										NE	LC
Anostomidae											
<i>Schizodon fasciatus</i> Spix & Agassiz, 1829										NE	LC
<i>Leporinus friderici</i> (Bloch, 1794)										NE	LC
Curimatidae											
<i>Steindachnerina elegans</i> (Steindachner, 1875)										NE	LC
Characidae											
<i>Astyanax bimaculatus</i> (Linnaeus, 1758)										NE	LC

												NE	LC
SILURIFORMES	<i>Hemigrammus marginatus</i> Ellis, 1911												
	Loricariidae												
	<i>Hypostomus plecostomus</i> (Linnaeus, 1758)											NE	NE
	<i>Loricaria parnahybae</i> Steindachner, 1907											NE	LC
	Ariidae												
	<i>Aspistor luniscutis</i> (Valenciennes, 1840)					x					x	NE	LC
	<i>Aspistor quadriscutis</i> (Valenciennes, 1840)	x		x							x	LC	LC
	<i>Bagre bagre</i> (Linnaeus, 1766)											LC	NT
	<i>Bagre marinus</i> (Mitchill, 1815)	x				x						LC	DD
	<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)					x					x	NE	LC
	<i>Cathorops spixii</i> (Agassiz, 1829)	x		x		x		x			x	NE	LC
	<i>Genidens genidens</i> (Cuvier, 1829)	x										LC	LC
	<i>Notarius grandicassis</i> (Valenciennes, 1840)										x	LC	LC
	<i>Sciades couma</i> (Valenciennes, 1840)											LC	DD
	<i>Sciades herzbergii</i> (Bloch, 1794)	x		x		x				x	x	LC	LC
	<i>Sciades parkeri</i> (Traill, 1832)					x					x	VU	VU
	<i>Sciades proops</i> (Valenciennes, 1840)	x				x						NE	DD
	Aspredinidae												
	<i>Aspredo aspredo</i> (Linnaeus, 1758)											LC	LC
	<i>Aspredinichthys tибicen</i> (Valenciennes, 1840)											LC	LC
	Auchenipteridae												
	<i>Ageneiosus inermis</i> (Linnaeus, 1766)											NE	LC
	<i>Pseudauchenipterus nodosus</i> (Bloch, 1794)											NE	LC
	<i>Trachelyopterus galeatus</i> (Linnaeus, 1766)											NE	LC
	Heptapteridae												
	<i>Pimelodella cristata</i> (Muller & Troschel, 1849)											LC	LC
	<i>Pimelodella lateristriga</i> (Lichtenstein, 1823)											NE	DD

	Pimelodidae											
	<i>Brachyplatystoma vaillantii</i> (Valenciennes, 1840)									NE	LC	
	<i>Pseudoplatystoma fasciatum</i> (Linnaeus, 1766)									NE	NE	
	<i>Sorubim lima</i> (Bloch & Schneider, 1801)									NE	LC	
AULOPIFORMES												
	Synodontidae											
	<i>Synodus foetens</i> (Linnaeus, 1766)					x		x		x	LC	LC
	<i>Synodus poeyi</i> Jordan, 1887					x					LC	LC
	<i>Trachinocephalus myops</i> (Forster, 1801)										LC	LC
HOLOCENTRIFORMES												
	Holocentridae											
	<i>Holocentrus adscensionis</i> (Osbeck, 1765)							x			LC	LC
BATRACHOIDIFORMES												
	Batrachoididae											
	<i>Amphichthys cryptocentrus</i> (Valenciennes, 1837)					x					LC	LC
	<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)					x					LC	LC
	<i>Thalassophryne nattereri</i> Steindachner, 1876	x	x		x	x	x	x		x	LC	LC
	<i>Thalassophryne punctata</i> Steindachner, 1876	x									NE	LC
KURTIFORMES												
	Apogonidae											
	<i>Phaeoptyx pigmentaria</i> (Poey, 1860)							x			LC	LC
GOBIIFORMES												
	Eleotridae											
	<i>Dormitator maculatus</i> (Bloch, 1792)					x		x			LC	NT
	<i>Eleotris pisonis</i> (Gmelin, 1789)	x	x			x					LC	LC
	<i>Erotelis smaragdus</i> (Valenciennes, 1837)					x			x		LC	LC
	<i>Guavina guavina</i> (Valenciennes, 1837)				x	x	x			x	LC	LC
	Gobiidae											

	<i>Awaous tajasica</i> (Lichtenstein, 1822)									LC	LC
	<i>Bathygobius mystacium</i> Ginsburg, 1947									LC	NE
	<i>Bathygobius soporator</i> (Valenciennes, 1837)	x	x	x	x	x	x		x	LC	LC
	<i>Coryphopterus glaucofraenum</i> Gill, 1863	x								LC	LC
	<i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882)	x	x	x	x	x	x	x	x	LC	LC
	<i>Ctenogobius saepepallens</i> (Gilbert & Randall, 1968)								x	LC	LC
	<i>Ctenogobius shufeldti</i> (Jordan & Eigenmann, 1887)				x					LC	LC
	<i>Ctenogobius smaragdus</i> (Valenciennes, 1837)	x			x	x	x			LC	LC
	<i>Ctenogobius stigmaticus</i> (Poey, 1860)				x					LC	LC
	<i>Evorthodus lyricus</i> (Girard, 1858)				x					LC	LC
	<i>Gobioides broussonneti</i> Lacepède, 1800								x	LC	LC
	<i>Gobionellus oceanicus</i> (Pallas, 1770)	x	x		x	x	x	x	x	LC	LC
	<i>Gobionellus stomatus</i> Starks, 1913		x		x	x	x			NE	LC
	<i>Microdesmus bahianus</i> Dawson, 1973									LC	LC
	<i>Microdesmus longipinnis</i> (Weymouth, 1910)									LC	LC
	<i>Microgobius meeki</i> Evermann & Marsh, 1899				x					LC	LC
	Pomacentridae										
	<i>Abudefduf saxatilis</i> (Linnaeus, 1758)	x			x			x		LC	LC
	<i>Stegastes fuscus</i> (Cuvier, 1830)	x								LC	LC
	<i>Stegastes variabilis</i> (Castelnau, 1855)	x			x			x		NE	LC
	Opistognathidae										
	<i>Opistognathus cuvierii</i> Valenciennes, 1836									NE	LC
MUGILIFORMES											
	Mugilidae										
	<i>Mugil brevirostris</i> (Ribeiro, 1915)							x		NE	NE
	<i>Mugil curema</i> Valenciennes, 1836	x	x		x	x	x	x	x	LC	DD
	<i>Mugil curvidens</i> Valenciennes, 1836	x	x		x		x	x	x	LC	DD
	<i>Mugil incilis</i> Hancock, 1830						x	x	x	LC	LC

	<i>Mugil liza</i> Valenciennes, 1836					x	x	x	x	x	DD	NT
	<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007					x		x			LC	DD
CICHLIFORMES												
	Cichlidae											
	<i>Cichla ocellaris</i> Bloch & Schneider, 1801										NE	LC
	<i>Oreochromis niloticus</i> (Linnaeus, 1758)										LC	NE
BLENNIIFORMES												
	Dactyloscopidae											
	<i>Dactyloscopus crossotus</i> Starks, 1913									x	LC	LC
	Blenniidae											
	<i>Scartella cristata</i> (Linnaeus, 1758)							x			LC	LC
	Labrisomidae											
	<i>Labrisomus nuchipinnis</i> (Quoy & Gaimard, 1824)				x			x			LC	LC
ATHERINIFORMES												
	Atherinidae											
	<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)	x	x	x	x	x	x	x	x	x	LC	LC
	<i>Atherinella cf. blackburni</i> (Schultz, 1949)									x	LC	LC
	<i>Odontesthes bonariensis</i> (Valenciennes, 1835)										NE	DD
BELONIFORMES												
	Exocoetidae											
	<i>Cheilopogon melanurus</i> (Valenciennes, 1847)										LC	LC
	<i>Hirundichthys affinis</i> (Gunther, 1866)										LC	LC
	Hemiramphidae											
	<i>Hemiramphus balao</i> Lesueur, 1821	x				x		x	x		LC	DD
	<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)	x				x	x	x			LC	LC
	<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)	x	x		x	x	x	x	x	x	LC	LC
	<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)										LC	NT
	Belonidae											

									x	LC	LC		
	<i>Ablennes hians</i> (Valenciennes, 1846)								x	LC	LC		
	<i>Strongylura marina</i> (Walbaum, 1792)			x					x	LC	LC		
	<i>Strongylura timucu</i> (Walbaum, 1792)	x	x		x		x	x	x	LC	LC		
	<i>Tylosurus acus acus</i> (Lacepède, 1803)								x	LC	LC		
	<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)								x	LC	LC		
CYPRINODONTIFORMES													
	Poeciliidae												
	<i>Poecilia vivipara</i> Bloch & Schneider, 1801				x		x		x		NE	LC	
CARANGIFORMES													
	Rachycentridae												
	<i>Rachycentron canadum</i> (Linnaeus, 1766)	x									LC	LC	
	Echeneidae												
	<i>Echeneis naucrates</i> Linnaeus, 1758	x	x					x			LC	LC	
	<i>Phtheichthys lineatus</i> (Menziés, 1791)										LC	LC	
	<i>Remora remora</i> (Linnaeus, 1758)										LC	LC	
	Carangidae												
	<i>Carangoides bartholomaei</i> (Cuvier, 1833)	x							x	x	x	LC	LC
	<i>Caranx crysos</i> (Mitchill, 1815)				x		x	x		x		LC	LC
	<i>Caranx hippos</i> (Linnaeus, 1766)				x		x	x	x	x	x	LC	LC
	<i>Caranx latus</i> Agassiz, 1831	x	x				x	x	x	x	x	LC	LC
	<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	x			x					x		LC	LC
	<i>Hemicaranx amblyrhynchus</i> (Cuvier, 1833)											LC	LC
	<i>Oligoplites palometa</i> (Cuvier, 1832)	x	x	x	x		x	x		x		LC	LC
	<i>Oligoplites saliens</i> (Bloch, 1793)				x					x		LC	LC
	<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)				x		x	x	x	x	x	LC	LC
	<i>Selene brownii</i> (Curvier, 1816)											LC	LC
	<i>Selene setapinnis</i> (Mitchill, 1815)						x		x			LC	LC
	<i>Selene vomer</i> (Linnaeus, 1758)	x			x		x	x	x	x		LC	LC

	<i>Trachinotus carolinus</i> (Linnaeus, 1766)								LC	LC	
	<i>Trachinotus cayennensis</i> Cuvier, 1832								LC	DD	
	<i>Trachinotus falcatus</i> (Linnaeus, 1758)	x			x		x	x	LC	LC	
	<i>Trachinotus goodei</i> Jordan & Evermann, 1896	x							LC	LC	
ISTIOPHORIFORMES											
	Sphyraenidae										
	<i>Sphyraena barracuda</i> (Edwards, 1771)	x		x	x	x	x	x	x	LC	LC
	<i>Sphyraena guachancho</i> Cuvier, 1829				x		x		x	LC	LC
PLEURONECTIFORMES											
	Paralichthyidae										
	<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900	x	x			x	x		x	LC	LC
	<i>Citharichthys cornutus</i> (Günther, 1880)									LC	LC
	<i>Citharichthys macrops</i> Dresel, 1885				x				x	LC	LC
	<i>Citharichthys spilopterus</i> Günther, 1862	x	x			x		x	x	LC	LC
	<i>Etropus crossotus</i> Jordan & Gilbert, 1882	x				x		x		LC	LC
	<i>Etropus longimanus</i> Norman, 1933					x				NE	LC
	<i>Paralichthys brasiliensis</i> (Ranzani, 1842)					x	x	x		NE	LC
	<i>Paralichthys orbignyanus</i> (Valenciennes, 1839)					x				DD	DD
	<i>Paralichthys tropicus</i> Ginsburg, 1933									DD	NE
	<i>Syacium micrurum</i> Ranzani, 1842	x				x		x		LC	LC
	<i>Syacium papillosum</i> (Linnaeus, 1758)									LC	LC
	Bothidae										
	<i>Bothus lunatus</i> (Linnaeus, 1758)									LC	LC
	<i>Bothus ocellatus</i> (Agassiz, 1831)	x		x	x			x		LC	LC
	<i>Bothus robinsi</i> Topp & Hoff, 1972								x	LC	LC
	Achiridae										
	<i>Achirus achirus</i> (Linnaeus, 1758)					x			x	LC	LC
	<i>Achirus declivis</i> Chabanaud, 1940	x				x			x	LC	LC

	<i>Achirus lineatus</i> (Linnaeus, 1758)	x	x	x	x	x	x	x	x	x	LC	LC
	<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009								x		NE	VU
	<i>Trinectes microphthalmus</i> (Chabanaud, 1928)	x							x		LC	LC
	<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	x	x			x			x	x	LC	LC
	Cynoglossidae											
	<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)								x		LC	LC
	<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)			x		x	x				LC	LC
	<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)	x				x			x	x	LC	LC
SYNGNATHIFORMES												
	Syngnathidae											
	<i>Bryx dunckeri</i> (Metzelaar, 1919)					x					LC	LC
	<i>Cosmocampus elucens</i> (Poey, 1868)			x	x	x	x				LC	LC
	<i>Hippocampus erectus</i> Perry, 1810					x			x		VU	VU
	<i>Hippocampus reidi</i> Ginsburg, 1933	x	x			x					NT	VU
	<i>Microphis brachyurus lineatus</i> (Kaup, 1856)			x			x				NE	NE
	<i>Microphis brachyurus brachyurus</i> (Bleeker, 1854)	x									LC	LC
	Fistulariidae											
	<i>Fistularia petimba</i> Lacepède, 1803						x	x			LC	LC
	<i>Fistularia tabacaria</i> Linnaeus, 1758					x	x	x			LC	LC
	<i>Syngnathus pelagicus</i> Linnaeus, 1758			x			x	x			LC	LC
	Dactylopteridae											
	<i>Dactylopterus volitans</i> (Linnaeus, 1758)	x				x	x		x	x	LC	LC
SCOMBRIFORMES												
	Trichiuridae											
	<i>Trichiurus lepturus</i> Linnaeus, 1758	x				x				x	LC	LC
	Scombridae											
	<i>Euthynnus alletteratus</i> (Rafinesque, 1810)	x									LC	LC

	<i>Scomberomorus brasiliensis</i> Collette, Russo & Zavala-Camin, 1978	x			x					LC	LC
	<i>Scomberomorus cavalla</i> (Cuvier, 1829)	x								LC	LC
	<i>Scomberomorus maculatus</i> (Mitchill, 1815)									LC	NE
	<i>Scomberomorus regalis</i> (Bloch, 1793)				x					LC	LC
	Stromateidae										
	<i>Peprilus paru</i> (Linnaeus, 1758)	x			x			x		LC	LC
TRACHINIFORMES											
	Uranoscopidae										
	<i>Astroscopus y-graecum</i> Cuvier, 1829								x	LC	LC
LABRIFORMES											
	Labridae										
	<i>Halichoeres poeyi</i> (Steindachner, 1867)	x								LC	LC
	Scaridae										
	<i>Nicholsina usta</i> (Valenciennes, 1840)									LC	LC
	<i>Sparisoma amplum</i> (Ranzani, 1842)						x		x	LC	NT
	<i>Sparisoma axillare</i> (Steindachner, 1878)	x			x				x	DD	VU
	<i>Sparisoma radians</i> (Valenciennes, 1840)			x	x				x	LC	LC
PERCIFORMES											
	Centropomidae										
	<i>Centropomus ensiferus</i> Poey, 1860	x							x	LC	LC
	<i>Centropomus mexicanus</i> Bocourt, 1868	x	x						x	LC	LC
	<i>Centropomus parallelus</i> Poey, 1860			x	x	x	x	x	x	LC	LC
	<i>Centropomus pectinatus</i> Poey, 1860	x	x		x				x	LC	LC
	<i>Centropomus undecimalis</i> (Bloch, 1792)	x	x	x	x	x	x	x	x	LC	LC
	Gerreidae										
	<i>Diapterus auratus</i> Ranzani, 1842	x			x	x	x	x	x	LC	LC
	<i>Diapterus rhombeus</i> (Cuvier, 1829)	x	x	x	x	x	x	x	x	LC	LC

<i>Eucinostomus argenteus</i> Baird & Girard, 1855	x		x	x	x	x	x	x	LC	LC
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	x		x	x	x	x	x	x	LC	LC
<i>Eucinostomus havana</i> (Nichols, 1912)				x		x		x	LC	LC
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	x	x	x	x	x	x	x	x	LC	LC
<i>Eugerres brasiliensis</i> (Cuvier, 1830)	x	x	x	x	x	x	x	x	LC	LC
<i>Gerres cinereus</i> (Walbaum, 1792)				x				x	LC	LC
<i>Ulaema lefroyi</i> (Goode, 1874)				x	x	x		x	LC	LC
Mullidae										
<i>Pseudupeneus maculatus</i> (Bloch, 1793)				x		x		x	LC	LC
Serranidae										
<i>Alphestes afer</i> (Bloch, 1793)									LC	DD
<i>Diplectrum radiale</i> (Quoy & Gaimard, 1824)						x			LC	LC
<i>Epinephelus adscensionis</i> (Osbeck, 1765)	x			x		x		x	LC	DD
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	x			x		x			VU	CR
<i>Hyporthodus nigritus</i> (Holbrook, 1855)									NT	EN
<i>Mycteroperca bonaci</i> (Poey, 1860)				x		x			NT	VU
<i>Mycteroperca tigris</i> (Valenciennes, 1833)									DD	DD
<i>Mycteroperca venenosa</i> (Linnaeus, 1758)									NT	DD
<i>Rypticus randalli</i> Courtenay, 1967	x	x		x				x	LC	LC
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)						x		x	LC	LC
<i>Serranus flaviventris</i> (Cuvier, 1829)	x					x			LC	LC
<i>Serranus phoebe</i> Poey, 1851									LC	LC
Pomacanthidae										
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)									LC	DD
<i>Pomacanthus paru</i> (Bloch, 1787)				x				x	LC	DD
Chaetodontidae										
<i>Chaetodon ocellatus</i> (Bloch, 1787)				x					LC	DD
<i>Chaetodon striatus</i> Linnaeus, 1758	x			x		x			LC	LC

Haemulidae											
<i>Anisotremus surinamensis</i> (Bloch, 1791)	x								DD	DD	
<i>Anisotremus virginicus</i> (Linnaeus, 1758)	x			x					LC	LC	
<i>Conodon nobilis</i> (Linnaeus, 1758)	x			x			x		LC	LC	
<i>Genyatremus luteus</i> (Bloch, 1790)				x			x		NE	LC	
<i>Haemulon aurolineatum</i> Cuvier, 1830				x			x		x	LC	LC
<i>Haemulon parra</i> (Desmarest, 1823)	x			x			x			LC	LC
<i>Haemulon plumierii</i> (Lacepède, 1801)	x						x			LC	DD
<i>Haemulon squamipinna</i> Rocha & Rosa, 1999										NE	LC
<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)	x					x			x	LC	LC
<i>Orthopristis ruber</i> (Cuvier, 1830)				x						LC	LC
<i>Pomadasys corvinaeformis</i> (Steindachner, 1868)	x		x	x	x		x	x	x	LC	LC
<i>Pomadasys crocro</i> (Cuvier, 1830)				x	x		x		x	DD	LC
<i>Pomadasys ramosus</i> Poey, 1860					x					NE	LC
Lutjanidae											
<i>Lutjanus alexandrei</i> Moura & Linderman, 2007	x			x	x	x	x	x	x	NE	LC
<i>Lutjanus analis</i> (Cuvier, 1828)	x		x	x			x			NT	NT
<i>Lutjanus apodus</i> (Walbaum, 1792)	x			x	x					LC	NE
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	x	x				x	x	x	x	VU	VU
<i>Lutjanus griseus</i> (Linnaeus, 1758)				x	x					LC	NE
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	x	x		x	x	x	x	x	x	DD	NT
<i>Lutjanus synagris</i> (Linnaeus, 1758)	x		x	x	x	x	x			NT	NT
<i>Ocyurus chrysurus</i> (Bloch, 1791)	x			x			x			DD	NT
Polynemidae											
<i>Polydactylus oligodon</i> (Gunther, 1860)										LC	LC
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	x	x	x	x			x	x	x	LC	LC
SCORPAENIFORMES											
Scorpaenidae											

	<i>Scorpaena isthmensis</i> Meek & Hilderbrand, 1928									LC	LC
	<i>Scorpaena plumieri</i> Bloch, 1789	x				x			x	LC	LC
	Triglidae										
	<i>Prionotus punctatus</i> (Bloch, 1793)	x				x	x		x	LC	LC
MORONIFORMES											
	Ephippidae										
	<i>Chaetodipterus faber</i> (Broussonet, 1782)	x	x	x	x	x	x	x	x	LC	LC
ACANTHURIFORMES											
	Sciaenidae										
	<i>Bairdiella ronchus</i> (Cuvier, 1830)	x	x			x	x	x	x	LC	LC
	<i>Cynoscion acoupa</i> (Lacepède, 1801)	x				x			x	LC	NT
	<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1833)									LC	LC
	<i>Cynoscion leiarchus</i> (Cuvier, 1830)	x				x				LC	LC
	<i>Cynoscion microlepidotus</i> (Cuvier, 1830)	x								LC	LC
	<i>Cynoscion steindachneri</i> (Jordan, 1889)									LC	LC
	<i>Cynoscion virescens</i> (Cuvier, 1830)					x				LC	LC
	<i>Isopisthus parvipinnis</i> (Cuvier, 1830)		x			x			x	LC	LC
	<i>Larimus breviceps</i> Cuvier, 1830	x	x							LC	LC
	<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)					x				LC	LC
	<i>Menticirrhus americanus</i> (Linnaeus, 1758)	x	x	x		x	x		x	LC	DD
	<i>Menticirrhus littoralis</i> (Holbrook, 1847)	x		x						LC	DD
	<i>Micropogonias furnieri</i> (Desmarest, 1823)	x	x			x			x	LC	LC
	<i>Nebris microps</i> Cuvier, 1830	x	x							LC	LC
	<i>Odontoscion dentex</i> (Cuvier, 1830)									LC	LC
	<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925	x	x						x	LC	DD
	<i>Paralanchurus brasiliensis</i> (Steindachner, 1875)									LC	LC
	<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)	x								LC	DD
	<i>Pogonias cromis</i> (Linnaeus, 1766)	x								LC	EN

	<i>Stellifer brasiliensis</i> (Schultz, 1945)	x		x						NE	LC	
	<i>Stellifer microps</i> (Steindachner, 1864)								x	LC	LC	
	<i>Stellifer naso</i> (Jordan, 1889)									LC	LC	
	<i>Stellifer rastrifer</i> (Jordan, 1889)	x							x	LC	LC	
	<i>Stellifer stellifer</i> (Bloch, 1790)									DD	LC	
	<i>Umbrina coroides</i> Cuvier, 1830									LC	LC	
	Acanthuridae											
	<i>Acanthurus bahianus</i> Castelnau, 1855	x							x		LC	LC
	<i>Acanthurus chirurgus</i> (Bloch, 1787)	x							x		LC	LC
	<i>Acanthurus coeruleus</i> Bloch & Schneider, 1801	x									LC	LC
SPARIFORMES												
	Lobotidae											
	<i>Lobotes surinamensis</i> (Bloch, 1790)										LC	LC
	Sparidae											
	<i>Archosargus probatocephalus</i> (Walbaum, 1792)										LC	DD
	<i>Archosargus rhomboidalis</i> (Linnaeus, 1758)	x									LC	LC
LOPHIIFORMES												
	Antennariidae											
	<i>Antennarius multiocellatus</i> (Valenciennes, 1837)										LC	DD
	<i>Antennarius striatus</i> (Shaw, 1794)										LC	DD
	Ogcocephalidae											
	<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	x									NE	LC
TETRAODONTIFORMES												
	Ostraciidae											
	<i>Acanthostracion quadricornis</i> (Linnaeus, 1758)										LC	LC
	<i>Lactophrys trigonus</i> (Linnaeus, 1758)										LC	LC
	<i>Lactophrys triqueter</i> (Linnaeus, 1758)										LC	NE
	Balistidae											

<i>Balistes vetula</i> Linnaeus, 1758									x	NT	NT
<i>Melichthys niger</i> (Bloch, 1786)										LC	LC
Monacanthidae											
<i>Aluterus heudelotii</i> Hollard, 1855										LC	LC
<i>Aluterus monoceros</i> (Linnaeus, 1758)										LC	NT
<i>Aluterus schoepfii</i> (Walbaum, 1792)						x				LC	LC
<i>Cantherhines pullus</i> (Ranzani, 1842)						x				LC	LC
<i>Monacanthus ciliatus</i> (Mitchill, 1818)										LC	LC
<i>Stephanolepis hispidus</i> (Linnaeus, 1766)										LC	LC
<i>Stephanolepis setifer</i> (Bennett, 1831)										LC	LC
Tetraodontidae											
<i>Canthigaster figueiredoi</i> Moura & Castro, 2002						x				LC	LC
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	x		x			x	x			LC	LC
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	x					x	x	x	x	LC	LC
<i>Sphoeroides greeleyi</i> Gilbert, 1900	x		x			x	x	x	x	LC	LC
<i>Sphoeroides nephelus</i> (Goode & Bean, 1882)										LC	NE
<i>Sphoeroides spengleri</i> (Bloch, 1785)					x	x		x		LC	LC
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	x	x		x		x	x	x	x	LC	DD
<i>Sphoeroides tyleri</i> Shipp, 1972										LC	LC
Diodontidae											
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	x							x		LC	LC
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)						x	x			LC	LC
<i>Diodon holocanthus</i> Linnaeus, 1758						x				LC	LC
<i>Diodon hystrix</i> Linnaeus, 1758						x				LC	LC

Table V: List of fish species recorded in 45 estuaries in north-eastern Brazil, along with their conservation status (ICMbio and IUCN list). Estuaries: Formoso River, Santo Antônio River, Mundaú River – AL, São Francisco River, Japaratuba, Paraguaçu River, Contas River, Itaparica Island and Joanes River. ICMbio and IUCN classification: endangered (EN), vulnerable (VU), near threatened (NT), least concern (LC), data deficient (DD) and not evaluated (NE).

Species	Formoso	Santo	Mundaú	São	Japaratuba-	Paraguaçu	Contas	Itaparica	Joanes	IUCN	ICMBIO
	River-PE	Antônio River-AL	River-AL	Francisco River-SE	SE	River-BA	River-BA	Island-BA	River-BA		
ORECTOLOBIFORMES											
Ginglymostomatidae											
<i>Ginglymostoma cirratum</i> (Bonnaterre, 1788)										VU	VU
CARCHARHINIFORMES											
Carcharhinidae											
<i>Carcharhinus leucas</i> (Muller & Henle, 1839)										NT	NT
<i>Carcharhinus limbatus</i> (Muller & Henle, 1839)										NT	NT
<i>Carcharhinus porosus</i> (Ranzani, 1839)										CR	CR
<i>Isogomphodon oxyrinchus</i> (Muller & Henle, 1839)										CR	CR
<i>Rhizoprionodon lalandii</i> (Muller & Henle, 1839)										VU	NT
<i>Rhizoprionodon porosus</i> (Poey, 1861)							x			VU	DD
Sphyrnidae											
<i>Sphyrna lewini</i> (Griffith & Smith, 1834)										CR	CR
<i>Sphyrna tiburo</i> (Linnaeus, 1758)										EN	CR
<i>Sphyrna tudes</i> (Valenciennes, 1822)										CR	CR
TORPEDINIFORMES											
Narcinidae											
<i>Narcine brasiliensis</i> (Olfers, 1831)							x		x	NT	DD
PRISTIFORMES											
Pristidae											

									CR	CR
	<i>Pristis pristis</i> Muller & Henle, 1841									
	Rhinobatidae									
	<i>Pseudobatos percellens</i> (Walbaum, 1792)						x		EN	DD
MYLIOBATIFORMES										
	Dasyatidae									
	<i>Fontitrygon geijskesi</i> (Boeseman, 1948)								CR	DD
	<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)	x					x		NT	DD
	<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	x					x		NT	LC
	<i>Hypanus marianae</i> (Gomes, Rosa & Gadig, 2000)								EN	DD
	<i>Hypanus say</i> (Lesueur, 1817)								NT	DD
	Gymnuridae									
	<i>Gymnura micrura</i> (Bloch & Scheneider, 1801)						x		NT	NT
	Myliobatidae									
	<i>Aetobatus narinari</i> (Euphrasen, 1790)								EN	DD
	<i>Mobula hypostoma</i> (Bancroft, 1831)								EN	VU
	<i>Rhinoptera bonasus</i> (Mitchill, 1815)						x		VU	DD
ELOPIFORMES										
	Elopidae									
	<i>Elops saurus</i> Linnaeus, 1766	x		x			x		LC	NE
	Megalopidae									
	<i>Megalops atlanticus</i> Valenciennes, 1847	x					x		VU	VU
ALBULIFORMES										
	Albulidae									
	<i>Albula vulpes</i> (Linnaeus, 1758)	x	x		x			x	NT	DD
ANGUILLIFORMES										
	Muraenidae									
	<i>Gymnothorax funebris</i> Ranzani, 1839	x							LC	DD
	<i>Gymnothorax moringa</i> (Cuvier, 1829)						x	x	LC	DD

	<i>Gymnothorax ocellatus</i> Agassiz, 1831	x			x		x	LC	DD	
	<i>Gymnothorax vicinus</i> (Castelnau, 1855)							LC	DD	
	Ophichthidae									
	<i>Ahlia egmontis</i> (Jordan, 1884)							LC	LC	
	<i>Myrichthys breviceps</i> (Richardson, 1848)							LC	LC	
	<i>Myrichthys ocellatus</i> (Lesueur, 1825)					x		LC	LC	
	<i>Myrophis punctatus</i> Lütken, 1852							LC	LC	
	<i>Ophichthus cylindroideus</i> (Ranzani, 1839)							LC	LC	
	<i>Ophichthus ophis</i> (Linnaeus, 1758)						x	LC	LC	
	Muraenesocidae									
	<i>Cynoponticus savanna</i> (Bancroft, 1831)							LC	LC	
	Congridae									
	<i>Ariosoma balearicum</i> (Delaroche, 1809)							LC	LC	
CLUPEIFORMES										
	Pristigasteridae									
	<i>Chirocentrodon bleekermanus</i> (Poey, 1867)							LC	LC	
	<i>Odontognathus mucronatus</i> Lacepède, 1800							LC	LC	
	<i>Pellona flavipinnis</i> (Valenciennes, 1837)							LC	LC	
	<i>Pellona harroweri</i> (Fowler, 1917)					x		LC	LC	
	Engraulidae									
	<i>Anchoa filifera</i> (Fowler, 1915)							x	LC	LC
	<i>Anchoa hepsetus</i> (Linnaeus, 1758)							LC	NE	
	<i>Anchoa januaria</i> (Steindachner, 1879)					x		LC	LC	
	<i>Anchoa lyolepis</i> (Evermann & Marsh, 1900)					x		LC	LC	
	<i>Anchoa parva</i> (Meek & Hildebrand, 1923)							LC	NE	
	<i>Anchoa spinifer</i> (Valenciennes, 1848)					x		LC	LC	
	<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)	x	x					LC	LC	
	<i>Anchovia clupeoides</i> (Swainson, 1839)	x	x	x		x	x	x	LC	LC

<i>Anchoiella lepidentostole</i> (Fowler, 1911)					x			x	LC	LC
<i>Cetengraulis edentulus</i> (Cuvier, 1829)	x		x	x			x		LC	LC
<i>Engraulis anchoita</i> Hubbs & Marini 1935									LC	LC
<i>Lycengraulis batesii</i> (Gunther, 1868)									LC	LC
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	x				x	x		x	LC	LC
<i>Pterengraulis atherinoides</i> (Linnaeus, 1766)									LC	LC
Clupeidae										
<i>Harengula clupeola</i> (Cuvier, 1829)	x	x			x	x			LC	LC
<i>Harengula jaguana</i> Poey, 1865								x	LC	LC
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)	x					x		x	LC	LC
<i>Opisthonema oglinum</i> (Lesueur, 1818)	x	x				x		x	LC	LC
<i>Platanichthys platana</i> (Regan, 1917)	x								LC	LC
<i>Rhinosardinia amazonica</i> (Steindachner, 1879)									LC	LC
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)									LC	LC
CHARACIFORMES										
Erythrinidae										
<i>Hoplias malabaricus</i> (Bloch, 1794)									LC	LC
<i>Prochilodus argenteus</i> Spix & Agassiz, 1829					x				NE	LC
Serrasalminidae										
<i>Pygocentrus nattereri</i> Kner, 1858									NE	LC
<i>Serrasalmus rhombeus</i> (Linnaeus, 1766)									NE	LC
<i>Serrasalmus spilopleura</i> Kner, 1858									NE	LC
Anostomidae										
<i>Schizodon fasciatus</i> Spix & Agassiz, 1829									NE	LC
<i>Leporinus friderici</i> (Bloch, 1794)									NE	LC
Curimatidae										
<i>Steindachnerina elegans</i> (Steindachner, 1875)									NE	LC
Characidae										

	<i>Pimelodella lateristriga</i> (Lichtenstein, 1823)							NE	DD
	Pimelodidae								
	<i>Brachyplatystoma vaillantii</i> (Valenciennes, 1840)							NE	LC
	<i>Pseudoplatystoma fasciatum</i> (Linnaeus, 1766)							NE	NE
	<i>Sorubim lima</i> (Bloch & Schneider, 1801)							NE	LC
AULOPIFORMES									
	Synodontidae								
	<i>Synodus foetens</i> (Linnaeus, 1766)	x			x		x	LC	LC
	<i>Synodus poeyi</i> Jordan, 1887							LC	LC
	<i>Trachinocephalus myops</i> (Forster, 1801)						x	LC	LC
HOLOCENTRIFORMES									
	Holocentridae								
	<i>Holocentrus adscensionis</i> (Osbeck, 1765)							LC	LC
BATRACHOIDIFORMES									
	Batrachoididae								
	<i>Amphichthys cryptocentrus</i> (Valenciennes, 1837)				x			LC	LC
	<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)							LC	LC
	<i>Thalassophryne nattereri</i> Steindachner, 1876	x		x	x		x	LC	LC
	<i>Thalassophryne punctata</i> Steindachner, 1876						x	NE	LC
KURTIFORMES									
	Apogonidae								
	<i>Phaeoptyx pigmentaria</i> (Poey, 1860)							LC	LC
GOBIIFORMES									
	Eleotridae								
	<i>Dormitator maculatus</i> (Bloch, 1792)	x						LC	NT
	<i>Eleotris pisonis</i> (Gmelin, 1789)	x					x	LC	LC
	<i>Erotelis smaragdus</i> (Valenciennes, 1837)							LC	LC
	<i>Guavina guavina</i> (Valenciennes, 1837)	x						LC	LC

Gobiidae									
								LC	LC
								LC	NE
	x	x			x		x	LC	LC
								LC	LC
					x		x	LC	LC
								LC	LC
	x				x		x	LC	LC
					x		x	LC	LC
								LC	LC
		x						LC	LC
	x				x			LC	LC
	x				x		x	NE	LC
	x							LC	LC
	x							LC	LC
					x			LC	LC
Pomacentridae									
							x	LC	LC
	x							LC	LC
							x	NE	LC
Opistognathidae									
					x			NE	LC
MUGILIFORMES									
Mugilidae									
		x						NE	NE
	x	x	x		x	x	x	LC	DD
	x	x	x	x			x	LC	DD

	<i>Mugil incilis</i> Hancock, 1830				x			x	LC	LC	
	<i>Mugil liza</i> Valenciennes, 1836	x	x		x	x			DD	NT	
	<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007	x	x						LC	DD	
CICHLIFORMES											
	Cichlidae										
	<i>Cichla ocellaris</i> Bloch & Schneider, 1801								NE	LC	
	<i>Oreochromis niloticus</i> (Linnaeus, 1758)								LC	NE	
BLENNIIFORMES											
	Dactyloscopidae										
	<i>Dactyloscopus crossotus</i> Starks, 1913								LC	LC	
	Blenniidae										
	<i>Scartella cristata</i> (Linnaeus, 1758)								LC	LC	
	Labrisomidae										
	<i>Labrisomus nuchipinnis</i> (Quoy & Gaimard, 1824)								LC	LC	
ATHERINIFORMES											
	Atherinidae										
	<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)	x	x			x		x	x	LC	LC
	<i>Atherinella cf. blackburni</i> (Schultz, 1949)									LC	LC
	<i>Odontesthes bonariensis</i> (Valenciennes, 1835)	x								NE	DD
BELONIFORMES											
	Exocoetidae										
	<i>Cheilopogon melanurus</i> (Valenciennes, 1847)									LC	LC
	<i>Hirundichthys affinis</i> (Gunther, 1866)	x								LC	LC
	Hemiramphidae										
	<i>Hemiramphus balao</i> Lesueur, 1821	x								LC	DD
	<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)		x							LC	LC
	<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)	x								LC	LC
	<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)	x				x		x		LC	NT

Belonidae											
	<i>Ablennes hians</i> (Valenciennes, 1846)								LC	LC	
	<i>Strongylura marina</i> (Walbaum, 1792)	x		x			x		LC	LC	
	<i>Strongylura timucu</i> (Walbaum, 1792)	x					x		LC	LC	
	<i>Tylosurus acus acus</i> (Lacepède, 1803)	x		x					LC	LC	
	<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)								LC	LC	
CYPRINODONTIFORMES											
Poeciliidae											
	<i>Poecilia vivipara</i> Bloch & Schneider, 1801	x						x	x	NE	LC
CARANGIFORMES											
Rachycentridae											
	<i>Rachycentron canadum</i> (Linnaeus, 1766)									LC	LC
Echeneidae											
	<i>Echeneis naucrates</i> Linnaeus, 1758	x								LC	LC
	<i>Phtheichthys lineatus</i> (Menzies, 1791)	x								LC	LC
	<i>Remora remora</i> (Linnaeus, 1758)									LC	LC
Carangidae											
	<i>Carangoides bartholomaei</i> (Cuvier, 1833)	x		x			x			LC	LC
	<i>Caranx crysos</i> (Mitchill, 1815)	x	x	x		x	x			LC	LC
	<i>Caranx hippos</i> (Linnaeus, 1766)	x		x		x	x		x	LC	LC
	<i>Caranx latus</i> Agassiz, 1831	x	x	x	x	x	x		x	LC	LC
	<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	x					x		x	LC	LC
	<i>Hemicaranx amblyrhynchus</i> (Cuvier, 1833)									LC	LC
	<i>Oligoplites palometa</i> (Cuvier, 1832)	x		x		x	x			LC	LC
	<i>Oligoplites saliens</i> (Bloch, 1793)	x			x	x	x			LC	LC
	<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	x	x	x			x		x	LC	LC
	<i>Selene brownii</i> (Curvier, 1816)	x								LC	LC
	<i>Selene setapinnis</i> (Mitchill, 1815)		x				x			LC	LC

	<i>Selene vomer</i> (Linnaeus, 1758)	x	x	x		x	x	x	LC	LC	
	<i>Trachinotus carolinus</i> (Linnaeus, 1766)	x						x	LC	LC	
	<i>Trachinotus cayennensis</i> Cuvier, 1832								LC	DD	
	<i>Trachinotus falcatus</i> (Linnaeus, 1758)	x		x		x			LC	LC	
	<i>Trachinotus goodei</i> Jordan & Evermann, 1896			x					LC	LC	
ISTIOPHORIFORMES											
Sphyaenidae											
	<i>Sphyaena barracuda</i> (Edwards, 1771)	x	x			x		x	x	LC	LC
	<i>Sphyaena guachancho</i> Cuvier, 1829	x				x			x	LC	LC
PLEURONECTIFORMES											
Paralichthyidae											
	<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900	x		x		x				LC	LC
	<i>Citharichthys cornutus</i> (Günther, 1880)									LC	LC
	<i>Citharichthys macrops</i> Dresel, 1885							x	x	LC	LC
	<i>Citharichthys spilopterus</i> Günther, 1862	x		x		x	x	x	x	LC	LC
	<i>Etropus crossotus</i> Jordan & Gilbert, 1882	x				x			x	LC	LC
	<i>Etropus longimanus</i> Norman, 1933									NE	LC
	<i>Paralichthys brasiliensis</i> (Ranzani, 1842)	x				x	x	x	x	NE	LC
	<i>Paralichthys orbignyanus</i> (Valenciennes, 1839)							x		DD	DD
	<i>Paralichthys tropicus</i> Ginsburg, 1933		x			x				DD	NE
	<i>Syacium micrurum</i> Ranzani, 1842					x	x			LC	LC
	<i>Syacium papillosum</i> (Linnaeus, 1758)							x		LC	LC
Bothidae											
	<i>Bothus lunatus</i> (Linnaeus, 1758)									LC	LC
	<i>Bothus ocellatus</i> (Agassiz, 1831)	x								LC	LC
	<i>Bothus robinsi</i> Topp & Hoff, 1972									LC	LC
Achiridae											
	<i>Achirus achirus</i> (Linnaeus, 1758)	x							x	LC	LC

	<i>Achirus declivis</i> Chabanaud, 1940	x		x		x				LC	LC
	<i>Achirus lineatus</i> (Linnaeus, 1758)	x		x		x		x		LC	LC
	<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009									NE	VU
	<i>Trinectes microphthalmus</i> (Chabanaud, 1928)					x				LC	LC
	<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)					x		x		LC	LC
	Cynoglossidae					x					
	<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)	x				x			x	LC	LC
	<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)	x		x					x	LC	LC
	<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)									LC	LC
SYNGNATHIFORMES											
	Syngnathidae										
	<i>Bryx dunckeri</i> (Metzelaar, 1919)									LC	LC
	<i>Cosmocampus elucens</i> (Poey, 1868)									LC	LC
	<i>Hippocampus erectus</i> Perry, 1810	x								VU	VU
	<i>Hippocampus reidi</i> Ginsburg, 1933	x				x		x		NT	VU
	<i>Microphis brachyurus lineatus</i> (Kaup, 1856)									NE	NE
	<i>Microphis brachyurus brachyurus</i> (Bleeker, 1854)									LC	LC
	Fistulariidae										
	<i>Fistularia petimba</i> Lacepède, 1803	x								LC	LC
	<i>Fistularia tabacaria</i> Linnaeus, 1758					x		x	x	LC	LC
	<i>Syngnathus pelagicus</i> Linnaeus, 1758							x		LC	LC
	Dactylopteridae										
	<i>Dactylopterus volitans</i> (Linnaeus, 1758)	x				x		x		LC	LC
SCOMBRIFORMES											
	Trichiuridae										
	<i>Trichiurus lepturus</i> Linnaeus, 1758	x				x				LC	LC
	Scombridae										
	<i>Euthynnus alletteratus</i> (Rafinesque, 1810)									LC	LC

<i>Scomberomorus brasiliensis</i> Collette, Russo & Zavala-Camin, 1978	x					x			x	LC	LC
<i>Scomberomorus cavalla</i> (Cuvier, 1829)	x		x							LC	LC
<i>Scomberomorus maculatus</i> (Mitchill, 1815)										LC	NE
<i>Scomberomorus regalis</i> (Bloch, 1793)	x									LC	LC
Stromateidae											
<i>Peprilus paru</i> (Linnaeus, 1758)										LC	LC
TRACHINIFORMES											
Uranoscopidae											
<i>Astroscopus y-graecum</i> Cuvier, 1829										LC	LC
LABRIFORMES											
Labridae											
<i>Halichoeres poeyi</i> (Steindachner, 1867)										LC	LC
Scaridae											
<i>Nicholsina usta</i> (Valenciennes, 1840)										LC	LC
<i>Sparisoma amplum</i> (Ranzani, 1842)	x									LC	NT
<i>Sparisoma axillare</i> (Steindachner, 1878)	x									DD	VU
<i>Sparisoma radians</i> (Valenciennes, 1840)	x									LC	LC
PERCIFORMES											
Centropomidae											
<i>Centropomus ensiferus</i> Poey, 1860			x	x						LC	LC
<i>Centropomus mexicanus</i> Bocourt, 1868	x									LC	LC
<i>Centropomus parallelus</i> Poey, 1860	x	x	x	x		x	x	x	x	LC	LC
<i>Centropomus pectinatus</i> Poey, 1860	x									LC	LC
<i>Centropomus undecimalis</i> (Bloch, 1792)	x	x	x	x	x	x		x		LC	LC
Gerreidae											
<i>Diapterus auratus</i> Ranzani, 1842	x	x	x			x	x		x	LC	LC
<i>Diapterus rhombeus</i> (Cuvier, 1829)	x	x	x	x	x	x	x	x	x	LC	LC

<i>Eucinostomus argenteus</i> Baird & Girard, 1855	x	x	x	x	x	x	x	x	LC	LC
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	x	x				x		x	LC	LC
<i>Eucinostomus havana</i> (Nichols, 1912)									LC	LC
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	x	x		x		x		x	LC	LC
<i>Eugerres brasiliensis</i> (Cuvier, 1830)	x				x	x			LC	LC
<i>Gerres cinereus</i> (Walbaum, 1792)	x					x			LC	LC
<i>Ulaema lefroyi</i> (Goode, 1874)	x								LC	LC
Mullidae										
<i>Pseudupeneus maculatus</i> (Bloch, 1793)	x					x		x	LC	LC
Serranidae										
<i>Alphestes afer</i> (Bloch, 1793)	x								LC	DD
<i>Diplectrum radiale</i> (Quoy & Gaimard, 1824)						x		x	LC	LC
<i>Epinephelus adscensionis</i> (Osbeck, 1765)	x								LC	DD
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	x							x	VU	CR
<i>Hyporthodus nigritus</i> (Holbrook, 1855)									NT	EN
<i>Mycteroperca bonaci</i> (Poey, 1860)	x							x	NT	VU
<i>Mycteroperca tigris</i> (Valenciennes, 1833)									DD	DD
<i>Mycteroperca venenosa</i> (Linnaeus, 1758)									NT	DD
<i>Rypticus randalli</i> Courtenay, 1967	x		x			x		x	LC	LC
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)	x							x	LC	LC
<i>Serranus flaviventris</i> (Cuvier, 1829)						x		x	LC	LC
<i>Serranus phoebe</i> Poey, 1851									LC	LC
Pomacanthidae										
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	x								LC	DD
<i>Pomacanthus paru</i> (Bloch, 1787)								x	LC	DD
Chaetodontidae										
<i>Chaetodon ocellatus</i> (Bloch, 1787)									LC	DD
<i>Chaetodon striatus</i> Linnaeus, 1758								x	LC	LC

Haemulidae										
	<i>Anisotremus surinamensis</i> (Bloch, 1791)								x	DD DD
	<i>Anisotremus virginicus</i> (Linnaeus, 1758)								x	LC LC
	<i>Conodon nobilis</i> (Linnaeus, 1758)		x							LC LC
	<i>Genyatremus luteus</i> (Bloch, 1790)	x							x	NE LC
	<i>Haemulon aurolineatum</i> Cuvier, 1830									LC LC
	<i>Haemulon parra</i> (Desmarest, 1823)	x							x	LC LC
	<i>Haemulon plumierii</i> (Lacepède, 1801)		x							LC DD
	<i>Haemulon squamipinna</i> Rocha & Rosa, 1999								x	NE LC
	<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)								x	LC LC
	<i>Orthopristis ruber</i> (Cuvier, 1830)									LC LC
	<i>Pomadasys corvinaeformis</i> (Steindachner, 1868)	x	x						x	LC LC
	<i>Pomadasys crocro</i> (Cuvier, 1830)	x							x	DD LC
	<i>Pomadasys ramosus</i> Poey, 1860									NE LC
Lutjanidae										
	<i>Lutjanus alexandrei</i> Moura & Linderman, 2007	x							x	NE LC
	<i>Lutjanus analis</i> (Cuvier, 1828)	x		x					x	NT NT
	<i>Lutjanus apodus</i> (Walbaum, 1792)		x							LC NE
	<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	x								VU VU
	<i>Lutjanus griseus</i> (Linnaeus, 1758)	x	x						x	LC NE
	<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	x	x		x	x			x	DD NT
	<i>Lutjanus synagris</i> (Linnaeus, 1758)	x							x	NT NT
	<i>Ocyurus chrysurus</i> (Bloch, 1791)	x							x	DD NT
Polynemidae										
	<i>Polydactylus oligodon</i> (Gunther, 1860)									LC LC
	<i>Polydactylus virginicus</i> (Linnaeus, 1758)	x		x		x	x	x	x	LC LC
SCORPAENIFORMES										
Scorpaenidae										

<i>Scorpaena isthmensis</i> Meek & Hilderbrand, 1928	x								LC	LC
<i>Scorpaena plumieri</i> Bloch, 1789	x				x		x	x	LC	LC
Triglidae										
<i>Prionotus punctatus</i> (Bloch, 1793)	x				x			x	LC	LC
MORONIFORMES										
Ephippidae										
<i>Chaetodipterus faber</i> (Broussonet, 1782)	x		x		x		x	x	LC	LC
ACANTHURIFORMES										
Sciaenidae										
<i>Bairdiella ronchus</i> (Cuvier, 1830)	x		x		x				LC	LC
<i>Cynoscion acoupa</i> (Lacepède, 1801)			x				x		LC	NT
<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1833)			x				x		LC	LC
<i>Cynoscion leiarchus</i> (Cuvier, 1830)	x						x	x	LC	LC
<i>Cynoscion microlepidotus</i> (Cuvier, 1830)			x				x	x	LC	LC
<i>Cynoscion steindachneri</i> (Jordan, 1889)									LC	LC
<i>Cynoscion virescens</i> (Cuvier, 1830)									LC	LC
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)	x						x		LC	LC
<i>Larimus breviceps</i> Cuvier, 1830							x		LC	LC
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)			x				x		LC	LC
<i>Menticirrhus americanus</i> (Linnaeus, 1758)	x						x		LC	DD
<i>Menticirrhus littoralis</i> (Holbrook, 1847)								x	LC	DD
<i>Micropogonias furnieri</i> (Desmarest, 1823)	x		x				x	x	LC	LC
<i>Nebris microps</i> Cuvier, 1830									LC	LC
<i>Odontoscion dentex</i> (Cuvier, 1830)								x	LC	LC
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925									LC	DD
<i>Paralanchurus brasiliensis</i> (Steindachner, 1875)									LC	LC
<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)									LC	DD
<i>Pogonias cromis</i> (Linnaeus, 1766)									LC	EN

	<i>Stellifer brasiliensis</i> (Schultz, 1945)	x					NE	LC
	<i>Stellifer microps</i> (Steindachner, 1864)						LC	LC
	<i>Stellifer naso</i> (Jordan, 1889)						LC	LC
	<i>Stellifer rastrifer</i> (Jordan, 1889)	x			x		LC	LC
	<i>Stellifer stellifer</i> (Bloch, 1790)				x	x	DD	LC
	<i>Umbrina coroides</i> Cuvier, 1830						LC	LC
	Acanthuridae							
	<i>Acanthurus bahianus</i> Castelnau, 1855				x		LC	LC
	<i>Acanthurus chirurgus</i> (Bloch, 1787)	x				x	LC	LC
	<i>Acanthurus coeruleus</i> Bloch & Schneider, 1801						LC	LC
SPARIFORMES								
	Lobotidae							
	<i>Lobotes surinamensis</i> (Bloch, 1790)	x			x		LC	LC
	Sparidae							
	<i>Archosargus probatocephalus</i> (Walbaum, 1792)	x					LC	DD
	<i>Archosargus rhomboidalis</i> (Linnaeus, 1758)	x	x		x		LC	LC
LOPHIIFORMES								
	Antennariidae							
	<i>Antennarius multiocellatus</i> (Valenciennes, 1837)						x	LC DD
	<i>Antennarius striatus</i> (Shaw, 1794)	x				x	x	LC DD
	Ogcocephalidae							
	<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	x			x	x	NE	LC
TETRAODONTIFORMES								
	Ostraciidae							
	<i>Acanthostracion quadricornis</i> (Linnaeus, 1758)				x	x	LC	LC
	<i>Lactophrys trigonus</i> (Linnaeus, 1758)	x				x	LC	LC
	<i>Lactophrys triqueter</i> (Linnaeus, 1758)	x					LC	NE
	Balistidae							

<i>Balistes vetula</i> Linnaeus, 1758								x	NT	NT		
<i>Melichthys niger</i> (Bloch, 1786)									LC	LC		
Monacanthidae												
<i>Aluterus heudelotii</i> Hollard, 1855								x	LC	LC		
<i>Aluterus monoceros</i> (Linnaeus, 1758)									LC	NT		
<i>Aluterus schoepfii</i> (Walbaum, 1792)									LC	LC		
<i>Cantherhines pullus</i> (Ranzani, 1842)									LC	LC		
<i>Monacanthus ciliatus</i> (Mitchill, 1818)									LC	LC		
<i>Stephanolepis hispidus</i> (Linnaeus, 1766)								x	LC	LC		
<i>Stephanolepis setifer</i> (Bennett, 1831)								x	LC	LC		
Tetraodontidae												
<i>Canthigaster figueiredoi</i> Moura & Castro, 2002									LC	LC		
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)								x	LC	LC		
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	x			x				x	x	LC	LC	
<i>Sphoeroides greeleyi</i> Gilbert, 1900	x					x		x	x	LC	LC	
<i>Sphoeroides nephelus</i> (Goode & Bean, 1882)	x									LC	NE	
<i>Sphoeroides spengleri</i> (Bloch, 1785)	x					x			x	LC	LC	
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	x	x		x		x		x	x	LC	DD	
<i>Sphoeroides tyleri</i> Shipp, 1972										LC	LC	
Diodontidae												
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	x					x		x		LC	LC	
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)									x	x	LC	LC
<i>Diodon holocanthus</i> Linnaeus, 1758											LC	LC
<i>Diodon hystrix</i> Linnaeus, 1758											LC	LC

References

- Affe HM de J, Conceição LP, Rocha DSB, et al (2021) Phytoplankton community in a tropical estuarine gradient after an exceptional harmful bloom of akashiwo sanguinea (Dinophyceae) in the todos os santos bay. *Ocean and Coastal Research* 69:. <https://doi.org/10.1590/2675-2824069.20-004hmdja>
- Barletta M, Barletta-Bergan A, Saint-Paul U, Hubold G (2003) Seasonal changes in density, biomass, and diversity of estuarine fishes in tidal mangrove creeks of the lower Caeté Estuary (northern Brazilian coast, east Amazon). *Marine Ecology Progress Series* 256:217–228. <https://doi.org/10.3354/meps256217>
- Barletta M, Blaber SJM (2007) Comparison of fish assemblages and guilds in tropical habitats of the Embley (Indo-West Pacific) and Caeté (Western Atlantic) Estuaries. *Bulletin of Marine Science* 80:647–680
- Barletta M, Jaureguizar AJ, Baigun C, et al (2010) Fish and aquatic habitat conservation in South America: A continental overview with emphasis on neotropical systems. *Journal of Fish Biology* 76:2118–2176. <https://doi.org/10.1111/j.1095-8649.2010.02684.x>
- Barletta M, Lima ARA (2019) Systematic Review of Fish Ecology and Anthropogenic Impacts in South American Estuaries: Setting Priorities for Ecosystem Conservation. *Frontiers in Marine Science* 6:. <https://doi.org/10.3389/fmars.2019.00237>
- Bernardes M, Knoppers B, Rezende C, et al (2012) Land-sea interface features of four estuaries on the South America Atlantic coast. *Brazilian Journal of Biology* 72:761–774. <https://doi.org/10.1590/s1519-69842012000400011>
- Blaber SJM (2000) Tropical estuarine fishes. ecology, exploitation and conservation. Blackwell Science 372. [https://doi.org/10.1643/0045-8511\(2001\)001\[0880:J2.0.CO:2](https://doi.org/10.1643/0045-8511(2001)001[0880:J2.0.CO:2)
- Branco JO, Freitas Júnior F, Fracasso HAA, Barbieri E (2011) Biodiversidade no estuário do Saco da Fazenda, Itajaí-SC. *O Mundo da Saúde*. 35(1): 12-22.
- Bruno DO, Barbini SA, Díaz de Astarloa JM, Martos P (2013) Fish abundance and distribution patterns related to environmental factors in a choked temperate coastal lagoon (Argentina). *Brazilian Journal of Oceanography* 61:43–53. <https://doi.org/10.1590/S1679-87592013000100005>
- Camara EM, Andrade-Tubino MF de, Franco TP, Araújo FG (2020) Multilevel decomposition of spatial and environmental effects on nearshore fish assemblages in tropical semi-enclosed ecosystems. *Estuarine, Coastal and Shelf Science* 237:. <https://doi.org/10.1016/j.ecss.2020.106691>
- Catelani PA, Petry, AC, Di Dario F, Santos VLM, Mincarone MM (2014) Fish composition (Teleostei) of the estuarine region of the Macaé River, southeastern Brazil. *Check List* 10(4). <https://10.15560/10.4.900>
- Chao NL, Frédou FL, Haimovici M, et al (2015) A popular and potentially sustainable fishery resource under pressure-extinction risk and conservation of Brazilian Sciaenidae (Teleostei: Perciformes). *Global Ecology and Conservation* 4:117–126. <https://doi.org/10.1016/j.gecco.2015.06.002>
- Clarke K, Warwick R (2001) Change in marine communities: an approach to statistical analysis and interpretation. 2nd edition. Primer-E, Plymouth
- Costanza R, de Groot R, Sutton P, et al (2014) Changes in the global value of ecosystem services. *Global Environmental Change*. <https://doi.org/10.1016/j.gloenvcha.2014.04.002>
- Elliott M, Whitfield AK, Potter IC, et al (2007) The guild approach to categorizing estuarine fish.pdf. *Fish and Fisheries* 8:241–268. <https://doi.org/10.1136/adc.2002.016303>
- Franco A, Elliott M, Franzoi P, Torricelli P (2008) Life strategies of fishes in European estuaries: The functional guild approach. *Marine Ecology Progress Series* 354:219–228. <https://doi.org/10.3354/meps07203>
- Ferreira B, Maida, M (2006) Monitoramento dos recifes de coral do Brasil. *Série Biodiversidade*, 18, MMA, Brasília
- Fricke, R., Eschmeyer, W. N. & Van der Laan R (2020) Eschmeyer's Catalog of Fishes: Genera, Species, References. <http://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp>. Accessed 21 Oct 2020
- Garnier S, Ross N, Rudis R, et al (2021) Rvision - Colorblind- Friendly Color Maps for R

- Harrison TD, Whitfield AK (2021) A global assessment of fish estuary associations: A numerical approach to assessing estuary-associated fish functional guilds. *Fish and Fisheries* 1–25. <https://doi.org/10.1111/faf.12558>
- ICMbio (2018) Livro Vermelho da Fauna Brasileira Ameaçada de Extinção: Volume I. ICMBIO/MMA, Brasília
- König C, Weigelt P, Schrader J, et al (2019) Biodiversity data integration—the significance of data resolution and domain. *PLoS Biology* 17:1–16. <https://doi.org/10.1371/journal.pbio.3000183>
- Lessa GC (2020) Aspectos Básicos da Circulação Estuarina e sua relação com o Ambiente Costeiro. In: Muehe D, Lins-de-Barros FM, Pinheiro LS (eds) *Geografia Marinha: Oceanos e Costas na Perspectiva de Geógrafos*. PGGM, Rio de Janeiro, pp 167–197
- Lins Oliveira JE, Leite TS, Castro PT, Araripe HGA (1999) Caracterização dos Ecossistemas Costeiros dos Estados do Rio Grande do Norte, Ceará e Piauí. Natal
- Loureiro SN, Reis-Filho JA, Giarrizzo T (2016) Evidence for habitat-driven segregation of an estuarine fish assemblage. *Journal of fish biology* 89:804–820. <https://doi.org/10.1111/jfb.13017>
- Mateus M, Mateus S, Barrett (2008) Basic concepts of estuarine ecology. In: *Perspectives on Integrated Coastal Zone Management in South America*. IST Press, Lisboa, pp 3–14
- Mathieson S, Cattrijsse A, Costa MJ, et al (2000) Fish assemblages of European tidal marshes: A comparison based on species, families and functional guilds. *Marine Ecology Progress Series* 204:225–242. <https://doi.org/10.3354/meps204225>
- Medeiros APM de (2013) Distribuição e dieta da ictiofauna em área estuarino-recifal da APA Barra do rio Mamanguape, Paraíba, Brasil. 87
- Melo FAG, Viana JQ, Araújo TM, et al (2021) Ichthyofauna of shallow zones of the estuary of Timonha and Ubatuba Rivers, Northeastern Brazil. *Biota Amazônia* 33–40. <https://doi.org/http://dx.doi.org/10.18561/2179-5746/biotaamazonia.v11n1p33-40>
- Mourão KRM, Ferreira V, Lucena-Frédou F (2014) Composition of functional ecological guilds of the fish fauna of the internal sector of the amazon estuary, Pará, Brazil. *Anais da Academia Brasileira de Ciências* 86:1783–1800. <https://doi.org/10.1590/0001-3765201420130503>
- Murase A, Angulo A, Miyazaki Y, et al (2014) Marine and estuarine fish diversity in the inner Gulf of Nicoya, Pacific coast of Costa Rica, Central America. *Check List* 10:1401–1413. <https://doi.org/10.15560/10.6.1401>
- Nelson JS, Grande TC, Wilson MVH (2016) *Fishes of the world*. John Wiley & Sons, New Jersey
- NOAA (2019) Life in an estuary. <https://www.noaa.gov/education/resource-collections/marine-life/life-in-estuary>. Accessed 10 Nov 2020
- Oksanen JF, Blanchet G, Friendly M, et al (2019) *Vegan: Community Ecology Package*. R package version 2.5-6.
- Paiva ACG de, Chaves P de T, Araújo ME de (2013) Distribution of estuarine fish fauna along coast of Brazil. *Tropical Oceanography Online* 41:1–110. <https://doi.org/10.59/14/♦to.2013.0077>
- Passarone R, Aparecido KC, Eduardo LN, et al (2019) Ecological and conservation aspects of bycatch fishes: An evaluation of shrimp fisheries impacts in northeastern Brazil. *Brazilian Journal of Oceanography* 67:1–10. <https://doi.org/10.1590/s1679-87592019029106713>
- Passos AC dos, Contente RF, Abatepaulo FV, et al (2013) Analysis of fish assemblages in sectors along a salinity gradient based on species, families and functional groups. *Brazilian Journal of Oceanography* 61:251–264. <https://doi.org/10.1590/S1679-87592013000400006>
- Pereira L de JG (2018) Análise Multidisciplinar de uma Pescaria Proibida: Estudo de Caso da Pesca do Mero *Epinephelus itajara* (Lichtenstein, 1822) na costa Amazônica Brasileira
- Pinheiro L de S, Coriolano LN, Costa MF da, Dias JA (2008) O Nordeste brasileiro e a gestão costeira. *Revista de Gestão Costeira Integrada* 8:5–10. <https://doi.org/10.5894/rgci58>

- Potter IC, Chuwen BM, Hesp SA, et al (2011) Implications of the divergent use of a suite of estuaries by two exploited marine fish species. *Journal of Fish Biology* 79:662–691. <https://doi.org/10.1111/j.1095-8649.2011.03051.x>
- Potter IC, Tweedley JR, Elliott M, Whitfield AK (2015) The ways in which fish use estuaries: a refinement and expansion of the guild approach. *Fish and Fisheries* 16:230–239. <https://doi.org/10.1111/faf.12050>
- R Core T (2019) R: A language and environment for statistical computing.
- Reis RE, Albert JS, di Dario F, et al (2016) Fish biodiversity and conservation in South America. *Journal of fish biology* 89:12–47. <https://doi.org/10.1111/jfb.13016>
- Roshni K, Renjithkumar CR, Raghavan R, Ranjeet K (2021a) Fish distribution and assemblage structure in a hydrologically fragmented tropical estuary on the south-west coast of India. *Regional Studies in Marine Science* 43:101693. <https://doi.org/10.1016/j.rsma.2021.101693>
- Roshni K, Renjithkumar CR, Raghavan R, Ranjeet K (2021b) Fish distribution and assemblage structure in a hydrologically fragmented tropical estuary on the south-west coast of India. *Regional Studies in Marine Science* 43:101693. <https://doi.org/10.1016/j.rsma.2021.101693>
- Rotundo MM, Gaulia LA, Cardoso GS, et al (2020) Ichthyofauna from Santos-São Vicente upper estuary: a study before and during fire at Santos port terminal. *Research, Society and Development* 9:e6269108901. <https://doi.org/10.33448/rsd-v9i10.8901>
- Sampaio De Souza C, Júnior PM Distribution and Abundance of Carangidae (Teleostei, Perciformes) Associated with Oceanographic Factors along the Northeast Brazilian Exclusive Economic Zone. *Braz arch biol technol* v 51:1267–1278
- Santos JAP, Schmiegelow JMM, Rotundo MM, Barrella W (2015) Composição e variação temporal da assembleia de peixes do alto sistema estuarino de Santos, São Paulo, Brasil. *Boletim do Instituto de Pesca*, 41(4): 945-959.
- Spier D, Gerum HLN, Bornatowski H, et al (2018) Ictiofauna da plataforma interna do Paraná, Brasil: Checklist, distribuição geográfica, importância econômica e estado de conservação. *Biota Neotropica* 18:. <https://doi.org/10.1590/1676-0611-bn-2017-0385>
- Suguio K, Bezerra FHR, Barreto AMF (2011) Luminescence dated Late Pleistocene wave-built terraces in northeastern Brazil. *Anais da Academia Brasileira de Ciências* 83:907–920
- Team RC (2020) R: A language and environment for statistical computing.
- Vasconcelos RP, Henriques S, França S, et al (2015) Global patterns and predictors of fish species richness in estuaries. *Journal of Animal Ecology* 84:1331–1341. <https://doi.org/10.1111/1365-2656.12372>
- Vendel AL, Chaves PDT (2006) Use of an estuarine environment (Barra do Saí lagoon, Brazil) as nursery by fish. *Revista Brasileira de Zoologia* 23:1117–1122. <https://doi.org/10.1590/S0101-81752006000400020>
- Vicente F, Loeb M v., de Paiva ACG, et al (2020) Integrative systematics unveils the controversial identity of Engraulidae fishing stocks in a Neotropical estuary, northeast Brazil. *Neotropical Ichthyology* 18:1–17. <https://doi.org/10.1590/1982-0224-2020-0037>
- Vilar CC, Joyeux J-C (2018a) Synopsis of the knowledge on the Brazilian estuarine fishes. *PeerJPreprints* 37. <https://doi.org/https://doi.org/10.7287/peerj.preprints.27085v1> |
- Vilar CC, Joyeux J-C (2018b) Synopsis of the knowledge on the Brazilian estuarine fishes. *PeerJ Preprints* 37. <https://doi.org/10.7287>
- Vilar CC, Joyeux JC, Giarrizzo T, et al (2013) Local and regional ecological drivers of fish assemblages in Brazilian estuaries. *Marine Ecology Progress Series* 485:181–197. <https://doi.org/10.3354/meps10343>
- Wickham H (2016) *Ggplot2: Elegant Graphics for Data Analysis*
- WoRMS EB (2020) World Register of Marine Species. Accessed 21 Oct 2020

- Yee DA, Yee SH, Kneitel JM, Juliano SA (2007) Richness-productivity relationships between trophic levels in a detritus-based system: significance of abundance. *Oecologia* 154:377–385. <https://doi.org/10.1007/s00442-007-0837-5>
- Zapata C, Puente A, Garca A, et al (2018a) Assessment of ecosystem services of an urbanized tropical estuary with a focus on habitats and scenarios. *PLoS ONE* 13:1–19. <https://doi.org/10.1371/journal.pone.0203927>
- Zapata C, Puente A, Garca A, et al (2018b) Assessment of ecosystem services of an urbanized tropical estuary with a focus on habitats and scenarios. *PLoS ONE* 13:1–19. <https://doi.org/10.1371/journal.pone.0203927>

Supplementary Material

Table I: Classification of feeding mode functional group (FMFG): zoobenthivore (ZB), piscivore (PV), zooplanktivore (ZP), herbivore (HV), omnivore (OV), detritivore (DV) and opportunistic (OP); estuarine use functional group (EUFG): marine migrant (MM), marine stragglers (MS), estuarine fishes (ES), anadromous species (AN), amphidromous (AM), catadromous (CA), freshwater migrants (FM) and freshwater species (FS), for species observed during the study and references used for each category.

Species	FMFG	REF	EUFG	REF
<i>Ginglymostoma cirratum</i> (Bonnaterre, 1788)	PV	Compagno, 2001	MM	Compagno, 2001
<i>Carcharhinus leucas</i> (Muller & Henle, 1839)	PV	WoRMS 2020	MM	WoRMS 2020
<i>Carcharhinus limbatus</i> (Muller & Henle, 1839)	PV	Castro 1996	MM	Castro, 1996
<i>Carcharhinus porosus</i> (Ranzani, 1839)	PV	Ebert et al. 2013	MM	Ebert et al., 2013
<i>Isogomphodon oxyrinchus</i> (Muller & Henle, 1839)	PV	Compagno 1984	MM	Ebert et al., 2013
<i>Rhizoprionodon lalandii</i> (Muller & Henle, 1839)	PV	Ferreira et al. 2019	MS	Ferreira et al., 2019
<i>Rhizoprionodon porosus</i> (Poey, 1861)	PV	Ferreira et al., 2019	MS	Ferreira et al., 2019
<i>Sphyrna lewini</i> (Griffith & Smith, 1834)	PV	Sommer et al. 1996	MM	Compagno, 1994
<i>Sphyrna tiburo</i> (Linnaeus, 1758)	ZB	Alves and Filho 1996	MM	Ebert et al., 2013
<i>Sphyrna tudes</i> (Valenciennes, 1822)	PV	Compagno, 1984	MM	Ebert et al., 2013
<i>Narcine brasiliensis</i> (Olfers, 1831)	ZB	Michael 1993	MS	Reis-Filho et al. 2010
<i>Pristis pristis</i> Muller & Henle, 1841	PV	Miller 2005	MM	Thorburn et al. 2007
<i>Pseudobatos percellens</i> (Walbaum, 1792)	ZB	Passos et al. 2013	MS	Reis-Filho et al., 2010
<i>Fontitrygon geijskesi</i> (Boeseman, 1948)	-	-	MM	Last et al. 2016
<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)	ZB	Lieske and Myers 1994	MS	Reis-Filho et al., 2010
<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	ZB	Freitas et al. 2019	MS	Uyeno et al. 1983
<i>Hypanus marianae</i> (Gomes, Rosa & Gadig, 2000)	ZB	Ferreira et al., 2019	MS	Ferreira et al., 2019
<i>Hypanus say</i> (Lesueur, 1817)	ZB	Smith 1997	MS	Last et al., 2016
<i>Gymnura micrura</i> (Bloch & Scheneider, 1801)	ZB	Lima et al. 2021	MS	Reis-Filho et al., 2010
<i>Aetobatus narinari</i> (Euphrasen, 1790)	ZB	Last et al., 2016	MS	Ajemian et al. 2012
<i>Mobula hypostoma</i> (Bancroft, 1831)	ZB	Michael, 1993	MS	Michael, 1993
<i>Rhinoptera bonasus</i> (Mitchill, 1815)	ZB	Last et al., 2016	MS	Reis-Filho et al., 2010

<i>Elops saurus</i> Linnaeus, 1766	PV	da Silva and Fabré 2019	MS	Vasconcelos Vasconcelos Filho and Oliveira 1999
<i>Megalops atlanticus</i> Valenciennes, 1847	PV	Whitehead and Vergara 1978	MM	Whitehead and Vergara, 1978
<i>Albula vulpes</i> (Linnaeus, 1758)	ZB	Passos et al., 2013	MM	Paiva et al. 2009
<i>Gymnothorax funebris</i> Ranzani, 1839	ZB	Carvalho-Filho 1999	MS	Vasconcelos Filho & Oliveira, 1999
<i>Gymnothorax moringa</i> (Cuvier, 1829)	ZB	Carvalho-Filho, 1999	MM	Reis-Filho et al., 2010
<i>Gymnothorax ocellatus</i> Agassiz, 1831	ZB	Santos and Castro 2003	MS	Vasconcelos Filho and Oliveira 1999
<i>Gymnothorax vicinus</i> (Castelnau, 1855)	OP	Miller 1987	MS	Claro 1994
<i>Ahlia egmontis</i> (Jordan, 1884)	ZB	Pimentel, 2018	MS	Ross and Rohde 2003
<i>Myrichthys breviceps</i> (Richardson, 1848)	ZB	Ross et al. 2007	MS	Robins and Ray 1986
<i>Myrichthys ocellatus</i> (Lesueur, 1825)	ZB	Luiz et al. 2008	MS	Monteiro-Neto et al. 2013
<i>Myrophis punctatus</i> Lütken, 1852	PV	Alves and Filho 1996	MM	Favero et al. 2019
<i>Ophichthus cylindroideus</i> (Ranzani, 1839)	PV	Favero et al. 2019	MM	Favero et al. 2019
<i>Ophichthus ophis</i> (Linnaeus, 1758)	PV	Randall 1967	MS	Schneider 1990
<i>Cynoponticus savanna</i> (Bancroft, 1831)	ZB	Carpenter 2003	ES	Carpenter 2003
<i>Ariosoma balearicum</i> (Delaroche, 1809)	-	-	MS	Miller 2002; Ross et al. 2007
<i>Chirocentron bleekermanus</i> (Poey, 1867)	ZP	Passos et al. 2013	MS	Passos et al. 2013
<i>Odontognathus mucronatus</i> Lacepède, 1800	HV	Passarone et al. 2019	MS	Passarone et al. 2019
<i>Pellona flavipinnis</i> (Valenciennes, 1837)	OV	di Dario 2013	FM	di Dario 2013
<i>Pellona harroweri</i> (Fowler, 1917)	ZB	Passarone et al. 2019	MS	Passarone et al. 2019
<i>Anchoa filifera</i> (Fowler, 1915)	ZB	Passarone et al. 2019	ES	Passarone et al. 2019
<i>Anchoa hepsetus</i> (Linnaeus, 1758)	ZP	Nizinski and Munroe 2002	MM	Nizinski and Munroe 2002
<i>Anchoa januaria</i> (Steindachner, 1879)	ZP	Lima et al. 2021	ES	Reis-Filho et al. 2010
<i>Anchoa lyolepis</i> (Evermann & Marsh, 1900)	ZP	Passos et al. 2013	ES	Reis-Filho et al. 2010
<i>Anchoa parva</i> (Meek & Hildebrand, 1923)	ZP	Cervigón 1993	MM	Cervigón 1993
<i>Anchoa spinifer</i> (Valenciennes, 1848)	PV	Passarone et al. 2019	MM	Passarone et al. 2019

<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)	ZB	Ferreira et al. 2019	MM	Ferreira et al., 2019
<i>Anchovia clupeioides</i> (Swainson, 1839)	ZP	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Anchoviella lepidentostole</i> (Fowler, 1911)	PV	Favero et al. 2019	MM	Favero et al. 2019
<i>Cetengraulis edentulus</i> (Cuvier, 1829)	ZP	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Engraulis anchoita</i> Hubbs & Marini 1935	ZP	Vasconcellos et al. 1998	MS	Passos et al. 2013
<i>Lycengraulis batesii</i> (Gunther, 1868)	PV	Tejerina-Garro and de Merona 2010	FM	Bornbusch 1988
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	PV	Passarone et al. 2019	ES	Passarone et al. 2019
<i>Pterengraulis atherinoides</i> (Linnaeus, 1766)	PV	Krumme et al. 2005	MS	Krumme et al. 2004
<i>Harengula clupeiola</i> (Cuvier, 1829)	ZP	Paiva et al. 2008	MS	Vasconcelos Filho and Oliveira 1999
<i>Harengula jaguana</i> Poey, 1865	ZP	Passos et al. 2013	MS	Peralta-Meixueiro and Vega-Cendejas 2011
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)	ZP	Favero et al. 2019	ES	Reis-Filho et al. 2010
<i>Opisthonema oglinum</i> (Lesueur, 1818)	ZP	Vasconcelos-Filho 1979	MS	Vasconcelos Filho and Oliveira 1999
<i>Platanichthys platana</i> (Regan, 1917)	ZP	Passos et al. 2013	ES	Passos et al. 2013
<i>Rhinosardinia amazonica</i> (Steindachner, 1879)	OV	Alves and Filho 1996	FM	Whitehead 1985
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)	ZP	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Hoplias malabaricus</i> (Bloch, 1794)	PV	Alves and Filho 1996	ES	Costa Novaes and Carvalho 2011
<i>Prochilodus argenteus</i> Spix & Agassiz, 1829	HV	Alves and Filho 1996	FM	Melo 2011
<i>Pygocentrus nattereri</i> Kner, 1858	ZB	Mills and Vevers 1989	FS	Ferreira et al. 2014
<i>Serrasalmus rhombeus</i> (Linnaeus, 1766)	PV	Planquette et al. 1996	FS	Planquette et al. 1996
<i>Serrasalmus spilopleura</i> Kner, 1858	PV	Alves and Filho 1996	FS	Agostinho and Júlio 200)
<i>Schizodon fasciatus</i> Spix & Agassiz, 1829	HV	Planquette et al. 1996	FS	Planquette et al. 1996
<i>Leporinus friderici</i> (Bloch, 1794)	ZP	Boujard et al. 1997	FS	Cordiviola de Yuan and Pignalberi de Hassan 1985 Cordiviola de Yuan and

				Pignalberi de Hassan 1985
<i>Steindachnerina elegans</i> (Steindachner, 1875)	DV	Ribeiro et al. 2007	FS	Ribeiro et al. 2007
<i>Astyanax bimaculatus</i> (Linnaeus, 1758)	ZB	Alves and Filho 1996	FS	Kenny 1995
<i>Hemigrammus marginatus</i> Ellis, 1911	OV	Barreto et al. 2018	FS	Barreto et al. 2018
<i>Hypostomus plecostomus</i> (Linnaeus, 1758)	HV	Alves and Filho 1996	FS	Oliveira and Isaac 2013
<i>Loricaria parnabybae</i> Steindachner, 1907	ZB	Alves and Filho 1996	FS	Londoño-Burbano et al. 2020
<i>Aspistor luniscutis</i> (Valenciennes, 1840)	OV	Denadai et al. 2012	MS	Denadai et al. 2012
<i>Aspistor quadriscutis</i> (Valenciennes, 1840)	ZB	Denadai et al. 2012	MS	Denadai et al. 2012
<i>Bagre bagre</i> (Linnaeus, 1766)	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Bagre marinus</i> (Mitchill, 1815)	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)	ZB	Passarone et al. 2019	ES	Passarone et al. 2019
<i>Cathorops spixii</i> (Agassiz, 1829)	ZB	Passarone et al. 2019	ES	Passarone et al. 2019
<i>Genidens genidens</i> (Cuvier, 1829)	PV	Passos et al. 2013	MM	Passos et al. 2013
<i>Notarius grandicassis</i> (Valenciennes, 1840)	ZP	Mendes 1999	ES	Cervigón, 1992
<i>Sciades couma</i> (Valenciennes, 1840)	ZP	le Bail et al. 2000	ES	Reis-Filho et al. 2010
<i>Sciades herzbergii</i> (Bloch, 1794)	ZB	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Sciades parkeri</i> (Traill, 1832)	PV	Mendes 1999	ES	Leopold 2004
<i>Sciades proops</i> (Valenciennes, 1840)	ZB	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Aspredo aspredo</i> (Linnaeus, 1758)	OV	Herbert 2015	FS	Froese and Pauly 2018
<i>Aspredinichthys tibicen</i> (Valenciennes, 1840)	-	-	FS	le Bail et al. 2000
<i>Ageneiosus inermis</i> (Linnaeus, 1766)	PV	Boujard et al. 1997	FM	Boujard et al. 1997
<i>Pseudauchenipterus nodosus</i> (Bloch, 1794)	OV	le Bail et al. 2000	FM	Kenny 1995
<i>Trachelyopterus galeatus</i> (Linnaeus, 1766)	ZB	Alves and Filho 1996	-	-
<i>Pimelodella cristata</i> (Muller & Troschel, 1849)	-	-	FS	Mol 1958
<i>Pimelodella lateristriga</i> (Lichtenstein, 1823)	PV	Alves and Filho 1996	-	-

<i>Brachyplatystoma vaillantii</i> (Valenciennes, 1840)	PV	Cuvier and Valenciennes 1840	FS	le Bail et al. 2000
<i>Pseudoplatystoma fasciatum</i> (Linnaeus, 1766)	OV	Burgess 1989	FS	Burgess 1989
<i>Sorubim lima</i> (Bloch & Schneider, 1801)	PV	Goulding 1981	-	-
<i>Synodus foetens</i> (Linnaeus, 1766)	PV	Cruz-Escalona et al. 2005	MS	Vasconcelos Filho and Oliveira 1999
<i>Synodus poeyi</i> Jordan, 1887	-	-	-	-
<i>Trachinocephalus myops</i> (Forster, 1801)	PV	Fischer et al. 1990	MS	Heemstra 1995
<i>Holocentrus adscensionis</i> (Osbeck, 1765)	ZB	Greenfield and Thomerson 1997	MS	Silva Júnior et al. 2015
<i>Amphichthys cryptocentrus</i> (Valenciennes, 1837)	ZB	Collette 2003	ES	Reis-Filho et al. 2010
<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)	ZB	Collette 2003	MS	Uyeno et al. 1983
<i>Thalassophryne nattereri</i> Steindachner, 1876	ZB	Collette 2003b	MS	Vasconcelos Filho and Oliveira 1999
<i>Thalassophryne punctata</i> Steindachner, 1876	-	-	-	-
<i>Phaeoptyx pigmentaria</i> (Poey, 1860)	ZP	Greenfield and Johnson 1990	MS	Greenfield & Johnson, 1990
<i>Dormitator maculatus</i> (Bloch, 1792)	PV	Alves and Filho 1996	AM	Elliott et al. 2007
<i>Eleotris pisonis</i> (Gmelin, 1789)	ZB	Alves and Filho 1996	FM	Nordlie 1981
<i>Erotelis smaragdus</i> (Valenciennes, 1837)	-	-	MM	Dawson 1971
<i>Guavina guavina</i> (Valenciennes, 1837)	ZB	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Awaous tajasica</i> (Lichtenstein, 1822)	ZB	Miller 2005	AM	Miller 2005
<i>Bathygobius mystacium</i> Ginsburg, 1947	PV	Silva 2018	MS	Tornabene et al. 2010
<i>Bathygobius soporator</i> (Valenciennes, 1837)	ZB	Passos et al. 2013	MM	Passos et al. 2013
<i>Coryphopterus glaucofraenum</i> Gill, 1863	ZB	Smith 1997	MS	Finley and Forrester 2003
<i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882)	DV	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Ctenogobius saepepallens</i> (Gilbert & Randall, 1968)	ZB	Randall 2004	MS	Baldwin and Smith 2003; Ross and Rhode 2004
<i>Ctenogobius shufeldti</i> (Jordan & Eigenmann, 1887)	OV	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Ctenogobius smaragdus</i> (Valenciennes, 1837)	DV	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Ctenogobius stigmaticus</i> (Poey, 1860)	DV	Ferreira et al. 2019	ES	Ferreira et al. 2019

<i>Evorthodus lyricus</i> (Girard, 1858)	DV	Ferreira et al. 2019	MS	Vasconcelos Filho and Oliveira 1999
<i>Gobioides broussonnetii</i> Lacepède, 1800	DV	da Silva and Fabré et al. 2019	ES	Passos et al. 2013
<i>Gobionellus oceanicus</i> (Pallas, 1770)	DV	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Gobionellus stomatus</i> Starks, 1913	DV	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Microdesmus bahianus</i> Dawson, 1973	-	-	MS	Castro 2005
<i>Microdesmus longipinnis</i> (Weymouth, 1910)	-	-	ES	Harrison et al. 2003
<i>Microgobius meeki</i> Evermann & Marsh, 1899	ZB	Ferreira et al. 2019	MS	Ferreira et al. 2019
<i>Abudefduf saxatilis</i> (Linnaeus, 1758)	OP	Emery 1978	MS	Emery 1978
<i>Stegastes fuscus</i> (Cuvier, 1830)	OV	Paiva et al. 2009	MS	Cervigón 1993
<i>Stegastes variabilis</i> (Castelnau, 1855)	ZB	Cervigón 1993	MS	Cervigón 1993
<i>Opistognathus cuvierii</i> Valenciennes, 1836	ZP	Reis-Filho 2011	MS	Haimovici and Klippel 1999
<i>Mugil brevisrostris</i> (Ribeiro, 1915)	-	-	MM	Barletta and Dantas 2016
<i>Mugil curema</i> Valenciennes, 1836	DV	Elliott et al. 2007	MM	Ferreira et al. 2019
<i>Mugil curvidens</i> Valenciennes, 1836	DV	da Silva and Fabré et al. 2019	ES	Passos et al. 2013
<i>Mugil incilis</i> Hancock, 1830	DV	Favero et al. 2019	ES	Passos et al. 2013
<i>Mugil liza</i> Valenciennes, 1836	DV	da Silva and Fabré et al. 2019	CA	Passos et al. 2013
<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007	DV	da Silva and Fabré et al. 2019	MM	da Silva and Fabré et al. 2019
<i>Cichla ocellaris</i> Bloch & Schneider, 1801	PV	Alves and Filho 1996	FS	Keith et al. 2000
<i>Oreochromis niloticus</i> (Linnaeus, 1758)	ZB	Alves and Filho 1996	FS	de Moor and Bruton 1988
<i>Dactyloscopus crossotus</i> Starks, 1913	ZB	Guedes et al. 2015	MS	Paiva et al. 2009
<i>Scartella cristata</i> (Linnaeus, 1758)	HV	Randall 1996	MM	Cervigón 1994
<i>Labrisomus nuchipinnis</i> (Quoy & Gaimard, 1824)	ZB	Randall 1996	MS	Cervigón 1994
<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)	OV	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Atherinella cf. blackburni</i> (Schultz, 1949)	PV	Favero et al. 2019	MM	Favero et al. 2019
<i>Odontesthes bonariensis</i> (Valenciennes, 1835)	ZP	Piedras and Pouey 2005	FM	Nakamura et al. 1986
<i>Cheilopogon melanurus</i> (Valenciennes, 1847)	PV/ZP	Rahaman 2015	MS	Rahaman 2015
<i>Hirundichthys affinis</i> (Gunther, 1866)	PV	Oxenford 2007	MS	Cervigón et al. 1992
<i>Hemiramphus balao</i> Lesueur, 1821	HV	Alves and Filho 1996	MS	Collette 1999

<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)	PV	Passos et al. 2013	MM	Passos et al. 2013
<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)	-	-	MS	Passos et al. 2013
<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)	OV	Passos et al. 2013	MM	Ferreira et al. 2019
<i>Ablennes hians</i> (Valenciennes, 1846)	PV	Collette 1995	MS	Sasekumar et al. 1992
<i>Strongylura marina</i> (Walbaum, 1792)	PV	Passos et al. 2013	MM	Passos et al. 2013
<i>Strongylura timucu</i> (Walbaum, 1792)	PV	Passos et al. 2013	MM	Passos et al. 2013
<i>Tylosurus acus acus</i> (Lacepède, 1803)	PV	Contente et al. 2009	MS	Ferreira et al. 2019
<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)	PV	Alves and Filho 1996	MM	Favero et al. 2019
<i>Poecilia vivipara</i> Bloch & Schneider, 1801	PV	Paiva et al., 2009	ES	Keith et al. 2000
<i>Rachycentron canadum</i> (Linnaeus, 1766)	PV	Collette 1999a	MS	Collette 1999
<i>Echeneis naucrates</i> Linnaeus, 1758	DV	Williams Jr et al., 2003	MS	Vasconcelos Filho and Oliveira 1999
<i>Phtheichthys lineatus</i> (Menziés, 1791)	OP	Heemstra 1995	MS	Heemstra 1995
<i>Remora remora</i> (Linnaeus, 1758)	PV	Alves and Filho 1996	MS	Cressey and Lachner 1970
<i>Carangoides bartholomaei</i> (Cuvier, 1833)	PV	Paiva et al. 2008	MS	Santos 2012
<i>Caranx crysos</i> (Mitchill, 1815)	PV	Sley et al. 2009	MS	Ferreira 2018
<i>Caranx hippos</i> (Linnaeus, 1766)	PV	Temóteo et al. 2015	MS	Vasconcelos Filho and Oliveira 1999
<i>Caranx latus</i> Agassiz, 1831	ZB	Passos et al. 2013	MS	Vasconcelos Filho and Oliveira 1999
<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	OP	Passos et al. 2013	MS	Vasconcelos Filho and Oliveira 1999
<i>Hemicaranx amblyrhynchus</i> (Cuvier, 1833)	ZB	Cerqueira 2018	MM	Menezes and Figueiredo 1980
<i>Oligoplites palometa</i> (Cuvier, 1832)	PV	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Oligoplites saliens</i> (Bloch, 1793)	PV	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	PV	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Selene brownii</i> (Curvier, 1816)	ZB	Bomfim 2014	MS	Ferreira et al. 2019
<i>Selene setapinnis</i> (Mitchill, 1815)	PV	Menezes and Figueiredo 1980	MS	Passos et al. 2013
<i>Selene vomer</i> (Linnaeus, 1758)	ZB	Passos et al. 2013	MM	Passos et al. 2013

<i>Trachinotus carolinus</i> (Linnaeus, 1766)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Trachinotus cayennensis</i> Cuvier, 1832	ZB	Castro et al. 2019	MM	Cervigón 1993
<i>Trachinotus falcatus</i> (Linnaeus, 1758)	ZB	Passos et al. 2013	MS	Vasconcelos Filho and Oliveira 1999
<i>Trachinotus goodei</i> Jordan & Evermann, 1896	ZB	Ferreira et al. 2019	MS	Ferreira et al. 2019
<i>Sphyraena barracuda</i> (Edwards, 1771)	PV	Ferreira et al. 2019	MM	Osório et al. 2011
<i>Sphyraena guachancho</i> Cuvier, 1829	PV	Lopes et al. 2012	MS	Ferreira 2018
<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900	ZB	Passos et al. 2013	MS	da Silva and Fabré et al. 2019
<i>Citharichthys cornutus</i> (Günther, 1880)	-	-	-	-
<i>Citharichthys macrops</i> Dresel, 1885	ZB	Passos et al., 2013	MS	Passos et al, 2013
<i>Citharichthys spilopterus</i> Günther, 1862	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Etropus crossotus</i> Jordan & Gilbert, 1882	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Etropus longimanus</i> Norman, 1933	ZB	Figueiredo and Menezes 2000	MS	Saad and Fagundes Netto 1992; Derisio et al. 2012
<i>Paralichthys brasiliensis</i> (Ranzani, 1842)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Paralichthys orbignyanus</i> (Valenciennes, 1839)	ZB	Prisco et al. 2001	MM	Prisco et al. 2001
<i>Paralichthys tropicus</i> Ginsburg, 1933	ZB	Rosas et al. 1999	MM	Reis-Filho et al. 201
<i>Syacium micrurum</i> Ranzani, 1842	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Syacium papillosum</i> (Linnaeus, 1758)	ZB	Ferreira et al. 2019	MS	Ferreira et al. 2019
<i>Bothus lunatus</i> (Linnaeus, 1758)	ZB	Robins and Ray 1986	MS	Robins and Ray 1986
<i>Bothus ocellatus</i> (Agassiz, 1831)	ZB	Ferreira et al., 201	MM	Ferreira et al. 2019
<i>Bothus robinsi</i> Topp & Hoff, 1972	ZB	Figueiredo and Menezes 2000	MS	Mendonça and Araújo 2002; Marancik et al. 2005
<i>Achirus achirus</i> (Linnaeus, 1758)	ZB	Carpenter 2003	ES	Carpenter 2003
<i>Achirus declivis</i> Chabanaud, 1940	ZB	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Achirus lineatus</i> (Linnaeus, 1758)	ZB	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009	-	-	-	-
<i>Trinectes microphthalmus</i> (Chabanaud, 1928)	OV	Favero 201)	MM	Reis-Filho et al. 2010

<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)	ZB	Carpenter 2003	MS	Reis-Filho et al. 2010
<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)	PV	Paiva et al. 2009	MM	Reis-Filho et al. 2010
<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Bryx dunckeri</i> (Metzelaar, 1919)	PV	Favero et al. 2019	MM	Favero et al. 2019
<i>Cosmocampus elucens</i> (Poey, 1868)	ZB	Fritzsche 1990	MS	Passos et al, 2013
<i>Hippocampus erectus</i> Perry, 1810	ZB	Vieira and Musick 1993	MS	Vasconcelos Filho and Oliveira 1999
<i>Hippocampus reidi</i> Ginsburg, 1933	ZP	Felício et al. 2006	MM	Osório et al. 2011
<i>Microphis brachyurus lineatus</i> (Kaup, 1856)	ZP	Frias-Torres 2004	ES	Frias-Torres 2004
<i>Microphis brachyurus brachyurus</i> (Bleeker, 1854)	ZP	Dawson 1985	AN	Gilmore and Hastings 1983
<i>Fistularia petimba</i> Lacepède, 1803	PV	Paiva et al. 2009	ES	Passos et al. 2013
<i>Fistularia tabacaria</i> Linnaeus, 1758	ZP	Passos et al. 2013	MM	Passos et al. 2013
<i>Syngnathus pelagicus</i> Linnaeus, 1758	ZP	Passos et al, 2013	MS	Passos et al. 2013
<i>Dactylopterus volitans</i> (Linnaeus, 1758)	PV	Paiva et al. 2009	MS	Passos et al. 2013
<i>Trichiurus lepturus</i> Linnaeus, 1758	PV	Vasconcelos Filho et al. 2010	MS	Vasconcelos Filho and Oliveira 1999
<i>Euthynnus alletteratus</i> (Rafinesque, 1810)	PV	Collette and Nauen 1983	MS	Collette and Nauen 1983
<i>Scomberomorus brasiliensis</i> Collette, Russo & Zavala-Camin, 1978	PV	Helfman et al. 2009	MS	Vasconcelos Filho and Oliveira 1999
<i>Scomberomorus cavalla</i> (Cuvier, 1829)	PV	da Silva and Fabré et al. 2019	MS	da Silva and Fabré et al. 2019
<i>Scomberomorus maculatus</i> (Mitchill, 1815)	PV	Collette and Nauen 1983	MS	Elliott et al. 2007
<i>Scomberomorus regalis</i> (Bloch, 1793)	PV	Helfman et al. 2009	MS	Riede 2004
<i>Peprilus paru</i> (Linnaeus, 1758)	OV	Passos et al. 2013	MS	Passos et al. 2013
<i>Astroscoptes y-graecum</i> Cuvier, 1829	PV	Cervigón 1994	MS	Grimes et al. 1982; Peterson et al. 2000; Cézár Felix et al. 2006
<i>Halichoeres poeyi</i> (Steindachner, 1867)	ZB	(Nunes et al. 2013	MS	Carpenter 2003

<i>Nicholsina usta</i> (Valenciennes, 1840)	HV	Carpenter 2003	MS	Carpenter 2003
<i>Sparisoma amplum</i> (Ranzani, 1842)	HV	Ferreira and Gonçalves 2006	MS	Moura et al. 2001
<i>Sparisoma axillare</i> (Steindachner, 1878)	HV	Ferreira and Gonçalves, 2006	MS	Moura et al. 2001
<i>Sparisoma radians</i> (Valenciennes, 1840)	HV	Paiva et al. 2008	MS	Vasconcelos Filho and Oliveira 1999
<i>Centropomus ensiferus</i> Poey, 1860	OV	Favero 2019	MM	da Silva and Fabré et al. 2019
<i>Centropomus mexicanus</i> Bocourt, 1868	PV	Keith et al. 2000	MM	Keith et al. 2000
<i>Centropomus parallelus</i> Poey, 1860	PV	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Centropomus pectinatus</i> Poey, 1860	PV	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Centropomus undecimalis</i> (Bloch, 1792)	PV	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Diapterus auratus</i> Ranzani, 1842	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Diapterus rhombeus</i> (Cuvier, 1829)	ZP	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Eucinostomus argenteus</i> Baird & Girard, 1855	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	ZB	Ferreira et al. 2019	MM	Passos et al. 2013
<i>Eucinostomus havana</i> (Nichols, 1912)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	ZB	Ferreira et al. 2019	MM	Passos et al. 2013
<i>Eugerres brasilianus</i> (Cuvier, 1830)	OV	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Gerres cinereus</i> (Walbaum, 1792)	ZB	Carpenter 2003	MM	Reis-Filho et al. 2010
<i>Ulaema lefroyi</i> (Goode, 1874)	PV	Paiva et al. 2009	MM	Favero et al. 2019
<i>Pseudupeneus maculatus</i> (Bloch, 1793)	ZB	Soares et al. 2020	MS	Vasconcelos Filho and Oliveira 1999
<i>Alphestes afer</i> (Bloch, 1793)	ZB	Randall 1967	MS	Medeiros et al. 2009; Marques and Ferreira 2011
<i>Diplectrum radiale</i> (Quoy & Gaimard, 1824)	PV	Alves and Filho 1996	MM	Reis-Filho et al. 2010
<i>Epinephelus adscensionis</i> (Osbeck, 1765)	ZB	Medeiros et al. 2010	MS	Pereira 2014
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	PV/OP	Freitas et al. 2015	MS	Paiva et al. 2009
<i>Epinephelus marginatus</i> (Lowe, 1834)	OP	Medeiros et al. 2010	MS	Andrade et al. 2003

<i>Hyporthodus nigrurus</i> (Holbrook, 1855)	ZB	Passos et al. 2013	MS	Damasceno et al. 2016
<i>Mycteroperca bonaci</i> (Poey, 1860)	PV	Daros 2014	MS	Daros 2014
<i>Mycteroperca tigris</i> (Valenciennes, 1833)	PV	Carpenter 2003	MS	Carpenter 2003
<i>Mycteroperca venenosa</i> (Linnaeus, 1758)	PV	Carpenter 2003	MS	Carpenter 2003
<i>Rypticus randalli</i> Courtenay, 1967	PV	Medeiros et al. 2017	MS	Nakayama et al. 2020
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)	ZB	Dantas et al. 2012	MS	Vasconcelos Filho and Oliveira 1999
<i>Serranus flaviventris</i> (Cuvier, 1829)	OP	Maia-Nogueira et al. 2008	MS	Lieske and Myers 1994
<i>Serranus phoebe</i> Poey, 1851	ZB	Robins and Starck 1961	MS	Cervigón et al. 1992
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	ZB	Allee et al. 2000	MS	Gasparini et al. 2005
<i>Pomacanthus paru</i> (Bloch, 1787)	ZB	Allee et al. 2000	MS	Vasconcelos Filho and Oliveira 1999
<i>Chaetodon ocellatus</i> (Bloch, 1787)	ZB	Ferreira 2018	MS	Ferreira 2018
<i>Chaetodon striatus</i> Linnaeus, 1758	ZP	Liedke et al. 2016	MS	Vasconcelos Filho and Oliveira 1999
<i>Anisotremus surinamensis</i> (Bloch, 1791)	ZB	Robins and Ray 1986	MS	Passos et al. 2013
<i>Anisotremus virginicus</i> (Linnaeus, 1758)	OV	Ferreira et al., 201	MS	Ferreira et al. 2019
<i>Conodon nobilis</i> (Linnaeus, 1758)	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Genyatremus luteus</i> (Bloch, 1790)	ZB	Passos et al. 2013	MS	Vasconcelos Filho and Oliveira 1999
<i>Haemulon aurolineatum</i> Cuvier, 1830	ZB	Pereira et al. 2015	MS	Vasconcelos Filho and Oliveira 1999
<i>Haemulon parra</i> (Desmarest, 1823)	ZB	Paiva et al. 2008	MS	Vasconcelos Filho and Oliveira 1999
<i>Haemulon plumierii</i> (Lacepède, 1801)	ZB	Ferreira et al. 2019	MS	Ferreira et al. 2019
<i>Haemulon squamipinna</i> Rocha & Rosa, 1999	ZP	Pereira et al. 2015	MS	Reis-Filho et al. 2010
<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)	ZB	Pereira et al. 2015	MM	Passos et al. 2013
<i>Orthopristis ruber</i> (Cuvier, 1830)	ZB	Cervigón 1993	MS	Cervigón 1993
<i>Pomadasys corvinaeformis</i> (Steindachner, 1868)	ZB	Denadai et al. 2013	MM	Passos et al. 2013
<i>Pomadasys crocro</i> (Cuvier, 1830)	ZB	Ferreira et al. 2019	MS	Riede 2004

<i>Pomadasys ramosus</i> Poey, 1860	ZB	Silva 2018	MM	Passos et al. 2013
<i>Lutjanus alexandrei</i> Moura & Linderman, 2007	ZB	Moraes 2012	MS	Fernandes et al. 2012
<i>Lutjanus analis</i> (Cuvier, 1828)	PV	Passos et al.,2013	MM	Passos et al. 2013
<i>Lutjanus apodus</i> (Walbaum, 1792)	ZB	Alves and Filho 1996	MS	Lieske and Myers 1994
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	ZB	Allen 1985	MS	Bastos et al. 2013
<i>Lutjanus griseus</i> (Linnaeus, 1758)	PV	Paiva et al. 2009	MS	Smith 1997
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	ZB	Monteiro et al. 2009	MS	Vasconcelos Filho and Oliveira 1999
<i>Lutjanus synagris</i> (Linnaeus, 1758)	ZB	Ferreira 2018	MS	Vasconcelos Filho and Oliveira 1999
<i>Ocyurus chrysurus</i> (Bloch, 1791)	ZB	Heck and Weinstein 1989	MS	Vasconcelos Filho and Oliveira 1999
<i>Polydactylus oligodon</i> (Gunther, 1860)	ZB	Passos et al. 2013	MS	Passos et al. 2013
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Scorpaena isthmensis</i> Meek & Hilderbrand, 1928	PV	Paiva et al. 2009	MS	Passos et al. 2013
<i>Scorpaena plumieri</i> Bloch, 1789	PV	Passos et al.,2013	MS	Edwards 1990
<i>Prionotus punctatus</i> (Bloch, 1793)	ZB	Longo et al. 2015	MS	Vasconcelos Filho and Oliveira 1999
<i>Chaetodipterus faber</i> (Broussonet, 1782)	OV	Passarone et al. 2019	MS	Passos et al. 2013
<i>Bairdiella ronchus</i> (Cuvier, 1830)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Cynoscion acoupa</i> (Lacepède, 1801)	ZB	Passos et al.,2013	MM	da Silva and Fabré et al. 2019
<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1833)	PV	da Silva and Fabré et al. 2019	MS	da Silva and Fabré et al. 2019
<i>Cynoscion leiarchus</i> (Cuvier, 1830)	PV	Alves and Filho 1996	MS	Passos et al. 2013
<i>Cynoscion microlepidotus</i> (Cuvier, 1830)	PV	da Silva and Fabré et al. 2019	ES	Passos et al. 2013
<i>Cynoscion steindachneri</i> (Jordan, 1889)	OV	Carpenter 2003	ES	Carpenter 2003
<i>Cynoscion virescens</i> (Cuvier, 1830)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)	PV	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Larimus breviceps</i> Cuvier, 1830	ZB	Passarone et al. 2019	MM	Passarone et al. 2019

<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)	PV	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Menticirrhus americanus</i> (Linnaeus, 1758)	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Menticirrhus littoralis</i> (Holbrook, 1847)	ZB	Alves and Filho 1996	MM	Passos et al. 2013
<i>Micropogonias furnieri</i> (Desmarest, 1823)	OV	(Lima et al. 2021)	MM	da Silva and Fabré et al. 2019
<i>Nebris microps</i> Cuvier, 1830	ZB	Carpenter 2003	ES	Carpenter 2003
<i>Odontoscion dentex</i> (Cuvier, 1830)	OV	Carpenter 2003	MS	Carpenter 2003
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Paralonchurus brasiliensis</i> (Steindachner, 1875)	ZB	Ferreira et al. 2019	MM	Ferreira et al. 2019
<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)	ZB	Froese and Pauly 2017	MM	Chao 1976
<i>Pogonias cromis</i> (Linnaeus, 1766)	ZB	Passos et al. 2013	MS	Passos et al. 2013
<i>Stellifer brasiliensis</i> (Schultz, 1945)	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Stellifer microps</i> (Steindachner, 1864)	ZB	Passarone et al. 2019	ES	Passarone et al. 2019
<i>Stellifer naso</i> (Jordan, 1889)	ZB	Silva et al. 2007	ES	Cervigón 1993
<i>Stellifer rastrifer</i> (Jordan, 1889)	ZB	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Stellifer stellifer</i> (Bloch, 1790)	ZB	Passarone et al. 2019	ES	Passarone et al. 2019
<i>Umbrina coroides</i> Cuvier, 1830	ZB	Passos et al. 2013	MS	Passos et al. 2013
<i>Acanthurus bahianus</i> Castelnau, 1855	HV	(Pimentel et al. 2018)	MS	Vasconcelos Filho and Oliveira 1999
<i>Acanthurus chirurgus</i> (Bloch, 1787)	HV	Longo et al. 2015	MS	Vasconcelos Filho and Oliveira 1999
<i>Acanthurus coeruleus</i> Bloch & Schneider, 1801	HV	Ferreira et al. 2019	MS	Ferreira et al. 2019
<i>Lobotes surinamensis</i> (Bloch, 1790)	ZB	Passos et al. 2013	MS	Vasconcelos Filho and Oliveira 1999
<i>Archosargus probatocephalus</i> (Walbaum, 1792)	ZB	Passos et al. 2013	MS	Costa et al. 2007
<i>Archosargus rhomboidalis</i> (Linnaeus, 1758)	ZB	Pauly and Yáñez-Arancibia 2012	MM	Passos et al. 2013
<i>Antennarius multiocellatus</i> (Valenciennes, 1837)	PV	Randall 1996	ES	Gasparini et al. 2005
<i>Antennarius striatus</i> (Shaw, 1794)	ZB	Nakane et al. 2011	MS	Paiva et al. 2009

<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	ZB	Gibran and Castro 1999	MS	Vasconcelos Filho and Oliveira 1999
<i>Acanthostracion quadricornis</i> (Linnaeus, 1758)	OV	Eduardo et al. 2020	MS	Carpenter 2003
<i>Lactophrys trigonus</i> (Linnaeus, 1758)	ZB	Ferreira et al. 2019	MS	Ferreira et al. 2019
<i>Lactophrys triqueter</i> (Linnaeus, 1758)	ZB	Lieske and Myers 1994	MS	Lieske and Myers 1994
<i>Balistes vetula</i> Linnaeus, 1758	ZB	Menezes 1979	MS	Schärer et al. 2010; Rivera Hernández et al. 2019
<i>Melichthys niger</i> (Bloch, 1786)	OV	Eduardo et al. 2020	MS	Carpenter 2003
<i>Aluterus heudelotii</i> Hollard, 1855	ZP	Eduardo et al. 2020	MS	de Andrade et al. 2015
<i>Aluterus monoceros</i> (Linnaeus, 1758)	OV	Eduardo et al. 2020	MS	de Andrade et al. 2015
<i>Aluterus schoepfii</i> (Walbaum, 1792)	HV	Eduardo et al. 2020	MS	de Andrade et al. 2015
<i>Cantherhines pullus</i> (Ranzani, 1842)	OV	Eduardo et al. 2020	MS	Carpenter 2003
<i>Monacanthus ciliatus</i> (Mitchill, 1818)	ZP	Eduardo et al. 2020	MS	Carpenter 2003
<i>Stephanolepis hispidus</i> (Linnaeus, 1766)	OV	Eduardo et al. 2020	ES	Passos et al. 2013
<i>Stephanolepis setifer</i> (Bennett, 1831)	OV	Eduardo et al. 2020	MS	Carpenter 2003
<i>Canthigaster figueiredoi</i> Moura & Castro, 2002	PV	Eduardo et al. 2020	MS	Carpenter 2003
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	ZB	Ferreira et al. 2019	MS	Ferreira et al. 2019
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	HV	Passarone et al. 2019	MM	Passarone et al. 2019
<i>Sphoeroides greeleyi</i> Gilbert, 1900	ZB	Ferreira et al. 2019	ES	Ferreira et al. 2019
<i>Sphoeroides nephelus</i> (Goode & Bean, 1882)	PV	Carpenter 2003	MM	Carpenter 2003
<i>Sphoeroides spengleri</i> (Bloch, 1785)	PV/ZB	Eduardo et al. 2020	ES	Favero et al. 2019
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	ZB	Ferreira et al. 2019	ES	Osório et al. 2011
<i>Sphoeroides tyleri</i> Shipp, 1972	PV	Eduardo et al. 2020	MS	de Andrade et al. 2015
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	-	-	MS	Paiva et al. 2009
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)	ZB	Almeida-Silva et al. 2015	ES	Passos et al. 2013
<i>Diodon holocanthus</i> Linnaeus, 1758	PV	Eduardo et al. 2020	MS	Mundy 2005

Diodon hystrix Linnaeus, 1758

PV

Eduardo et al.
2020

MS

Gasparini and
Floeter 2001

References

- Agostinho CS, Júlio HF (2002) Observation of an invasion of the piranha *Serrasalmus marginatus* Valenciennes, 1847 (Osteichthyes, Serrasalminidae) into the Upper Paraná River, Brazil. *Acta Scientiarum - Biological and Health Sciences* 24:391–395
- Ajemian MJ, Powers SP, Murdoch TJT (2012) Estimating the potential impacts of large mesopredators on benthic resources: Integrative assessment of spotted eagle ray foraging ecology in Bermuda. *PLoS ONE* 7:. <https://doi.org/10.1371/journal.pone.0040227>
- Allee RJ, Dethier MN, Brown D, et al (2000) Marine and estuarine ecosystem and habitat classification. NOAA Technical Memorandum 51
- Allen GR (1985) FAO Species catalogue Volume 6. Snappers of the world. An annotated and illustrated catalogue of lutjanid species known to date. FAO Spec. Cat. Fish. Purp, Rome
- Almeida-Silva PH, Tubino RA, Zambrano LC, et al (2015) Trophic ecology and food consumption of fishes in a hypersaline tropical lagoon. *Journal of Fish Biology* 86:1781–1795. <https://doi.org/10.1111/jfb.12689>
- Alves MIM, Filho AAS (1996) Peixes do estuário do rio Jaguaribe (Ceará - Brasil): Aspectos Fisiocológicos. *Ciencia Agronomica* 27:5–16
- Andrade ÁB, Machado LF, Hostim-Silva M, Barreiros JP (2003) Reproductive Biology of *Epinephelus marginatus* (Lowe, 1834). *Brazilian Archives of Biology and Technology* 46:373–381. <https://doi.org/10.1590/S1516-89132003000300009>
- Baldwin CC, Smith DG (2003) Larval gobiidae (Teleostei: Perciformes) of Carrie Bow Cay, Belize, central America. *Bulletin of Marine Science* 72:639–674
- Barletta M, Dantas D (2016) Biogeography and Distribution of Mugilidae in the Americas. Taylor & Francis
- Barreto SB, Silva AT, Souza FB, Jucá-Chagas R (2018) Diet of *Hemigrammus marginatus* (Characiformes: Characidae) in the Upper Contas River, Diamantina Plateau (Bahia, Brazil). *Iheringia Série Zoologia* 108:1–8. <https://doi.org/10.1590/1678-4766e2018036>
- Bastos RF, Condini MV, Garcia AM (2013) Fish species list of coastal streams in southern Brazil, with notes on austral distribution limits of marine and freshwater endangered species. *Pan-American Journal of Aquatic Sciences* 8:347–351
- Bomfim A da C (2014) Bioecologia da ictiofauna marinha descartada pelo arrasto camaroeiro em praias da Bacia Potiguar, Brasil. Universidade Federal do Rio Grande do Norte
- Bornbusch AH (1988) Gill Raker Morphologies of Anchovies (Teleostei: Engraulidae) from the Río Orinoco, Venezuela. *Copeia* 174–182
- Boujard T, Pascal M, Meunier FJ, le Bail PY (1997) Poissons de Guyane. Guide écologique de l'Approuague et de la réserve des Nouragues. Institut National de la Recherche Agronomique, Paris
- Burgess WE (1989) An atlas of freshwater and marine catfishes. A preliminary survey of the Siluriformes. T.F.H. Publications, New Jersey
- Carpenter KE (2003) The living marine resources of the Western Central Atlantic. Volume 2: Bony fishes part 1 (Acipenseridae to Grammatidae). FAO Species Identification Guide for Fishery Purposes and American Society of Ichthyologists and Herpetologists Special Publication No. 5., Virginia
- Carvalho-Filho A (1999) Peixes: costa Brasileira. Melro, São Paulo
- Castro ACL, Azevedo JWJ, Ferreira HRS, et al (2019) Feeding activity of the cayenne pompano *Trachinotus cayennensis* (Cuvier 1832) (perciformes, carangidae) in estuaries on the western coast of the state of maranhão, Brazil. *Brazilian Journal of Biology* 79:311–320. <https://doi.org/10.1590/1519-6984.182683>
- Castro MF de (2005) Abundância, Distribuição e Desenvolvimento das fases iniciais de peixes no estuário do Rio Formoso, Pernambuco - Brasil

- Castro JI (1996) BIOLOGY OF THE BLACKTIP SHARK , *CARCHARHINUS LIMBATUS* , 1841), is a cosmopolitan species found throughout tropical and subtropical waters (Bigelow and Schroeder , 1948). It ranges along the eastern coast of North America from New England to the Florida. 59:508–522
- Cerqueira NNCD (2018) Análise comparativa da composição genética de exemplares da fauna de peixes marinho-estuarinos encontrados na costa do Brasil
- Cervigón F (1993) Los peces marinos de Venezuela. Volume 2. Fundación Científica Los Roques, Caracas
- Cervigón F (1994) Los peces marinos de Venezuela. Volume 3. Fundación Científica Los Roques, Caracas
- Cervigón F, Cipriani R, Fischer W, et al (1992) Fichas FAO de identificación de especies para los fines de la pesca. Guía de campo de las especies comerciales marinas y de aguas salobres de la costa septentrional de Sur América. Preparado con el financiamiento de la Comisión de Comunidades Europeas y de. FAO, Rome
- Cézar Felix F, Louis Spach H, Werner Hackradt C, et al (2006) Abundância sazonal e a composição da assembléia de peixes em duas praias estuarinas da Baía de Paranaguá, Paraná. *Revista Brasileira de Zootecias* 8:35–47
- Chao LN (1976) Aspects of systematics, morphology, life history and feeding of western Atlantic sciaenidae (pisces : perciformes)
- Claro R (1994) Ecología de los peces marinos de Cuba. Instituto de Oceanología Académica de Ciencias de Cuba, CIQRO, Talleres Ferrándiz, Cuba
- Collette BB (2003a) *Batrachoididae*. In: *The Living Marine Resources of the Western central Atlantic*. FAO, Rome, pp 1026–1042
- Collette BB (1999a) *Rachycentridae*. In: *FAO Species Identification Guide for Fishery Purposes*. FAO, Rome, pp 98–99
- Collette BB (1999b) *Belonidae*. In: *The Living Marine Resources of the Western central Atlantic Pacific*. FAO, Rome, pp 2151–2161
- Collette BB (2003b) Family *Belonidae* Bonaparte 1832 - needlefishes. *California Academy of Sciences Annotated Checklists of Fishes* 1–22
- Collette BB, Nauen CE (1983) *FAO Species Catalogue: Vol. 2 Scombrids of the World*
- Compagno L (1984) Sharks of the world. An annotated and illustrated catalogue of shark species known to date. Part 2 - *Carchahiniformes*. In: *FAO Species Catalogue*. FAO Fish. Synop., Rome, pp 251–655
- Compagno LJ v. (2001) Sharks of the world. An annotated and illustrated catalogue of shark species known to date. In: Bullhead, mackerel and carpet sharks (*Heterodontiformes*, *Lamniformes* and *Orectolobiformes*). *FAO Species Catalogue for Fishery Purposes*, Rome, pp 188–191
- Contente RF, Stefanoni MF, Spach HL (2009) Size-related changes in diet of the slipper sole *Trinectes paulistanus* (*Actinopterygii*, *Achiridae*) juveniles in a subtropical Brazilian estuary. *Pan-American Journal of Aquatic Sciences* 4:63–69
- Cordiviola de Yuan E, Pignalberi de Hassan C (1985) Fish population in the Paraná River: lentic environments of Diamante and San Pedro areas (Argentine Republic). *Hydrobiologia* 127:213–218
- Costa Novaes JL, Carvalho ED (2011) Population structure and stock assessment of *Hoplias malabaricus* (characiformes: *Erythrinidae*) caught by artisanal fishermen in river-reservoir transition area in Brazil. *Revista de Biologia Tropical* 59:71–83. <https://doi.org/10.15517/rbt.v59i1.3179>
- Costa RC, Fransozo A, Freire FAM, Castilho AL (2007) Abundance and Ecological Distribution of the “Sete-Barbas” Shrimp *Xiphopenaeus kroyeri* (Heller, 1862) (Decapoda: Penaeoidea) in Three Bays of the Ubatuba Region, Southeastern Brazil. *Gulf and Caribbean Research* 19:33–41. <https://doi.org/10.18785/gcr.1901.04>
- Cressey RF, Lachner EA (1970) The Parasitic Copepod Diet and Life History of Diskfishes (*Echeneidae*). *Copeia* 1970:310–318. <https://doi.org/10.2307/1441652>
- Cruz-Escalona VH, Peterson MS, Campos-Dávila L, Zetina-Rejón M (2005) Feeding habits and trophic morphology of inshore lizardfish (*Synodus foetens*) on the central continental shelf off Veracruz, Gulf of Mexico. *Journal of Applied Ichthyology* 21:525–530. <https://doi.org/10.1111/j.1439-0426.2005.00651.x>

- Cuvier M, Valenciennes MA (1840) Histoire Naturelle des Poissons. Breger Levrault, Paris
- da Silva VEL, Fabré NN (2019) Rare Species Enhance Niche Differentiation Among Tropical Estuarine Fish Species. *Estuaries and Coasts* 42:890–899. <https://doi.org/10.1007/s12237-019-00524-2>
- Damasceno JS, Siccha-Ramirez R, Oliveira C, et al (2016) Molecular identification of Atlantic goliath grouper *epinephelus itajara* (Lichtenstein, 1822) (Perciformes: Epinephelidae) and related commercial species applying multiplex PCR. *Neotropical Ichthyology* 14:. <https://doi.org/10.1590/1982-0224-20150128>
- Dantas NCF de M, Feitosa CV, Araújo ME de (2012) Composition and assemblage structure of demersal fish from São Cristóvão beach, Areia Branca, RN. *Biota Neotropica* 12:108–117. <https://doi.org/10.1590/s1676-06032012000300012>
- Daros FAL de M (2014) Estudo da ictiofauna costeira no litoral do Paraná e adjacências através de censos visuais e microquímica de otólitos. Universidade Federal do Paraná
- Dawson CE (1971) Supplemental Observations on *Gunterichthys longipenis*, a Northern Gulf of Mexico Brotulid Fish. *Copeia* 1967:164–167. <https://doi.org/10.2307/1442254>
- Dawson CE (1985) Indo-Pacific pipefishes (Red Sea to the Americas). The Gulf Coast Research Laboratory, Ocean Springs
- de Andrade AC, Santos SR, Verani JR, Vianna M (2015) Guild composition and habitat use by Tetraodontiformes (Teleostei, Acanthopterygii) in a south-western Atlantic tropical estuary. *Journal of the Marine Biological Association of the United Kingdom* 1–14. <https://doi.org/10.1017/S0025315415001368>
- de Moor IJ, Bruton MN (1988) Atlas of alien and translocated indigenous aquatic animals in southern Africa. National Scientific Programmes Unit: CSIR, SANSP Report 144
- Denadai MR, Bessa E, Santos FB, et al (2012) Life history of three catfish species (Siluriformes: Ariidae) from southeastern Brazil. *Biota Neotrop* 12:
- Derisio C, Betti P, de Astarloa JMD, Machinandiarena L (2012) Desarrollo larval de *Etropus longimanus* (Paralichthyidae) y *Symphurus trewavasae* (Cynoglossidae) en la costa Bonaerense, Argentina. *Scientia Marina* 76:29–37. <https://doi.org/10.3989/scimar.2012.76n1029>
- di Dario F (2013) Pristigasteridae. In: Peixes do Rio Madeira. SantoAntonio Energia, São Paulo, pp 100–107
- Ebert DA, Fowler S, Compagno L (2013) Sharks of the World. Wid Nature Press, Plymouth
- Eduardo LN, Bertrand A, Frédou T, et al (2020) Biodiversity, ecology, fisheries, and use and trade of Tetraodontiformes fishes reveal their socio-ecological significance along the tropical Brazilian continental shelf. *Aquatic Conservation: Marine and Freshwater Ecosystems* 1:1–14. <https://doi.org/10.1002/aqc.3278>
- Edwards A (1990) Fish and Fisheries of Saint Helena Island. Centre for Tropical Coastal Management Studies, Newcastle
- Elliott M, Whitfield AK, Potter IK, et al (2007) The guild approach to categorizing estuarine fish assemblages: a global review. *Fish and Fisheries* 8:241–268. <https://doi.org/10.1111/j.1467-2679.2007.00253.x>
- Emery AR (1978) Pomacentridae. In: FAO Species Identification sheets for fishery purposes. FAO, Rome, p pag.var.
- Favero F de LT (2019) Diversidade funcional da ictiofauna da zona de arrebentação de Jaguaribe, Itamaracá, litoral norte de Pernambuco
- Favero F de LT, Araujo IM da S, Severi W (2019) Structure of the fish assemblage and functional guilds in the estuary of Maracápe, Northeastern coast of Brazil. *Boletim do Instituto de Pesca* 45:417. <https://doi.org/10.20950/1678-2305.2019.45.1.417>
- Felício AKC, Rosa IL, Souto A, Freitas RHA (2006) Feeding behavior of the longsnout seahorse *Hippocampus reidi* Ginsburg, 1933. *Journal of Ethology* 24:219–225. <https://doi.org/10.1007/s10164-005-0189-8>
- Fernandes CAF, de Oliveira PGV, Travassos PEP, Hazin FHV (2012) Reproduction of the Brazilian snapper, *Lutjanus alexandrei* Moura & Lindeman, 2007 (Perciformes: Lutjanidae), off the northern coast of Pernambuco, Brazil. *Neotropical Ichthyology* 10:587–592. <https://doi.org/10.1590/S1679-62252012005000022>
- Ferreira CEL, Gonçalves JEA (2006) Community structure and diet of roving herbivorous reef fishes in the Abrolhos Archipelago, south-western Atlantic. *Journal of Fish Biology* 69:1533–1551. <https://doi.org/10.1111/j.1095-8649.2006.01220.x>

- Ferreira FS, Vicentin W, Costa FE dos S, Suárez YR (2014) Trophic ecology of two piranha species, *Pygocentrus nattereri* and *Serrasalmus marginatus* (Characiformes, Characidae), in the floodplain of the Negro River, Pantanal. *Acta Limnologica Brasiliensia* 26:381–391. <https://doi.org/10.1590/s2179-975x2014000400006>
- Ferreira V (2018) Estrutura Trófica da Ictiofauna Estuarina e Marinha do Complexo Itapissuma/Itamaracá, Norte de Pernambuco, Brasil. Universidade Federal Rural de Pernambuco
- Ferreira V, le Loc'h F, Ménard F, et al (2019) Composition of the fish fauna in a tropical estuary: the ecological guild approach. *Scientia Marina* 83:133. <https://doi.org/10.3989/scimar.04855.25a>
- Figueiredo JL, Menezes NA (2000) Manual de Peixes Marinhos do Sudeste do Brasil. Museu de Zoologia- USP, São Paulo
- Finley RJ, Forrester GE (2003) Impact of ectoparasites on the demography of a small reef fish. *Marine Ecology Progress Series* 248:305–309. <https://doi.org/10.3354/meps248305>
- Fischer W, Souza I, Silva C, et al (1990) Fichas FAO de identificação de espécies para actividades de pesca. Guia de campo das espécies comerciais marinhas e de águas salobras de Moçambique. Publicação preparada em colaboração com o Instituto de Investigação Pesqueira de Moçambique, com financia. FAO, Rome
- Freitas MO, Abilhoa V, Giglio VJ, et al (2015) Diet and reproduction of the goliath grouper, *Epinephelus itajara* (Actinopterygii: Perciformes: Serranidae), in Eastern Brazil. *Acta Ichthyologica et Piscatoria* 45:1–11. <https://doi.org/10.3750/AIP2015.45.1.01>
- Freitas RHA, Aguiar AA, Freitas AKCHA, et al (2019) Unravelling the foraging behavior of the southern stingray, *Hypanus americanus* (Myliobatiformes: Dasyatidae) in a southwestern atlantic MPA. *Neotropical Ichthyology* 17:. <https://doi.org/10.1590/1982-0224-20180131>
- Frias-Torres S (2004) Notes on Aquarium Brood Release and Feeding of the Opossum Pipefish, *Microphis brachyurus lineatus*. *Gulf and Caribbean Research* 16:73–75. <https://doi.org/10.18785/gcr.1601.11>
- Fritzsche RA (1990) Check-list of the fishes of the eastern tropical Atlantic (CLOFETA). JNICT, Lisboa
- Froese R, Pauly D (2018) Fishbase. In: FishBase
- Froese R, Pauly D (2017) FishBase. World Wide electronic publication. www.fishbase.org
- Gasparini JL, Floeter SR (2001) The shore fishes of Trindade Island, western South Atlantic. *Journal of Natural History* 35:1639–1656. <https://doi.org/10.1080/002229301317092379>
- Gasparini JL, Floeter SR, Sazima I (2005) Marine Ornamental Trade in Brazil. *Biodiversity and Conservation* 14:2883–2899. <https://doi.org/10.1007/s10531-004-0222-1>
- Gibran FZ, Castro RMC (1999) Activity, feeding behaviour and diet of *Ogcocephalus vespertilio* in southern west Atlantic. *Journal of Fish Biology* 55:588–595. <https://doi.org/10.1006/jfbi.1999.1019>
- Gilmore RG, Hastings PA (1983) Observations on the ecology and distribution of certain tropical peripheral fishes in Florida. *Florida Scientist* 46:31–51
- Goulding M (1981) Man and fisheries on an Amazon frontier. In: *Developments in Hydrobiology*. W. Tunk Publishers, p 137
- Greenfield DW, Johnson RK (1990) Heterogeneity in habitat choice in cardinalfish community structure. *Copeia* 1107–1114
- Greenfield DW, Thomerson JE (1997) *Fishes of the continental waters of Belize*. University Press of Florida, Florida
- Grimes CB, Manooch CS, Huntsman GR (1982) Reef and Rock Outcropping Fishes of the Outer Continental Shelf of North Carolina and South Carolina, and Ecological Notes on the Red Porgy and Vermilion Snapper. *Bulletin of Marine Science* 32:277–289. <https://doi.org/10.1017/CBO9781107415324.004>
- Guedes APP, Araújo FG, Pessanha ALM, Milagre RR (2015) Partitioning of the feeding niche along spatial, seasonal and size dimensions by the fish community in a tropical Bay in Southeastern Brazil. *Marine Ecology* 36:38–56. <https://doi.org/10.1111/maec.12115>
- Haimovici M, Klippel S (1999) Diagnóstico da Biodiversidade dos Peixes Teleósteos Demersais Marinhos e Estuarinos do Brasil. 68

- Harrison IJ, Miller PJ, Pezold F (2003) Microdesmidae. In: The fresh and brackish water fishes of West Africa Volume 2. Muséum national d'histoire naturelle, Paris, pp 667–669
- Heck KL, Weinstein MP (1989) Feeding Habits of Juvenile Reef Fishes Associated with Panamanian Seagrass Meadows. *Bulletin of Marine Science* 45:629–636
- Heemstra PC (1995) Additions and corrections for the 1995 impression. In: Revised Edition of Smiths' Sea Fishes. Springer-Verlag Berlin Heidelberg, Berlin, pp v–xv
- Helfman GS, Collette BB, Facey DE, Bowen BW (2009) The diversity of fishes: biology, evolution, and ecology. John Wiley & Sons, Malaysia
- Herbert J (2015) The Online Guide to the Animals of Trinidad and Tobago. In: Aspreto aspreto. [https://sta.uwi.edu/fst/lifesciences/sites/default/files/lifesciences/images/Aspreto aspreto - Banjo Catfish.pdf](https://sta.uwi.edu/fst/lifesciences/sites/default/files/lifesciences/images/Aspreto%20aspreto%20-%20Banjo%20Catfish.pdf). Accessed 9 Mar 2021
- Keith P, Bail PY le, Planquette P (2000) Atlas des poissons d'eau douce de Guyane. Tome 2, Fascicule I: Batrachoidiformes, Mugiliformes, Beloniformes, Cyprinodontiformes, Synbranchiformes, Perciformes, Pleuronectiformes, Tetraodontiformes. *Publications scientifiques du Muséum national d'Histoire naturelle* 43:286
- Kenny JS (1995) Views from the bridge: a memoir on the freshwater fishes of Trinidad. JS Kenny Maracas, St Joseph Trinidad and Tobago, St. Joseph
- Krumme U, Keuthen H, Barletta M, et al (2005) Contribution to the feeding ecology of the predatory wingfin anchovy *Pterengraulis atherinoides* (L.) in north Brazilian mangrove creeks. *Journal of Applied Ichthyology* 21:469–477
- Krumme U, Saint-Paul U, Rosenblatt H (2004) Tidal and diel changes in the structure of a nekton assemblage in small intertidal mangrove creeks in northern Brazil. *Aquatic Living Resources* 17:215–229
- Last PR, Naylor GJP, Manjaji-Matsumoto BM (2016) A revised classification of the family Dasyatidae (Chondrichthyes: Myliobatiformes) based on new morphological and molecular insights. *Zootaxa* 4139:345–368. <https://doi.org/10.11646/zootaxa.4139.3.2>
- le Bail PYP, Keith P, Planquette P (2000) Atlas des poissons d'eau douce de Guyane. Tome 2, Fascicule II: Siluriformes. Collection Patrimoines Naturels. *Publications scientifiques du Muséum national d'Histoire naturelle, Paris*
- Leopold M (2004) Poissons de Mer de Guyane: Guide Illustre. Ifremer, Paris
- Liedke AMR, Barneche DR, Ferreira CEL, et al (2016) Abundance, diet, foraging and nutritional condition of the banded butterflyfish (*Chaetodon striatus*) along the western Atlantic. *Marine Biology* 163:1–13. <https://doi.org/10.1007/s00227-015-2788-4>
- Lieske E, Myers R (1994) Collins Pocket Guide. Coral reef fishes. Indo-Pacif & Caribbean including the Red Sea. HarperCollins Publishers, New York
- Lima RS, Neto A, Lira A, et al (2021) Fish bycatch caught by shrimp fisheries in Western Tropical Atlantic. CEPE, Recife
- Londoño-Burbano A, Urbano-Bonilla A, Thomas MR (2020) *Loricaria cuffyi* (Siluriformes: Loricariidae), a new species of loricariin catfish from the Guiana Shield. *Journal of Fish Biology* 98:154–167
- Longo GO, Morais RA, Martins CDL, et al (2015) Between-habitat variation of benthic cover, reef fish assemblage and feeding pressure on the benthos at the only atoll in South Atlantic: Rocas atoll, NE Brazil. *PLoS ONE* 10:1–29. <https://doi.org/doi.org/10.1371/journal.pone.0127176>
- Lopes PRD, Oliveira-Silva ;, Rocha ;, et al (2012) Notas sobre a alimentação de *Sphyaena guachancho* Cuvier, 1829 (Actinopterygii: Sphyaenidae) na praia do malhado, Ilhéus (Bahia)
- Luiz OJ, Carvalho-Filho A, Ferreira CEL, et al (2008) The reef fish assemblage of the Laje de Santos Marine State Park, Southwestern Atlantic: Annotated checklist with comments on abundance, distribution, trophic structure, symbiotic associations, and conservation. *Zootaxa* 25:1–25. <https://doi.org/10.11646/zootaxa.1807.1.1>
- Maia-Nogueira R, de Anchieta Nunes JC, C Coni EO, et al (2008) The twinspace bass *Serranus flaviventris* (Serranidae) as follower of the goldspotted eel *Myrichthys ocellatus* (Ophichthidae) in north-eastern Brazil, with notes on others serranids. *JMBA - Biodiversity Records* 1–3

- Marancik KE, Clough LM, Hare JA (2005) Cross-shelf and seasonal variation in larval fish assemblages on the southeast United States continental shelf off the coast of Georgia. *Fishery Bulletin* 103:108–129
- Marques S, Ferreira BP (2011) Sexual development and reproductive pattern of the Mutton hamlet, *Alphestes afer* (Teleostei: Epinephelidae): A dyandric, hermaphroditic reef fish. *Neotropical Ichthyology* 9:547–558. <https://doi.org/10.1590/S1679-62252011005000026>
- Medeiros APM de, Xavier JH de A, Rosa IM de L (2017) Diet and trophic organization of the fish assemblage from the Mamanguape River Estuary, Brazil. *Latin American Journal of Aquatic Research* 45:879–890. <https://doi.org/10.3856/vol45-issue5-fulltext-2>
- Medeiros DV, Nunes J de ACC, Sampaio CLS (2009) A mutton hamlet *Alphestes afer* (Bloch, 1793) reproductive event in northeast Brazil. *Pan-American Journal of Aquatic Sciences* 4:212–215
- Medeiros PR, Gempel RG, Souza AT, et al (2010) Non-random reef use by fishes at two dominant zones in a tropical, algal-dominated coastal reef. *Environmental Biology of Fishes* 87:237–246. <https://doi.org/10.1007/s10641-010-9593-1>
- Melo BF (2011) Genética de populações de *Prochilodus argenteus* e *P. costatus* do médio São Francisco
- Mendes FLS (1999) Alimentação, Distribuição Espacial de *Arius* (Siluriformes: Ariidae) do Estuário Amazônico
- Mendonça P, Araújo FG (2002) Composição das populações de linguados (Osteichthyes, Pleuronectiformes) da Baía de Sepetiba, Rio de Janeiro, Brasil. *Revista Brasileira de Zoologia* 19:339–347
- Menezes MF de (1979) Aspectos da biologia e biometria do cangulo, *Balistes vetula* Linnaeus, no Nordeste do Brasil. *Arq Ciên Mar* 19:57–68
- Menezes N, Figueiredo JL (1980) Manual de peixes marinhos do sudeste do Brasil. IV. Teleostei (3). Museu de Zoologia- USP, São Paulo
- Michael SW (1993) Reef sharks and rays of the world. A guide to their identification, behavior, and ecology. Sea Challengers, Monterey
- Miller MJ (2002) The distribution and ecology of *Ariosoma balearicum* (Congridae) leptocephali in the western North Atlantic. *Environmental Biology of Fishes* 63:235–252. <https://doi.org/10.1023/A:1014311429809>
- Miller RR (2005) *Freshwater Fishes of Mexico*. The University of Chicago Press, Chicago
- Miller TJ (1987) Knotting: A previously undescribed feeding behavior in muraenid eels. *American Society of Ichthyologists and Herpetologists* 1055–1057
- Mills D, Vevers G (1989) *The Tetra encyclopedia of freshwater tropical aquarium fishes*. Tetra Press, New Jersey
- Mol JHA (1958) *The freshwater fishes of Suriname*. Brill, Boston
- Monteiro DP, Giarrizzo T, Isaac V (2009) Feeding ecology of juvenile dog snapper *Lutjanus jocu* (Bloch and Schneider, 1801) (Lutjanidae) in intertidal mangrove creeks in Curuçá estuary (Northern Brazil). *Brazilian Archives of Biology and Technology* 52:1421–1430. <https://doi.org/10.1590/S1516-89132009000600014>
- Monteiro-Neto C, Bertoni ÁA, Chaves LDCT, et al (2013) Checklist of marine fish from coastal islands of Rio de Janeiro, with remarks on marine conservation. *Marine Biodiversity Records* 6:1–13. <https://doi.org/10.1017/S1755267213000973>
- Moraes RLG de (2012) Pesca, parasitismo e dieta alimentar da baúna *Lutjanus alexandrei* Moura & Lindeman, 2007 nos ambientes costeiros do litoral sul de Pernambuco. Universidade Federal de Pernambuco
- Moura RL, Figueiredo JL, Sazima I (2001) A new parrotfish (Scaridae) from Brazil, and revalidation of *Sparisoma amplum* (Ranzani, 1842), *Sparisoma frondosum* (Agassiz, 1831), *Sparisoma axillare* (Steindachner, 1878) and *Scarus trispinosus* Valenciennes, 1840. *Bulletin of Marine Science* 68:505–524
- Mundy BC (2005) *Checklist of the Fishes Hawaiian Archipelago*. Bishop Museum, Honolulu
- Nakamura I, Inada T, Takeda M, Hatanaka H (1986) Important fishes trawled off Patagonia. Japan Marine Fishery Resource Research Center, Tokyo
- Nakane Y, Suda Y, Sano M (2011) Food habits of fishes on an exposed sandy beach at Fukiagehama, South-West Kyushu Island, Japan. *Helgoland Marine Research* 65:123–131. <https://doi.org/10.1007/s10152-010-0208-1>

- Nakayama P, Peret AC, Adelar-Alves J, et al (2020) Environmental drivers of fish assemblages from the shallow infralittoral areas of the Paranaguá bay, Southern Brazil. *Oecologia Australis* 24:917–927. <https://doi.org/10.4257/oeco.2020.2404.14>
- Nizinski MS, Munroe TA (2002) Order Clupeiformes, Engraulididae. In: *FAO Species Identification Guide for Fishery Purposes; American Society of Ichthyologists and Herpetologists Special Publication No. 5. FAO Species Identification Guide for Fishery Purposes and American Society of Ichthyologists and Herpetologists Special Publication No. 5.*, Rome
- Nordlie FG (1981) Feeding and reproductive biology of eleotrid fishes in a tropical estuary. *Journal of Fish Biology* 18:97–110. <https://doi.org/10.1111/j.1095-8649.1981.tb03764.x>
- Nunes J de ACC, Sampaio CLS, Barros F (2013) How wave exposure, group size and habitat complexity influence foraging and population densities in fishes of the genus *Halichoeres* (Perciformes: Labridae) on tropical rocky shores. *Marine Biology* 160:2383–2394. <https://doi.org/10.1007/s00227-013-2233-5>
- Oliveira JCS, Isaac VJ (2013) Diet Breadth and Niche Overlap Between *Hypostomus plecostomus* (Linnaeus, 1758) and *Hypostomus emarginatus* (Valenciennes, 1840) (Siluriformes) in the Coaracy Nunes Hydroelectric Reservoir, Ferreira Gomes, Amapá-Brazil. *Biota Amazônia* 3:116–125. <https://doi.org/10.18561/2179-5746/biotaamazonia.v3n2p116-125>
- Osório FM, Godinho WO, Lotufo TM da C (2011) Ictiofauna associada às raízes de mangue do estuário do Rio Pacoti - CE, Brasil. *Biota Neotropica* 11:415–420. <https://doi.org/10.1590/s1676-06032011000100038>
- Oxenford HA (2007) Summary of the importance of flyingfish fisheries in the eastern Caribbean. In: *The biology and management of eastern Caribbean flyingfish*. Centre for Resource Management and Environmental Studies University of the West Indies, Barbados, pp 47–48
- Paiva A, Lima M, Souza J, Araújo M (2009) Spatial distribution of the estuarine ichthyofauna of the Rio Formoso (Pernambuco, Brazil), with emphasis on reef fish. *Zoologia* 26:266–278
- Paiva ACG de, Tarso P de, Chaves C, Araújo ME de (2008) Estrutura e organização trófica da ictiofauna de águas rasas em um estuário tropical. *Revista Brasileira de Zoologia* 25:647–661
- Passarone R, Aparecido KC, Eduardo LN, et al (2019) Ecological and conservation aspects of bycatch fishes: An evaluation of shrimp fisheries impacts in northeastern Brazil. *Brazilian Journal of Oceanography* 67:1–10. <https://doi.org/10.1590/s1679-87592019029106713>
- Passos AC dos, Contente RF, Abatepaulo FV, et al (2013) Analysis of fish assemblages in sectors along a salinity gradient based on species, families and functional groups. *Brazilian Journal of Oceanography* 61:251–264. <https://doi.org/10.1590/S1679-87592013000400006>
- Pauly D, Yáñez-Arancibia A (2012) Ecology, biology and population dynamics of *Archosargus rhomboidalis* (Pisces, Sparidae) in atropical coastal lagoon system, southern Gulf of Mexico
- Peralta-Meixueiro MA, Vega-Cendejas ME (2011) Spatial and temporal structure of fish assemblages in a hyperhaline coastal system: Ría Lagartos, Mexico. *Neotropical Ichthyology* 9:673–682. <https://doi.org/10.1590/s1679-62252011005000033>
- Pereira PHC (2014) “Swallowing it all” – Extreme ingestion capability of juvenile reef fish. *Biota Neotropica* 14:1–2. <https://doi.org/10.1590/s1676-06020140071>
- Pereira PHC, Barros B, Zemoi R, Ferreira BP (2015) Ontogenetic diet changes and food partitioning of *Haemulon* spp. coral reef fishes, with a review of the genus diet. *Reviews in Fish Biology and Fisheries* 25:245–260. <https://doi.org/10.1007/s11160-014-9378-2>
- Peterson MS, Comyns BH, Hendon JR, et al (2000) Habitat use by early life-history stages of fishes and crustaceans along a changing estuarine landscape: Differences between natural and altered shoreline sites. *Wetlands Ecology and Management* 8:209–219. <https://doi.org/10.1023/A:1008452805584>
- Piedras SRN, Pouey JLOF (2005) Alimentação do peixe-rei (*Odontesthes bonariensis*, Atherinopsidae) nas lagoas Mirim e Mangueira, Rio Grande do Sul, Brasil. *Iheringia Série Zoologia* 95:117–120. <https://doi.org/10.1590/s0073-47212005000200001>
- Pimentel CR, Soares LSH, Macieira RM, Joyeux JC (2018) Trophic relationships in tidepool fish assemblages of the tropical Southwestern Atlantic. *Marine Ecology* 39:1–11. <https://doi.org/10.1111/maec.12496>
- Planquette P, Keith P, Bail P-Y le (1996) *Atlas des Poissons d’Eau Douce de Guyane*. Institut d’Ecologie et de Gestion de la Biodiversité, Paris

- Prisco AR, de La Rosa SGB, Astrarloa JMD de (2001) Feeding ecology of flatfish juveniles (Pleuronectiformes) in Mar Chiquita coastal lagoon (Buenos Aires, Argentina). *Estuaries* 24:917–925. <https://doi.org/10.2307/1353182>
- Rahaman F (2015) *Cheilopogon melanurus* (Atlantic Flyingfish). UWI Ecology: The Online Guide to the Animals of Trinidad and Tobago 4
- Randall JE (1996) Caribbean reef fishes. Third Edition - revised and enlarged. T.F.H. Publications, Hong Kong
- Randall JE (1967) Food habits of reef fishes of the West Indies. *Studies in Tropical Oceanography* 5:665–847
- Randall JE (2004) Food Habits of Reef Fishes of the West Indies. *Stud Trop Oceanogr* 5:665–847
- Reis-Filho JA (2011) Efeito da perda de habitat natural sobre a ictiofauna de áreas rasas em um estuário tropical
- Reis-Filho JA, Nunes J de AC da C, Ferreira A (2010) Estuarine ichthyofauna of the Paraguaçu River, Todos os Santos Bay, Bahia, Brazil. *Biota Neotropica* 10:301–311. <https://doi.org/10.1590/s1676-06032010000400034>
- Ribeiro VMA, Santos GB, Bazzoli N (2007) Reproductive biology of *Steindachnerina insculpta* (Fernandez-Yépez) (Teleostei, Curimatidae) in Furnas reservoir, Minas Gerais, Brazil. *Revista Brasileira de Zoologia* 24:71–76
- Riede K (2004) Global Register of Migratory Species - from Global to Regional Scales. Final Report of the R&D-Project 808 05 081. Federal Agency for Nature Conservation, Bonn, Germany. Bonn
- Rivera Hernández JM, Peña Alvarado N, Correa Vélez K, et al (2019) Queen Triggerfish Reproductive Biology in U.S. Caribbean Waters. *Transactions of the American Fisheries Society* 148:134–147. <https://doi.org/10.1002/tafs.10124>
- Robins CR, Ray GC (1986) A field guide to Atlantic coast fishes of North America. Houghton Mifflin Company 368
- Robins R, Starck WA (1961) Materials for a revision of *Serranus* and related fish genera. *Proceedings of the Academy of Natural Sciences of Philadelphia* 113:259–314. <https://doi.org/10.1038/139103c0>
- Rosas J, Arana D, Cabrera T, et al (1999) The potential use of the Caribbean flounder *Paralichthys tropicus* as an aquaculture species. *Aquaculture* 176:51–54. [https://doi.org/10.1016/S0044-8486\(99\)00049-6](https://doi.org/10.1016/S0044-8486(99)00049-6)
- Ross S, Casazza T, Quattrini A, Sulak K (2007) Aguiliform larvae collected off North Carolina. *Marine Biology* 150:681–695
- Ross SW, Rhode FC (2004) The gobioid fishes of North Carolina (Pisces: Gobioidi). *Bulletin of Marine Science* 74:287–323
- Ross SW, Rhode FC (2003) Collections of ophichthid eels on the surface at night off North Carolina. *Bulletin of Marine Science* 72:241–246
- Saad AM, Fagundes Netto E (1992) Aspectos da biologia reprodutiva de fêmeas de *Etropus longimanus* Norman, 1933 (Bothidae) da região de Cabo Frio, Rio de Janeiro: 1. tamanho da primeira maturação, tipo e época de desova. *Boletim do Instituto Oceanográfico* 40:01–13. <https://doi.org/10.1590/s0373-55241992000100001>
- Santos FB, Castro RMC (2003) Activity, habitat utilization, feeding behaviour, and diet of the sand moray *Gymnothorax ocellatus* (Anguilliformes, Muraenidae) in the south western Atlantic. *Biota Neotropica* 3:1–7. <https://doi.org/10.1590/s1676-06032003000100003>
- Santos MNS (2012) REPRODUÇÃO E ALIMENTAÇÃO DA GUARAJUBA *Carangoides bartholomaei* (CUVIER, 1833) (PERCIFORMES: CARANGIDAE) NA PLATAFORMA CONTINENTAL DE PERNAMBUCO, BRASIL. Universidade Federal de Pernambuco
- Sasekumar A, Chong VC, Leh MU, D´Cruz (1992) Mangroves as a habitat for fish and prawns. In: Jaccarini V, Marters E (eds) *The Ecology of Mangrove and Related Ecosystems*. Springer Science+Business Media, Mombasa, p 265
- Schärer MT, Nemeth MI, Appeldoorn RS (2010) Protecting a Multi-species Spawning Aggregation at Mona Island, Puerto Rico. *Gulf and Caribbean Fisheries Institute Proceedings* 252–259
- Schneider W (1990) FAO species identification sheets for fishery purposes. Field guide to the commercial marine resources of the Gulf of Guinea. Prepared and published with the support of the FAO Regional Office for Africa. FAO, Rome. FAO, Rome
- Silva JB da, Pereira ECG, Torres MFA (2007) Estuário de Itapessoca, Pernambuco: Relação entre pescadores e pescadeiras, e as espécies de maior importância comercial. *Bol Téc Cient CEPENE* 15:81–88

- Silva JDB da (2018) Ecologia trófica e distribuição espaço-temporal das espécies *Pomadasys ramosus* e *Haemulopsis corvinaeformis* (Steindachner, 1868) (Haemulidae) ao longo do gradiente de variação no estuário do Rio Goiana (PE/PB)
- Silva Júnior CAB da, Viana AP, Frédou FL, Frédou T (2015) Aspects of the reproductive biology and characterization of Sciaenidae captured as bycatch in the prawn trawling in the northeastern Brazil. *Acta Scientiarum Biological Sciences* 37:1–8. <https://doi.org/10.4025/actascibiolsci.v37i1.24962>
- Sley A, Jarboui O, Ghorbel M, Bouain A (2009) Food and feeding habits of *Caranx crysos* from the Gulf of Gabs (Tunisia). *Journal of the Marine Biological Association of the United Kingdom* 89:1375–1380. <https://doi.org/10.1017/S0025315409000265>
- Smith CL (1997) National Audubon Society field guide to tropical marine fishes of the Caribbean, the Gulf of Mexico, Florida, the Bahamas, and Bermuda. Alfred A. Knopf, Inc., New York
- Soares A, Lira AS, Gonzalez JG, et al (2020) Feeding habits and population aspects of the spotted goatfish, *pseudupeneus maculatus* (Perciformes: Mullidae), on the continental shelf of northeast Brazil. *Scientia Marina* 84:. <https://doi.org/10.3989/scimar.04958.24A>
- Sommer C, Schneider W, Poutiers JM (1996) FAO Species identification field guide for fishery purposes. The living resources of Somalia. FAO, Rome
- Tejerina-Garro FL, de Merona B (2010) Flow seasonality and fish assemblage in a tropical river, French Guiana, South America. *Neotropical Ichthyology* 8:145–154
- Temóteo TAA, Pina J, Lira AS, et al (2015) Alimentação de duas espécies do gênero *Caranx* capturadas no litoral de Pernambuco.
- Thorburn DC, Morgan DL, Rowland AJ, Gill HS (2007) Freshwater Sawfish *Pristis microdon* Latham, 1794 (Chondrichthyes : Pristidae) in the Kimberley region of Western Australia. *Zootaxa* 1471:27–41
- Tornabene L, Baldwin CC, Weigt LA, Pezold FL (2010) Exploring the diversity of western Atlantic Bathygobius (Teleostei: Gobiidae) with cytochrome c oxidase-I, with descriptions of two new species. *Aqua, Journal of Ichthyology and Aquatic Biology* 16:141–170
- Uyeno T, Matsuura K, Fujii E (1983) Fishes trawled off Suriname and French Guiana. Japan Marine Fishery Resource Research Center, Tokyo
- Vasconcellos MC, Freire KF, Castello JP (1998) Distribution patterns and feeding success of anchovy, *Engraulis anchoita*, larvae off southern Brazil. *Scientia Marina* 62:385–392. <https://doi.org/10.3989/scimar.1998.62n4385>
- Vasconcelos Filho A de L, Oliveira AME de (1999) Composição e ecologia da ictiofauna do Canal de Santa Cruz (Itamaracá-PE, Brasil). *Trab Oceanograf Univ Fed PE* 27:101–113
- Vasconcelos Filho AL, Neumann-Leitão S, Eskinazi-Leça E, Oliveira AME de (2010) Hábitos alimentares de peixes consumidores secundários do Canal de Santa Cruz, Pernambuco, Brasil. *Tropical Oceanography* 38:122–129. <https://doi.org/10.5914/tropocean.v38i2.5166>
- Vasconcelos-Filho AL (1979) Alimentação da sardinha-bandeira, *Ophistonema oglinun* (LeSueur, 1817) no Canal de Santa Cruz. *Trab Oceanogr* 14:105–116
- Vieira JP, Musick JA (1993) Latitudinal patterns in diversity of fishes in warm-temperate and tropical estuarine waters of the Western Atlantic. *Atlântica* 15:115–133
- Whitehead PJP (1985) Vol. 7. Clupeoid fishes of the world (Suborder Clupeoidei). An annotated and illustrated catalogue of the herrings, sardines, pilchards, sprats, shads, anchovies and wolf-herrings. FAO Fish. Synop., Rome
- Whitehead PJP, Vergara RR (1978) Megalopidae. In: Food Agriculture Organization species identification sheets for fishery purposes. Food and Agriculture Organization of the United Nations, Rome
- WoRMS EB (2020) World Register of Marine Species. Accessed 21 Oct 2020

Supplementary Material

Table II: Species and its original name throughout the references.

Species	Original Name
<i>Ginglymostoma cirratum</i> (Bonnaterre, 1788)	<i>Ginglymostoma cirratum</i>
<i>Carcharhinus leucas</i> (Muller & Henle, 1839)	
<i>Carcharhinus limbatus</i> (Muller & Henle, 1839)	
<i>Carcharhinus porosus</i> (Ranzani, 1839)	<i>Carcharhinus porosus</i>
<i>Isogomphodon oxyrhynchus</i> (Muller & Henle, 1839)	
<i>Rhizoprionodon lalandii</i> (Muller & Henle, 1839)	<i>Rhizoprionodon lalandei</i>
<i>Rhizoprionodon porosus</i> (Poey, 1861)	
<i>Sphyrna lewini</i> (Griffith & Smith, 1834)	
<i>Sphyrna tiburo</i> (Linnaeus, 1758)	<i>Sphyrna tiburo tiburo</i>
<i>Sphyrna tudes</i> (Valenciennes, 1822)	
<i>Narcine brasiliensis</i> (Olfers, 1831)	
<i>Pristis pristis</i> Muller & Henle, 1841	<i>Pristis perotteti</i>
<i>Pseudobatos percellens</i> (Walbaum, 1792)	<i>Rhinobatos percellens</i>
<i>Fontitrygon geijskesi</i> (Boeseman, 1948)	<i>Dasyatis geijskesi</i>
<i>Hypanus americanus</i> (Hildebrand & Schroeder, 1928)	
<i>Hypanus guttatus</i> (Bloch & Schneider, 1801)	<i>Dasyatis guttata</i>
<i>Hypanus marianae</i> (Gomes, Rosa & Gadig, 2000)	<i>Dasyatis marianae</i>
<i>Hypanus say</i> (Lesueur, 1817)	<i>Dasyatis say</i>
<i>Gymnura micrura</i> (Bloch & Scheneider, 1801)	
<i>Aetobatus narinari</i> (Euphrasen, 1790)	
<i>Mobula hypostoma</i> (Bancroft, 1831)	<i>Mobula hypostoma</i>
<i>Rhinoptera bonasus</i> (Mitchill, 1815)	
<i>Elops saurus</i> Linnaeus, 1766	
<i>Megalops atlanticus</i> Valenciennes, 1847	<i>Tarpon atlanticus</i>
<i>Albula vulpes</i> (Linnaeus, 1758)	
<i>Gymnothorax funebris</i> Ranzani, 1839	<i>Lycodontis funebris</i>
<i>Gymnothorax moringa</i> (Cuvier, 1829)	
<i>Gymnothorax ocellatus</i> Agassiz, 1831	
<i>Gymnothorax vicinus</i> (Castelnaud, 1855)	
<i>Ahlia egmontis</i> (Jordan, 1884)	
<i>Myrichthys breviceps</i> (Richardson, 1848)	
<i>Myrichthys ocellatus</i> (Lesueur, 1825)	
<i>Myrophis punctatus</i> Lütken, 1852	
<i>Ophichthus cylindroideus</i> (Ranzani, 1839)	<i>Ophichthus parilus</i>
<i>Ophichthus ophis</i> (Linnaeus, 1758)	
<i>Cynoponticus savanna</i> (Bancroft, 1831)	
<i>Ariosoma balearicum</i> (Delaroche, 1809)	
<i>Chirocentron bleekermanus</i> (Poey, 1867)	
<i>Odontognathus mucronatus</i> Lacepède, 1800	

<i>Pellona flavipinnis</i> (Valenciennes, 1837)	
<i>Pellona harroweri</i> (Fowler, 1917)	
<i>Anchoa filifera</i> (Fowler, 1915)	
<i>Anchoa hepsetus</i> (Linnaeus, 1758)	
<i>Anchoa januaria</i> (Steindachner, 1879)	
<i>Anchoa lyolepis</i> (Evermann & Marsh, 1900)	
<i>Anchoa parva</i> (Meek & Hildebrand, 1923)	
<i>Anchoa spinifer</i> (Valenciennes, 1848)	
<i>Anchoa tricolor</i> (Spix & Agassiz, 1829)	
<i>Anchovia clupeoides</i> (Swainson, 1839)	<i>Anchovia nigra</i>
<i>Anchoviella lepidentostole</i> (Fowler, 1911)	
<i>Cetengraulis edentulus</i> (Cuvier, 1829)	
<i>Engraulis anchoita</i> Hubbs & Marini 1935	
<i>Lycengraulis batesii</i> (Gunther, 1868)	<i>Lycengraulis barbouri</i>
<i>Lycengraulis grossidens</i> (Spix & Agassiz, 1829)	
<i>Pterengraulis atherinoides</i> (Linnaeus, 1766)	
<i>Harengula clupeola</i> (Cuvier, 1829)	
<i>Harengula jaguana</i> Poey, 1865	
<i>Lile piquitinga</i> (Schreiner & Miranda Ribeiro, 1903)	
<i>Opisthonema oglinum</i> (Lesueur, 1818)	
<i>Platanichthys platana</i> (Regan, 1917)	
<i>Rhinosardinia amazonica</i> (Steindachner, 1879)	
<i>Rhinosardinia bahiensis</i> (Steindachner, 1879)	
<i>Hoplias malabaricus</i> (Bloch, 1794)	
<i>Prochilodus argenteus</i> Spix & Agassiz, 1829	
<i>Pygocentrus nattereri</i> Kner, 1858	<i>Serrasalmus nattereri</i>
<i>Serrasalmus rhombeus</i> (Linnaeus, 1766)	
<i>Serrasalmus spilopleura</i> Kner, 1858	
<i>Schizodon fasciatus</i> Spix & Agassiz, 1829	
<i>Leporinus friderici</i> (Bloch, 1794)	
<i>Steindachnerina elegans</i> (Steindachner, 1875)	
<i>Astyanax bimaculatus</i> (Linnaeus, 1758)	
<i>Hemigrammus marginatus</i> Ellis, 1911	
<i>Hypostomus plecostomus</i> (Linnaeus, 1758)	
<i>Loricaria parnahybae</i> Steindachner, 1907	
<i>Aspistor luniscutis</i> (Valenciennes, 1840)	<i>Tachysurus luniscutis</i>
<i>Aspistor quadriscutis</i> (Valenciennes, 1840)	<i>Arius quadriscutis</i>
<i>Bagre bagre</i> (Linnaeus, 1766)	
<i>Bagre marinus</i> (Mitchill, 1815)	
<i>Cathorops agassizii</i> (Eigenmann & Eigenmann, 1888)	
<i>Cathorops spixii</i> (Agassiz, 1829)	
<i>Genidens genidens</i> (Cuvier, 1829)	
<i>Notarius grandicassis</i> (Valenciennes, 1840)	<i>Arius grandicassis</i>
<i>Sciades couma</i> (Valenciennes, 1840)	

<i>Sciades herzbergii</i> (Bloch, 1794)	<i>Arius herzbergii</i> ; <i>Hexanematichthys herzbergii</i> ; <i>Tachysurus herzbergii</i>
<i>Sciades parkeri</i> (Traill, 1832)	<i>Arius parkeri</i>
<i>Sciades proops</i> (Valenciennes, 1840)	<i>Hexanematichthys proops</i>
<i>Aspredo aspredo</i> (Linnaeus, 1758)	
<i>Aspredinichthys tibicen</i> (Valenciennes, 1840)	
<i>Ageneiosus inermis</i> (Linnaeus, 1766)	<i>Ageneiosus brevifilis</i>
<i>Pseudauchenipterus nodosus</i> (Bloch, 1794)	
<i>Trachelyopterus galeatus</i> (Linnaeus, 1766)	
<i>Pimelodella cristata</i> (Muller & Troschel, 1849)	
<i>Pimelodella lateristriga</i> (Lichtenstein, 1823)	
<i>Brachyplatystoma vaillantii</i> (Valenciennes, 1840)	
<i>Pseudoplatystoma fasciatum</i> (Linnaeus, 1766)	
<i>Sorubim lima</i> (Bloch & Schneider, 1801)	
<i>Synodus foetens</i> (Linnaeus, 1766)	
<i>Synodus poeyi</i> Jordan, 1887	
<i>Trachinocephalus myops</i> (Forster, 1801)	
<i>Holocentrus adscensionis</i> (Osbeck, 1765)	<i>Holocentrus adscensionis</i>
<i>Amphichthys cryptocentrus</i> (Valenciennes, 1837)	
<i>Batrachoides surinamensis</i> (Bloch & Schneider, 1801)	
<i>Thalassophryne nattereri</i> Steindachner, 1876	
<i>Thalassophryne punctata</i> Steindachner, 1876	
<i>Phaeoptyx pigmentaria</i> (Poey, 1860)	
<i>Dormitator maculatus</i> (Bloch, 1792)	<i>Dormitator maculatus</i>
<i>Eleotris pisonis</i> (Gmelin, 1789)	
<i>Erotelis smaragdus</i> (Valenciennes, 1837)	
<i>Guavina guavina</i> (Valenciennes, 1837)	
<i>Awaous tajasica</i> (Lichtenstein, 1822)	
<i>Bathygobius mystacium</i> Ginsburg, 1947	
<i>Bathygobius soporator</i> (Valenciennes, 1837)	
<i>Coryphopterus glaucofraenum</i> Gill, 1863	
<i>Ctenogobius boleosoma</i> (Jordan & Gilbert, 1882)	<i>Gobionellus boleosoma</i>
<i>Ctenogobius saepepallens</i> (Gilbert & Randall, 1968)	
<i>Ctenogobius shufeldti</i> (Jordan & Eigenmann, 1887)	
<i>Ctenogobius smaragdus</i> (Valenciennes, 1837)	<i>Gobionellus smaragdus</i>
<i>Ctenogobius stigmaticus</i> (Poey, 1860)	
<i>Evorthodus lyricus</i> (Girard, 1858)	
<i>Gobioides broussonnetii</i> Lacepède, 1800	<i>Gobioides broussonneti</i>
<i>Gobionellus oceanicus</i> (Pallas, 1770)	
<i>Gobionellus stomatus</i> Starks, 1913	
<i>Microdesmus bahianus</i> Dawson, 1973	
<i>Microdesmus longipinnis</i> (Weymouth, 1910)	
<i>Microgobius meeki</i> Evermann & Marsh, 1899	
<i>Abudefduf saxatilis</i> (Linnaeus, 1758)	

<i>Stegastes fuscus</i> (Cuvier, 1830)	
<i>Stegastes variabilis</i> (Castelnau, 1855)	
<i>Opistognathus cuvierii</i> Valenciennes, 1836	<i>Opistognathus cuvieri</i>
<i>Mugil brevisrostris</i> (Ribeiro, 1915)	
<i>Mugil curema</i> Valenciennes, 1836	
<i>Mugil curvidens</i> Valenciennes, 1836	<i>Mugil trichodon</i>
<i>Mugil incilis</i> Hancock, 1830	
<i>Mugil liza</i> Valenciennes, 1836	
<i>Mugil rubrioculus</i> Harrison, Nirchio, Oliveira, Ron & Gaviria, 2007	
<i>Cichla ocellaris</i> Bloch & Schneider, 1801	
<i>Oreochromis niloticus</i> (Linnaeus, 1758)	
<i>Dactyloscopus crossotus</i> Starks, 1913	
<i>Scartella cristata</i> (Linnaeus, 1758)	
<i>Labrisomus nuchipinnis</i> (Quoy & Gaimard, 1824)	
<i>Atherinella brasiliensis</i> (Quoy & Gaimard, 1825)	<i>Xenomelaniris brasiliensis</i>
<i>Atherinella cf. blackburni</i> (Schultz, 1949)	
<i>Odontesthes bonariensis</i> (Valenciennes, 1835)	
<i>Cheilopogon melanurus</i> (Valenciennes, 1847)	
<i>Hirundichthys affinis</i> (Gunther, 1866)	
<i>Hemiramphus balao</i> Lesueur, 1821	<i>Hemiramphus balao</i>
<i>Hemiramphus brasiliensis</i> (Linnaeus, 1758)	
<i>Hyporhamphus roberti roberti</i> (Valenciennes, 1847)	
<i>Hyporhamphus unifasciatus</i> (Ranzani, 1841)	
<i>Ablennes hians</i> (Valenciennes, 1846)	
<i>Strongylura marina</i> (Walbaum, 1792)	
<i>Strongylura timucu</i> (Walbaum, 1792)	
<i>Tylosurus acus acus</i> (Lacepède, 1803)	
<i>Tylosurus crocodilus crocodilus</i> (Péron & Lesueur, 1821)	
<i>Poecilia vivipara</i> Bloch & Schneider, 1801	
<i>Rachycentron canadum</i> (Linnaeus, 1766)	<i>Rachycentron canadus</i>
<i>Echeneis naucrates</i> Linnaeus, 1758	
<i>Phtheirichthys lineatus</i> (Menziés, 1791)	
<i>Remora remora</i> (Linnaeus, 1758)	
<i>Carangoides bartholomaei</i> (Cuvier, 1833)	
<i>Caranx crysos</i> (Mitchill, 1815)	
<i>Caranx hippos</i> (Linnaeus, 1766)	
<i>Caranx latus</i> Agassiz, 1831	
<i>Chloroscombrus chrysurus</i> (Linnaeus, 1766)	
<i>Hemicaranx amblyrhynchus</i> (Cuvier, 1833)	
<i>Oligoplites palometa</i> (Cuvier, 1832)	
<i>Oligoplites saliens</i> (Bloch, 1793)	
<i>Oligoplites saurus</i> (Bloch & Schneider, 1801)	
<i>Selene brownii</i> (Cuvier, 1816)	

<i>Selene setapinnis</i> (Mitchill, 1815)	
<i>Selene vomer</i> (Linnaeus, 1758)	
<i>Trachinotus carolinus</i> (Linnaeus, 1766)	
<i>Trachinotus cayennensis</i> Cuvier, 1832	
<i>Trachinotus falcatus</i> (Linnaeus, 1758)	
<i>Trachinotus goodei</i> Jordan & Evermann, 1896	
<i>Sphyraena barracuda</i> (Edwards, 1771)	
<i>Sphyraena guachancho</i> Cuvier, 1829	
<i>Citharichthys arenaceus</i> Evermann & Marsh, 1900	
<i>Citharichthys cornutus</i> (Günther, 1880)	
<i>Citharichthys macrops</i> Dresel, 1885	
<i>Citharichthys spilopterus</i> Günther, 1862	
<i>Etropus crossotus</i> Jordan & Gilbert, 1882	
<i>Etropus longimanus</i> Norman, 1933	
<i>Paralichthys brasiliensis</i> (Ranzani, 1842)	
<i>Paralichthys orbignyanus</i> (Valenciennes, 1839)	
<i>Paralichthys tropicus</i> Ginsburg, 1933	
<i>Syacium micrurum</i> Ranzani, 1842	
<i>Syacium papillosum</i> (Linnaeus, 1758)	
<i>Bothus lunatus</i> (Linnaeus, 1758)	
<i>Bothus ocellatus</i> (Agassiz, 1831)	
<i>Bothus robinsi</i> Topp & Hoff, 1972	
<i>Achirus achirus</i> (Linnaeus, 1758)	
<i>Achirus declivis</i> Chabanaud, 1940	
<i>Achirus lineatus</i> (Linnaeus, 1758)	
<i>Achirus mucuri</i> Ramos, Ramos & Lopes, 2009	
<i>Trinectes microphthalmus</i> (Chabanaud, 1928)	
<i>Trinectes paulistanus</i> (Miranda Ribeiro, 1915)	
<i>Symphurus diomedeanus</i> (Goode & Bean, 1885)	
<i>Symphurus plagusia</i> (Bloch & Schneider, 1801)	
<i>Symphurus tessellatus</i> (Quoy & Gaimard, 1824)	
<i>Bryx dunckeri</i> (Metzelaar, 1919)	<i>Sygnathus duncheri</i>
<i>Cosmocampus elucens</i> (Poey, 1868)	
<i>Hippocampus erectus</i> Perry, 1810	
<i>Hippocampus reidi</i> Ginsburg, 1933	
<i>Microphis brachyurus lineatus</i> (Kaup, 1856)	<i>Oostethus lineatus</i>
<i>Microphis brachyurus brachyurus</i> (Bleeker, 1854)	
<i>Fistularia petimba</i> Lacepède, 1803	
<i>Fistularia tabacaria</i> Linnaeus, 1758	
<i>Sygnathus pelagicus</i> Linnaeus, 1758	
<i>Dactylopterus volitans</i> (Linnaeus, 1758)	
<i>Trichiurus lepturus</i> Linnaeus, 1758	
<i>Euthynnus alletteratus</i> (Rafinesque, 1810)	<i>Euthynnus alleteratus</i>

<i>Scomberomorus brasiliensis</i> Collette, Russo & Zavala-Camin, 1978	
<i>Scomberomorus cavalla</i> (Cuvier, 1829)	
<i>Scomberomorus maculatus</i> (Mitchill, 1815)	
<i>Scomberomorus regalis</i> (Bloch, 1793)	
<i>Peprilus paru</i> (Linnaeus, 1758)	
<i>Astroscopus y-graecum</i> Cuvier, 1829	
<i>Halichoeres poeyi</i> (Steindachner, 1867)	
<i>Nicholsina usta</i> (Valenciennes, 1840)	
<i>Sparisoma amplum</i> (Ranzani, 1842)	
<i>Sparisoma axillare</i> (Steindachner, 1878)	
<i>Sparisoma radians</i> (Valenciennes, 1840)	
<i>Centropomus ensiferus</i> Poey, 1860	
<i>Centropomus mexicanus</i> Bocourt, 1868	
<i>Centropomus parallelus</i> Poey, 1860	
<i>Centropomus pectinatus</i> Poey, 1860	
<i>Centropomus undecimalis</i> (Bloch, 1792)	
<i>Diapterus auratus</i> Ranzani, 1842	<i>Diapterus olisthostomus</i>
<i>Diapterus rhombeus</i> (Cuvier, 1829)	
<i>Eucinostomus argenteus</i> Baird & Girard, 1855	
<i>Eucinostomus gula</i> (Quoy & Gaimard, 1824)	
<i>Eucinostomus havana</i> (Nichols, 1912)	
<i>Eucinostomus melanopterus</i> (Bleeker, 1863)	
<i>Eugerres brasilianus</i> (Cuvier, 1830)	
<i>Gerres cinereus</i> (Walbaum, 1792)	
<i>Ulaema lefroyi</i> (Goode, 1874)	
<i>Pseudupeneus maculatus</i> (Bloch, 1793)	
<i>Alphestes afer</i> (Bloch, 1793)	
<i>Diplectrum radiale</i> (Quoy & Gaimard, 1824)	<i>Diplectrum radeale</i>
<i>Epinephelus adscensionis</i> (Osbeck, 1765)	
<i>Epinephelus itajara</i> (Lichtenstein, 1822)	
<i>Epinephelus marginatus</i> (Lowe, 1834)	
<i>Hyporthodus nigrilus</i> (Holbrook, 1855)	
<i>Mycteroperca bonaci</i> (Poey, 1860)	<i>Mycteroperca bonasi</i>
<i>Mycteroperca tigris</i> (Valenciennes, 1833)	
<i>Mycteroperca venenosa</i> (Linnaeus, 1758)	
<i>Rypticus randalli</i> Courtenay, 1967	
<i>Rypticus saponaceus</i> (Bloch & Schneider, 1801)	
<i>Serranus flaviventris</i> (Cuvier, 1829)	
<i>Serranus phoebe</i> Poey, 1851	
<i>Pomacanthus arcuatus</i> (Linnaeus, 1758)	
<i>Pomacanthus paru</i> (Bloch, 1787)	
<i>Chaetodon ocellatus</i> (Bloch, 1787)	
<i>Chaetodon striatus</i> Linnaeus, 1758	

<i>Anisotremus surinamensis</i> (Bloch, 1791)	
<i>Anisotremus virginicus</i> (Linnaeus, 1758)	
<i>Conodon nobilis</i> (Linnaeus, 1758)	
<i>Genyatremus luteus</i> (Bloch, 1790)	
<i>Haemulon aurolineatum</i> Cuvier, 1830	
<i>Haemulon parra</i> (Desmarest, 1823)	
<i>Haemulon plumierii</i> (Lacepède, 1801)	
<i>Haemulon squamipinna</i> Rocha & Rosa, 1999	
<i>Haemulon steindachneri</i> (Jordan & Gilbert, 1882)	
<i>Orthopristis ruber</i> (Cuvier, 1830)	<i>Orthopristes ruber</i>
<i>Pomadasys corvinaeformis</i> (Steindachner, 1868)	<i>Pomadosys corvinaeformis</i>
<i>Pomadasys crocro</i> (Cuvier, 1830)	
<i>Pomadasys ramosus</i> Poey, 1860	
<i>Lutjanus alexandrei</i> Moura & Linderman, 2007	
<i>Lutjanus analis</i> (Cuvier, 1828)	
<i>Lutjanus apodus</i> (Walbaum, 1792)	
<i>Lutjanus cyanopterus</i> (Cuvier, 1828)	
<i>Lutjanus griseus</i> (Linnaeus, 1758)	
<i>Lutjanus jocu</i> (Bloch & Schneider, 1801)	
<i>Lutjanus synagris</i> (Linnaeus, 1758)	
<i>Ocyurus chrysurus</i> (Bloch, 1791)	
<i>Polydactylus oligodon</i> (Gunther, 1860)	
<i>Polydactylus virginicus</i> (Linnaeus, 1758)	
<i>Scorpaena isthmensis</i> Meek & Hilderbrand, 1928	
<i>Scorpaena plumieri</i> Bloch, 1789	
<i>Prionotus punctatus</i> (Bloch, 1793)	
<i>Chaetodipterus faber</i> (Broussonet, 1782)	
<i>Bairdiella ronchus</i> (Cuvier, 1830)	
<i>Cynoscion acoupa</i> (Lacepède, 1801)	
<i>Cynoscion jamaicensis</i> (Vaillant & Bocourt, 1833)	
<i>Cynoscion leiarchus</i> (Cuvier, 1830)	
<i>Cynoscion microlepidotus</i> (Cuvier, 1830)	
<i>Cynoscion steindachneri</i> (Jordan, 1889)	
<i>Cynoscion virescens</i> (Cuvier, 1830)	
<i>Isopisthus parvipinnis</i> (Cuvier, 1830)	
<i>Larimus breviceps</i> Cuvier, 1830	
<i>Macrodon ancylodon</i> (Bloch & Schneider, 1801)	
<i>Menticirrhus americanus</i> (Linnaeus, 1758)	<i>Menticirrhus martinicensis</i>
<i>Menticirrhus littoralis</i> (Holbrook, 1847)	
<i>Micropogonias furnieri</i> (Desmarest, 1823)	<i>Micropogon furnieri</i>
<i>Nebris microps</i> Cuvier, 1830	
<i>Odontoscion dentex</i> (Cuvier, 1830)	
<i>Ophioscion punctatissimus</i> Meek & Hildebrand, 1925	
<i>Paralanchurus brasiliensis</i> (Steindachner, 1875)	

<i>Pareques acuminatus</i> (Bloch & Schneider, 1801)	
<i>Pogonias cromis</i> (Linnaeus, 1766)	
<i>Stellifer brasiliensis</i> (Schultz, 1945)	
<i>Stellifer microps</i> (Steindachner, 1864)	
<i>Stellifer naso</i> (Jordan, 1889)	
<i>Stellifer rastrifer</i> (Jordan, 1889)	
<i>Stellifer stellifer</i> (Bloch, 1790)	
<i>Umbrina coroides</i> Cuvier, 1830	
<i>Acanthurus bahianus</i> Castelnau, 1855	
<i>Acanthurus chirurgus</i> (Bloch, 1787)	
<i>Acanthurus coeruleus</i> Bloch & Schneider, 1801	
<i>Lobotes surinamensis</i> (Bloch, 1790)	
<i>Archosargus probatocephalus</i> (Walbaum, 1792)	
<i>Archosargus rhomboidalis</i> (Linnaeus, 1758)	
<i>Antennarius multiocellatus</i> (Valenciennes, 1837)	
<i>Antennarius striatus</i> (Shaw, 1794)	
<i>Ogcocephalus vespertilio</i> (Linnaeus, 1758)	
<i>Acanthostracion quadricornis</i> (Linnaeus, 1758)	<i>Achantostracion quadricornis</i>
<i>Lactophrys trigonus</i> (Linnaeus, 1758)	
<i>Lactophrys triqueter</i> (Linnaeus, 1758)	<i>Rhonesomus triquerte</i>
<i>Balistes vetula</i> Linnaeus, 1758	
<i>Melichthys niger</i> (Bloch, 1786)	<i>Melichthys piceus</i>
<i>Aluterus heudelotii</i> Hollard, 1855	
<i>Aluterus monoceros</i> (Linnaeus, 1758)	
<i>Aluterus schoepfii</i> (Walbaum, 1792)	
<i>Cantherhines pullus</i> (Ranzani, 1842)	
<i>Monacanthus ciliatus</i> (Mitchill, 1818)	
<i>Stephanolepis hispidus</i> (Linnaeus, 1766)	
<i>Stephanolepis setifer</i> (Bennett, 1831)	
<i>Canthigaster figueiredoi</i> Moura & Castro, 2002	
<i>Colomesus psittacus</i> (Bloch & Schneider, 1801)	
<i>Lagocephalus laevigatus</i> (Linnaeus, 1766)	
<i>Sphoeroides greeleyi</i> Gilbert, 1900	
<i>Sphoeroides nephelus</i> (Goode & Bean, 1882)	
<i>Sphoeroides spengleri</i> (Bloch, 1785)	
<i>Sphoeroides testudineus</i> (Linnaeus, 1758)	
<i>Sphoeroides tyleri</i> Shipp, 1972	
<i>Chilomycterus antillarum</i> Jordan & Rutter, 1897	<i>Cyclichthys antillarum</i>
<i>Chilomycterus spinosus spinosus</i> (Linnaeus, 1758)	<i>Chilomycterus spinosus</i> / <i>Ciclichthys spinosus spinosus</i>
<i>Diodon holocanthus</i> Linnaeus, 1758	
<i>Diodon hystrix</i> Linnaeus, 1758	