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CRESCIMENTO E SOBREVIVÊNCIA LARVAL DO MARISCO
Anomalocardia brasiliiana (GMELIN, 1791) ALIMENTADO COM DIFERENTES
DIETAS ALGAIS

RECIFE,
2014



UNIVERSIDADE FEDERAL RURAL DE PERNAMBUCO
PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM RECURSOS PESQUEIROS E AQUICULTURA

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***Anomalocardia brasiliiana* (GMELIN, 1791) ALIMENTADO COM DIFERENTES**
DIETAS ALGAIS

Isabela Bacalhau de Oliveira

Tese apresentada ao Programa de Pós-Graduação em Recursos Pesqueiros e Aquicultura da Universidade Federal Rural de Pernambuco como exigência para obtenção do título de Doutor.

Prof.(a) Dr.(a) Alfredo Olivera Gálvez
Orientador

Recife,
Agosto/2014

Ficha catalográfica

Setor de Processos Técnicos da Biblioteca Central - UFRPE

- O48c Oliveira, Isabela Bacalhau de
Crescimento e sobrevivência larval do marisco
Anomalocardia brasiliiana (Gmelin, 1791) alimentado com
diferentes dietas algais / Isabela Bacalhau de Oliveira. –
Recife, 2014.
95 f.
- Orientador: Alfredo Olivera Gálvez.
Tese (Doutorado em Recursos Pesqueiros e
Aquicultura) – Universidade Federal Rural de Pernambuco,
Departamento de Pesca e Aquicultura, Recife, 2014.
Inclui referências e anexo(s).
1. Microalgas 2. Densidade de estocagem 3. Larvas
4. Pós-larvas 5. Manejo I. Olivera Gálvez, Alfredo, orientador
II. Título

CDD 639

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**CRESCIMENTO E SOBREVIVÊNCIA LARVAL DO MARISCO *Anomalocardia
brasiliiana* (GMELIN, 1791) ALIMENTADO COM DIFERENTES DIETAS ALGAIS**

Isabela Bacalhau de Oliveira

Tese julgada adequada para obtenção do título de doutor em Recursos Pesqueiros e Aquicultura. Defendida e aprovada em 25/08/2014 pela seguinte Banca Examinadora.

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Dedicatória

Dedico este trabalho as pessoas mais importantes de minha vida e que sempre estarão presentes em meu coração.

Minha mãe Maria das Graças Bacalhau;

Meu pai Moisés Cavalcanti;

Meu irmão Márcio Bacalhau;

Minha irmã Fabiane Bacalhau;

Meu sobrinho Otávio Bacalhau, e

Minhas sobrinhas Alice e

Letícia Bacalhau.

Agradecimentos

À Universidade Federal Rural de Pernambuco e ao Programa de Pós-Graduação em Recursos Pesqueiros e Aquicultura, em especial a Selma Santiago (secretária) pela enorme atenção e carinho;

Ao Instituto Federal de Sergipe, Campus Estância, principalmente a Coordenadoria de Recursos Pesqueiros pelo apoio durante o doutorado;

Ao orientador Alfredo Olivera Gálvez, pela amizade e confiança, pelas inúmeras correções de trabalhos, mas, principalmente, pelos valiosos ensinamentos que serão eternos;

Aos membros que fizeram parte da Banca Examinadora, os professores Dr. Alfredo Olivera Gálvez, Dr. Paulo Oliveira, Dr. José Carlos, Dra. Lilia Santos, Dra. Danielli Dantas, e aos suplentes Dr. Paulo Roberto Oliveira Filho e Dra. Raquel Coimbra pelas sugestões e colaborações para este trabalho;

Aos professores da Pós-graduação em Recursos Pesqueiros e Aquicultura pela excelência no ensino, Dra Rosângela Lessa, Dr. Silvio Peixoto, Dr. Paulo Travassos, Dr. Paulo de Paula Mendes, Dr. Eudes Correia, Dra. Roberta Soares, Dr. Fábio Hazin, Dr. Ronaldo Cavalli e os demais que já foram citados anteriormente;

Aos que fazem o Laboratório de Maricultura Sustentável, pela cumplicidade e companheirismo, que certamente tornaram este trabalho melhor, em especial à equipe marisco, Henrique, Sérgio, Leônidas, Priscila e Luciana;

Ao querido Emanuell Felipe, por ser o irmão que a vida me deu o prazer de escolher e pelas histórias que compartilhamos e que sempre serão motivos de muitas risadas em todos os momentos de encontros;

Aos meus amigos e companheiros de profissão do grupo Ruralindas: Adriana, Leilane, Suzianny, Hozana, Emanuell, Joana, Danielli, Wanessa, João Paulo, Juliana, Ana Melo, Bruna, Penélope e Fabiana Penalva;

Aos meus amigos da escola: Adilson, Amanda, Helzevon, Juliana, Milena, Paula, Renato, Talita e Victor;

E a todos aqueles que injustamente não foram citados.

Resumo

O presente estudo descreve a larvicultura do molusco bivalve *Anomalocardia brasiliiana*, com avaliação de diferentes dietas e densidade de estocagem. As dietas avaliadas foram: microalgas *Isochrysis galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo), *Chaetoceros calcitrans* (Cca), *Pavlova lutheri* (Pl) e as combinações (Ig + Cca), (Ig + Phaeo), (Cca + Phaeo) e (Cca + Pl), totalizando oito dietas fornecidas por um período de 15 dias de cultivo. A microalga *I. galbana* fornecida isoladamente, apresentou menor sobrevivência e crescimento em relação as demais dietas testadas. O uso de *C. calcitrans* e *P. tricornutum* isoladamente ou combinada as outras microalgas apresentou melhores valores de sobrevivência e crescimento. A dieta combinada Cca + Pl, obteve maior crescimento e sobrevivência nas larvas de *A. brasiliiana* ($261,67 \pm 9,64 \mu\text{m}$ e $31,50 \pm 0,87 \%$). Todas as dietas avaliadas obtiveram resultados satisfatórios quanto ao crescimento e sobrevivência de larvas de *A. brasiliiana*, exceto a dieta *I. galbana* quando fornecida isoladamente. As densidades de estocagem avaliadas nas pós-larvas de *Anomalocardia brasiliiana* foi de 40, 80 e 160 ind.cm⁻² em um período experimental de 28 dias. A densidade de 40 ind.cm⁻² apresentou a maior taxa de crescimento específico diária. As maiores sobrevivências das pós-larvas foram observadas nas menores densidades $53,24 \pm 4,60 \%$ (40 ind.cm⁻²) e $52,95 \pm 3,32 \%$ (80 ind.cm⁻²), diferindo significativamente da maior densidade de estocagem com $31,54 \pm 0,70 \%$. Assim pôde-se concluir que podemos produzir larvas de *A. brasiliiana* utilizando qualquer uma das dietas avaliadas, exceto *I. galbana* se fornecida isoladamente. No cultivo de pós-larvas desta mesma espécie devemos realizar o manejo na densidade de estocagem no decorrer do crescimento, a densidade de 160 ind.cm⁻² pode ser utilizada até que as larvas alcancem 600 μm de comprimento, larvas maiores que 600 μm devem ser cultivadas na densidade de 40 ind.cm⁻² para manter a taxa de crescimento máximo diária.

Palavras-chave: microalgas, densidade de estocagem, larvas, pós-larvas, manejo.

Abstract

The present study describes the larval rearing of bivalve mollusc *Anomalocardia brasiliiana*, with evaluation of different diets and stocking density. The diets were evaluated: microalgae *Isochrysis galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo), *Chaetoceros calcitrans* (Cca), *Pavlova lutheri* (Pl) and combinations (Ig + Cca), (Ig + Phaeo) (Cca + Phaeo) and (Cca + Pl), totaling eight diets provided for a period of 15 days of cultivation. The microalgae *I. galbana* provided alone had lower survival and growth compared with other diets tested. The use of *C. calcitrans* and *P. tricornutum* alone or combined other microalgae showed highest values for survival and growth. The combined diet Cca + Pl, demonstrated the better growth and survival of larval *A. brasiliiana* ($261.67 \pm 9.64 \mu\text{m}$ and $31.50 \pm 0.87\%$). All diets evaluated with satisfactory results regarding the growth and survival of larvae of *A. brasiliiana* except *I. galbana* diet when given alone. The stocking densities evaluated in post-larvae *Anomalocardia brasiliiana* was 40, 80 and 160 ind.cm⁻² in a trial period of 28 days. The density of 40 ind.cm⁻² had the highest rate of daily specific growth. The highest survival of post-larvae were observed at lower densities $53.24 \pm 4.60\%$ (40 ind.cm⁻²) and $52.95 \pm 3.32\%$ (80 ind.cm⁻²), differing from greater stocking density with $31.54 \pm 0.70\%$. So we concluded that we can produce larvae of *A. brasiliiana* evaluated using any of the diets, except *I. galbana* was provided separately. In cultivation post-larvae of the same species must perform storage management in density during growth, density 160 ind.cm⁻² can be used to achieve the larvae 600 μm in length, greater than 600 μm larvae must be grown at a density of 40 ind.cm⁻² to keep the rate of maximum growth rate.

Key words: microalgae, stocking density, larvae, post-larvae, management.

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1 - Introdução

A produção aquícola mundial baseia-se no cultivo de seis grupos de organismos aquáticos: peixes de água doce, peixes diádromos, peixes marinhos, algas, crustáceos e moluscos. Em 2012 a produção mundial de moluscos foi de 15,2 milhões de toneladas, sendo mais que o dobro da produção de crustáceos com 6,4 milhões de toneladas (FAO, 2014).

No Brasil o cultivo de moluscos é representado por três espécies, a ostra japonesa (*Crassostrea gigas*) espécie exótica, a ostra nativa (*Crassostrea brasiliiana*) e a vieira (*Nodipecten nodosus*). Entretanto, existem várias outras espécies nativas que apresentam potencial para o cultivo dentre elas pode-se citar o marisco *Anomalocardia brasiliiana*, sendo cultivadas espécies da mesma família (Veneridae) em outras partes do mundo e que apresentam um mercado consolidado. O maior problema, para produção em laboratório, é conseguir uma produção regular de sementes, pois pouco se conhece sobre as exigências desta espécie para determinação de uma metodologia de produção.

As Microalgas marinhas são cultivadas para serem utilizadas como alimento para as várias fases de vida dos moluscos bivalves e, até o momento, têm constituído o principal alimento de larvas e sementes de bivalves cultivados (HELM et al., 2004). Outra função do cultivo de microalgas na aquicultura é proporcionar a melhoria da qualidade da água, através da absorção de produtos nitrogenados tóxicos (amônia e nitrito) e combate as bactérias patogênicas pela sua produção de substâncias antibióticas (LAVENS e SORGELLOS, 1996; REITAN et al. 1994). As microalgas mais utilizadas nas larviculturas de moluscos bivalves são as espécies dos seguintes gêneros: *Isochrysis*, *Pavlova*, *Chaetoceros*, *Phaeodactylum*, *Skeletonema*, *Thalassiosira*, *Dunaliella*, *Nannochloris*, *Tetraselmis* e *Rhodomonas* (COUTTEAU e SORGELOOS,

1992; MOUEZA et al., 1999; BROWN, 2002; HELM e BOURNE, 2004; CRAGG, 2006; LIU et al., 2009).

A larvicultura é uma das etapas mais importantes no cultivo de organismos aquáticos, e uma alimentação nutricionalmente balanceada, trará mais resistência aos animais quando forem cultivados fora do laboratório. No entanto, uma das principais causas de mortalidade na larvicultura é o desenvolvimento da comunidade bacteriana. Por exemplo larvas de vieira, *Pecten maximus*, são suscetíveis à contaminação bacteriana causando alta mortalidade nos cultivos intensos (COMELY, 1972; NICHOLAS et al., 1996). Além disso, fatores como alta temperatura, alta densidade de estocagem e excesso de nutrientes podem favorecer a proliferação de patógenos oportunistas (ANDERSEN et al., 2000).

O efeito da densidade de estocagem no crescimento e sobrevivência de larvas, juvenis e adultos de moluscos bivalves tem sido amplamente estudado (FRÉCHETTE, 2005), principalmente para aquelas espécies com importância comercial para aquicultura, como: *Crassostrea gigas*, *Ruditapes philippinarum*, *Perna perna*. A maioria dos estudos avaliaram o período de crescimento nas fases de D véliger até a metamorfose para pé de véliger (LIU et al., 2006; YAN et al., 2006; LIU et al., 2010), pois nesta fase as larvas são frágeis e altamente susceptíveis a doenças.

O conhecimento da melhor microalga, e/ou suas combinações na alimentação de *A. brasiliana* assim como o manejo adequado é essencial para aumentar o desenvolvimento e sobrevivência de larvas e pós-larvas de marisco.

2 - Objetivos

2.1 - Objetivo geral

A presente proposta visa contribuir com o aprimoramento de tecnologias para a larvicultura do marisco *A. brasiliiana* na região Nordeste do Brasil.

2.2 - Objetivo(s) específico(s)

1. Avaliar o efeito de diferentes dietas microalgais no crescimento e sobrevivência de larvas do marisco *A. brasiliiana*;
2. Analisar o efeito de dietas combinadas de microalgas no crescimento e sobrevivência de pós-larvas do marisco *A. brasiliiana*
3. Avaliar o efeito da densidade de estocagem no crescimento e sobrevivência de pós-larvas do marisco *A. brasiliiana*;
4. Analisar o efeito da densidade de estocagem no crescimento e sobrevivência de pós-larvas do marisco *A. brasiliiana*, ao longo do tempo.

3 - Revisão de literatura

Anomalocardia brasiliiana pertence ao Reino Animal, Filo Mollusca, Classe Bivalvia, Subclasse Heterodonta, Infraclasse Euheterodonta, Ordem Veneroidea, Superfamília Veneroidea, Família Veneridae, Gênero *Anomalocardia* e espécie *brasiliiana* (Gmelin, 1791). Reúnem aproximadamente 500 espécies viventes, pertencentes à aproximadamente cinquenta gêneros e doze subfamílias (CANAPA et al., 1996). Essa diversidade está relacionada à grande variedade de habitats que estão adaptados, tais como: praias arenosas, areno-lodosas, manguezais e fundos arenosos de ambientes coralíneos (CANTERA, 1991). No Brasil foram registradas 35 espécies de venerídeos, pertencentes a quatorze gêneros e sete subfamílias (RIOS, 1994).

A. brasiliiana vive em profundidades de 0,3 – 5 m, superficialmente escavado na areia próximo ao manguezal, sensíveis a variações ecológicas, com alta mortalidade devido às chuvas, causando grandes flutuações no tamanho e na distribuição das populações (MOUEZA et al., 1999; MONTI et al., 1991). Distribui-se desde a costa das Antilhas até o Uruguai (RIOS, 1994), e o tamanho máximo já encontrado foi de 38 mm segundo dados do Malacolog version 4.1.1. (A Database of Western Atlantic Marine Mollusca). Encontra-se em maior abundância na região entre marés nos bancos naturais formados da interação dos mesmos no sedimento com uma grande variedade de bancos naturais, *micro-habitats*. É conhecido popularmente na região sudeste e sul do Brasil como berbigão, papafumo, sarnambi ou vôngoli, e na região nordeste é chamado de marisco-pedra e maçunim (OLIVEIRA et al., 2014).

A *A. brasiliiana* é uma espécie dióica, ou seja, apresenta sexos separados, não apresenta características morfológicas externas (diferença na coloração ou tamanho das conchas) ou internas (diferença na coloração das gônadas) que permitam a diferenciação macroscópica dos sexos, sendo necessária análise microscópica dos gametas ou estudos

histológicos (GROTTA & LUNETTA, 1980). A Reprodução é sexuada com lançamento de gametas na água e fecundação externa.

O desenvolvimento larval da *A. brasiliiana* passa pelas fases trocófora, véliger (D-véliger, umbonada e pé-de-véliger) e pós-larva ou juvenil. MOÛEZA et al (1999) relataram o surgimento de larvas trocóforas após 9 horas do início da fertilização, e após 24 h de vida já apresentavam aproximadamente 95 µm de comprimento e estavam no estágio D-véliger. A partir do quinto dia passaram para véliger com comprimento médio de 150 µm. No sétimo dia inicia a metamorfose para fase pé de véliger, sendo esta fase considerada crítica, pois as larvas são frágeis e susceptíveis a doenças durante o manejo (OLIVEIRA et al., 2007). Aos 11 dias de vida, as larvas migram para o fundo, fase pé de véliger, 180 µm de comprimento, e com 15 dias apresentando todas as características de juvenil, sifão e o pé totalmente desenvolvidos, com aproximadamente 300 µm de comprimento (MOÛEZA et al., 1999).

No habitat natural a espécie pode reproduzir o ano todo, porém apresenta picos de reprodução em algumas épocas do ano. O período reprodutivo para uma mesma espécie pode variar substancialmente de região para região, principalmente em função de diferentes condições climáticas e ambientais (LAVANDER et al., 2011). Sendo o Brasil um país com grande extensão costeira, possivelmente o padrão maturacional para *A. brasiliiana* é diferenciado entre as regiões do país.

Estudos realizados sobre a biologia reprodutiva da *A. brasiliiana* no Brasil identificaram que em São Paulo, Santa Catarina e no Paraná a espécie apresenta ciclo reprodutivo contínuo, com dois períodos de liberação de gametas na primavera e outono, assim como um período de repouso parcial no inverno (NARCHI, 1976; BOEHS, 2000; ARAÚJO, 2001).

No estado da Paraíba, a produção de gametas ocorre durante todo o ano, devido às condições ambientais favoráveis, temperatura constante e baixo índice de precipitação (GROTTA & LUNETTA, 1980). No Ceará foi observado dois picos reprodutivos: julho a outubro (primavera e inverno) e fevereiro a abril (verão e outono) (BARREIRA & ARAÚJO, 2005). Em Pernambuco, da mesma forma, apresenta ciclo reprodutivo contínuo, com eliminação de gametas de outubro a junho (LAVANDER et al., 2011).

Concomitantemente aos estudos de biologia reprodutiva foram realizados estudos sobre a dinâmica populacional de *A. brasiliana* na costa brasileira (PEZZUTO e ECHTERNACHT, 1999; BOEHS, 2008; RODRIGUES, 2009; EL-DEIR, 2009; OLIVEIRA et al., 2011; RODRIGUES et al., 2013). Em Pernambuco, El-deir (2009) observou grandes variações na distribuição da espécie, apresentando uma densidade média de 1348 ind.m². Para o mesmo Estado, Oliveira et al. (2011) encontraram valores médios de densidade no período de verão e inverno de 298 ind.m² e 173 ind.m² respectivamente. Esses resultados são semelhantes aos encontrados por Rodrigues et al. (2013), que também estudaram a espécie no litoral de Pernambuco e verificou que em ambiente natural a espécie pode ser encontrada em até 268 ind.m².

Os dados de dinâmica populacional e biologia reprodutiva são necessários para o estabelecimento de medidas regulatórias da atividade pesqueira. Contudo, até o momento, as medidas legais tomadas não têm apresentado um resultado satisfatório na preservação dos estoques naturais da *A. brasiliana*.

A crescente demanda por frutos do mar e a facilidade com que os estoques costeiros podem ser capturados (DAME et al., 2002) resultou em uma super exploração e colapso de algumas espécies de pesca dos ecossistemas costeiros (MORA et al., 2009; FAO, 2010). Na costa brasileira, a pesca de *A. brasiliana* é amplamente difundida e intensa, com grande importância econômica para grupos de famílias de pescadores

artesanais (OLIVEIRA et al., 2014). A coleta de *A. brasiliiana* está registrada desde 1970 em Pernambuco, quando a atividade era praticada apenas por mulheres, porém ao longo dos anos, o número de pescadores do sexo masculino tem aumentado significativamente (SILVA-CAVALCANTI, 2009).

Dentre as principais espécies de moluscos, no estado de Pernambuco, o marisco *A. brasiliiana* é o que mais se destaca. A produção no ano de 2007 foi de 2.479,2 t responsável por 20% da produção total de moluscos (CEPENE, 2008). É o principal recurso pesqueiro entre os bivalves, e a principal fonte de renda de marisqueiras, e teve sua exploração quase que dobrada entre os anos de 2003 a 2005 (PEZZUTO & ECHTERNACHT, 1999). Infelizmente não há dados recentes sobre a produção pesqueira de *A. brasiliiana* no estado de Pernambuco, porém as comunidades tradicionais perceberam a diminuição deste recurso tanto em relação ao tamanho quanto ao volume (SILVA-CAVALCANTI, 2009). ARRUDA-SOARES et al. (1982) recomendam a captura de espécimes de *A. brasiliiana* com comprimento acima de 20 mm, neste tamanho os indivíduos já teriam alcançado um grau de desenvolvimento reprodutivo que possibilitasse a reposição e manutenção dos estoques naturais.

O declínio dos bancos naturais de moluscos bivalves por todo mundo, tornou-se um incentivo ao crescimento das larviculturas para o fornecimento de sementes (PRADO et al., 2010). As atividades básicas de qualquer larvicultura de moluscos bivalves são condicionamento e desova do estoque reprodutor, cultivo e assentamento das larvas e sementes até um tamanho aceitável, assim como a produção de grandes quantidades de alimento (microalgas) para todas as fases do ciclo de produção (HELM et al., 2004).

A microalga *Isochrysis galbana* é considerada um ótimo alimento para as larvas dos moluscos bivalves *Mercenaria mercenaria* e *Tapes semidecussata*, porém para as

ostras *Crassostrea gigas* e *Crassostrea rhizophorae* e o marisco *Donax trunculus* não foi obtido um bom desenvolvimento larval (HELM e LAING, 1987; RUIZ-AZCONA et al., 1996). Liu et al. (2009) avaliaram o efeito de dietas de microalgas no crescimento e sobrevivência de larvas e pós-larvas de *Clinocardium nuttallii*. Os autores constataram que o uso de dietas espécie-específica de *I. galbana* (TISO) e *Chaetoceros muelleri* (CM) são suficientes para proporcionar um máximo crescimento de larvas e pós-larvas de *C. nuttallii*. Entretanto o uso destas microalgas utilizadas de forma combinada apresentou um melhor crescimento na fase larval ou no início da fase pós-larval, sendo recomendado o uso desta dieta bialgal para atender as necessidades nutricionais das sementes de *C. nuttallii*.

Martínéz-Fernandez e Southgate (2007) obtiveram uma maior taxa de crescimento na fase D-larva de *Pinctada margaritifera* para aquelas cultivadas com a combinação ternária de *Pavlova* sp. / *Pavlova salina* / TISO. No entanto, esta taxa de crescimento não foi significativamente maior do que a das larvas alimentadas com a combinação binária do *Pav. salina* / TISO que tem sido usado como uma “dieta padrão” para a larvicultura de *P. margaritifera* (SOUTHGATE e BEER, 1997). Martínéz-Fernandez e Southgate (2007) sugerem que a alimentação com uma única espécie de microalga para larvas de *P. margaritifera* pode ter uma finalidade prática, durante os primeiros 10 dias de larvicultura. No entanto, a adição de uma microalga diatomácea flagelada na dieta teve um incremento na taxa de crescimento e sobrevivência das larvas umbonadas de *P. margaritifera* quando comparada às combinações sem diatomáceas.

As haptophyta da classe Pavlovophyceae são microalgas flageladas com tamanho de 3-5 µm, ou seja apropriado para ingestão de larvas de moluscos (PONIS et al., 2006), são ricas em ácidos graxos polinsaturados (PUFA) (KANAZAWA et al., 1979; VOLKMAN et al., 1991) . *Pavlova lutheri* é conhecida como um excelente alimento de

larvas de *Pecten maximus* e *Pecten fumatus*, contudo para larvas de *Crassostrea gigas* apresentou baixo valor alimentar, com menor sobrevivência das larvas (HEASMAN et al., 2000; PONIS et al., 2003a,b; LAING, 2004).

A utilização de *Pavlova pinguis* e *Radomonas salina* como alimento complementar a alimentação natural de juvenis da ostra do pacífico, *Crassostrea gigas*, em uma larvicultura comercial, produz aumento de custo-benefício na produção animal (BROWN et al., 1998). Estes mesmos autores observaram que a utilização de dietas mono-específica de *Tetraselmis* sp. CS-362 e *Nannochloropsis* sp. CS-246 foram mal ingeridas, e é provável que este fator tenha causado crescimento inferior das ostras.

Rivero-Rodríguez et al. (2007) constataram que a dieta mono-específica de *Chaetoceros calcitrans* para juvenis de *Crassostrea corteziensis* foi superior a outras dietas testadas (*Tetraselmis suecica*, *Isochrysis galbana* clone T-iso, *Phaeodactylum tricornutum*, *Chaetoceros muelleri*, *Chaetoceros calcitrans* e suas combinações bi-algas). No geral, os juvenis de *C. corteziensis* alimentados com as dietas com qualquer combinação de *C. calcitrans* obtiveram melhor crescimento. O contrário foi observado para as microalgas *I. galbana*, *T. suecica*, *P. tricornutum* e suas combinações bi-algais.

Estudos afirmam que o uso da microalga *P. tricornutum*, em outros bivalves, afeta o crescimento, deixando-o mais lento (EPIFANIO et al., 1981; ALBENTOSA et al., 1996; RIVERO-RODRIGUEZ et al., 2007), devido à microalga ser de difícil digestão (RIVERO-RODRIGUEZ et al., 2007), possivelmente devido a falta de triptofano na dieta (EPIFANIO et al., 1981). Tang et al. (2006) obtiveram baixo crescimento de larvas de *Meretrix meretrix* quando alimentadas com *P. tricornutum* e *Pavlova viridis*.

O sistema Europeu de cultivo berçário de larvas de vieiras, inclui a troca completa da água várias vezes durante a semana, e alimentação uma ou duas vezes ao dia

(BUESTEL et al., 1982; COCHARD e DEVAUCHELLE, 1993; TAYLOR et al., 1994), que resulta em altos valores de matéria orgânica, e um ambiente microbiano instável, o qual é comumente controlado com uso profilático de agentes antibacteriano (ROBERT et al., 1996; TORKILDSEN et al., 2000). As larviculturas de moluscos bivalves são frequentemente afetadas por surtos de doenças, como infecções bacterianas do gênero *Vibrio* (PRADO et al, 2010). Andersen et al. (2000) observaram que o uso de antibióticos pode, hipoteticamente, alterar a atividade bacteriana ou a composição das espécies no sistema, ocorrendo menor degradação ou aumento na produção de amônia.

Prado et al. (2010) observaram em sua revisão que todos os tratamentos considerados (Filtração, Radiação Ultravioleta e Quimioterápicos) tem como objetivo a eliminação completa da microbiota presente na água, entretanto populações de bactérias ou parte delas tem efeito benéfico no desenvolvimento larval de bivalves. As bactérias parecem satisfazer as exigências metabólicas, fornecendo vitaminas ou outros fatores de crescimento (PRIEUR et al., 1990).

Foram realizados diversos estudos para estabelecer um protocolo de produção de larvas de bivalves, incluindo bivalves de areia (LIU et al, 2002; LIU et al, 2006; YAN et al, 2006; LIU et al, 2008; LIU et al, 2010), mexilhões (GALLEY et al, 2010; PETTERSEN et al, 2010), ostras (RICO-VILLA et al, 2006) e vieiras (RUPP e PARSONS, 2004; GOUDA et al, 2006), pois a identificação dos fatores que influenciam no crescimento e na sobrevivência das larvas de moluscos é essencial para o sucesso da larvicultura.

Fatores como temperatura, salinidade e alimentação podem influenciar na sobrevivência, crescimento e período de metamorfose larval para assentamento na espécie *A. brasiliiana*. A sobrevivência e crescimento de moluscos são afetados por parâmetros físicos como temperatura e salinidade (KINNE, 1964), assim como manejo

e alimentação (DEVAKIE & ALI, 2000; OLIVEIRA et al, 2007). Para o molusco de areia mais cultivado no mundo, *Ruditapes philippinarum*, Delgado & Camacho (2007) descreveram que o aumento da temperatura influencia grande parte dos processos fisiológicos, afetando seu crescimento. Em *A. brasiliiana* a temperatura também é fator primordial para seu crescimento (BARREIRA & ARAUJO, 2005).

Outra importante variável a ser considerada, é a densidade de estocagem em larvas de moluscos bivalves. O efeito da densidade no crescimento e sobrevivência de larvas, juvenis e adultos tem sido amplamente estudado (FRÉCHETTE, 2005), principalmente nas espécies com importância comercial para aquicultura. A maioria dos estudos avaliaram o período de crescimento nas fases de D véliger até a metamorfose para pé de véliger (LIU et al., 2006; YAN et al., 2006; LIU et al., 2010), pois é considerada crítica, devido as larvas serem frágeis e susceptíveis a doenças durante o manejo.

Yan et al., (2006) avaliaram a dieta, densidade de estocagem, intensidade luminosa, filtração da água, troca de água e sedimento, tendo obtido que a densidade de 5-10 larvas.mL⁻¹ mantém um crescimento normal em larvas de *Ruditapes philippinarum*. Para larvas de *Meretrix meretrix* a densidade de estocagem recomendada é de 10-20 larvas.mL⁻¹ (LIU et al., 2006), enquanto que para larvas de *Clinocardium nuttallii* a densidade de estocagem deve ser igual ou inferior a 4 larval.mL⁻¹ e ao se aproximar da metamorfose para a pé de véliger, reduz a densidade para 2,5 larvas.mL⁻¹ (LIU et al., 2010).

Quando as larvas de moluscos bivalves realizam a metamorfose para a etapa pé de véliger, estas tornam-se mais resistentes ao manejo e passam a ser cultivadas em densidades mais elevadas, por um período maior de tempo (semanas) até alcançarem o tamanho de semente (1mm) quando poderão ser cultivadas a campo (NICOLAS e

ROBERT, 2001; EPELBAUM et al., 2011; LIU et al., 2011). Portanto avaliar a densidade de estocagem durante a fase pós-larval, tem como finalidade promover um ambiente de cultivo adequado para que estas alcancem a fase de semente em um menor tempo possível.

Para larvas de *Clinocardium nuttallii* é recomendado a densidade de estocagem de 160 ind.cm⁻² até alcançarem 1 mm de comprimento, passando para 40 ind.cm⁻² até 2 mm, depois 20 ind.cm⁻² até 3 mm, finalizando com 10 ind.cm⁻² até 4 mm (LIU et al., 2011). Resultados similares foram encontrados por Jones et al. (1993), sendo 150-200 pé de veliger.cm⁻² de Manila clam *Tapes philippinarum* em down-wellers.

Uma larvicultura comercial tem como meta melhorar o cultivo, aumentando a produção por unidade em um volume limitado de água com o menor custo possível. Nas larviculturas de moluscos é comum utilizar os sistemas de fluxo contínuo de água, diferenciados pelo sentido da corrente, sentido de cima para baixo (Down-welling) e de baixo para cima (Up-welling). Contudo quando se utiliza unidades de pequeno volume, o sistema de cultivo fechado com circulação da água também é bastante utilizado.

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5. - Artigo científico

5. 1. - Artigo científico I

Artigo científico encaminhado a Revista Latin American Journal of Aquatic Research.

Todas as normas de redação e citação, deste capítulo, atendem as estabelecidas pela referida revista (em anexo).

1 **Growth and survival of *Anomalocardia brasiliiana* larvae (Bivalvia:**
2 **Veneridae) fed microalgal diets**

3 **ABSTRACT.** Laboratory production of bivalve molluscs depends of the use of
4 microalgae. For this reason, the aim of this study was to evaluate the effect of using
5 different microalgal diets on growth and survival of *Anomalocardia brasiliiana* larvae
6 for 15 days of cultivation, between the trochophore and pediveliger stages. The diets
7 were evaluated in two separate experiments. The first experiment tested the microalgae
8 *Isochrysis galbana* (Ig), *Phaeodactylum tricornutum* (Phaeo), *Chaetoceros calcitrans*
9 (Cca) and the combinations (Ig + Cca), (Ig + Phaeo) and (Cca + Phaeo). The second
10 tested the microalgae *C. calcitrans* (Cca), *Pavlova lutheri* (Pl) and the combination (Cca
11 + Pl). The microalgae *I. galbana* when given alone had lower survival and growth when
12 compared to other diets tested. The use of diatoms *P. tricornutum* and *C. calcitrans* in
13 this bivalve diet alone or combined other microalgae achieved better values for survival
14 and growth. The diet (Cca + Pl) showed higher data concerning the growth and survival
15 of *A. brasiliiana* larvae ($261.67 \pm 9.64 \mu\text{m}$ and $31.50 \pm 0.87 \%$). The use of microalgae
16 *C. calcitrans* combined with *P. lutheri* is recommended to provide a good development
17 in *A. brasiliiana* larvae. Alone, microalgae *P. lutheri* and *C. calcitrans* no showed
18 significant difference, however, there is a synergistic effect when they are used in
19 combination. The use of a specific diet of *P. lutheri* combined with *C. calcitrans* is
20 recommended to provide satisfactory performance in *A. brasiliiana* larvae, cultured from
21 the trochophore stage to pediveliger.

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23 **Keywords:** *Anomalocardia brasiliiana* microalgae, larvae, growth, clam, diet,
24 pediveliger.

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34 **Efecto de las dietas de microalgas en crecimiento larvario y la supervivencia**
35 **de *Anomalocardia brasiliiana* (Bivalvia: Veneridae)**

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37 **RESUMEN.** La producción de moluscos bivalvos en laboratorio depende del uso de
38 microalgas. En este sentido, el objetivo de este estudio fue evaluar el efecto del uso de
39 diferentes dietas de microalgas sobre crecimiento y supervivencia de larvas de
40 *Anomalocardia brasiliiana* durante los 15 días de cultivo entre las fases trocófora y
41 pediveliger. Las dietas fueron evaluadas en dos experimentos separados. En el primer
42 experimento, se evaluaron las microalgas *Isochrysis galbana* (Ig), *Phaeodactylum*
43 *tricornutum* (Phaeo), *Chaetoceros calcitrans* (Cca) y las combinaciones (Ig + Cca), (Ig
44 + Phaeo) y (Cca + Phaeo). En el segundo experimento, las microalgas, *C. calcitrans*
45 (Cca), *Pavlova lutheri* (Pl) y su combinación (Cca + Pl). En el primer experimento, las
46 dietas que contienen *C. calcitrans* mostraron la mayor longitud de la concha al final del
47 cultivo, con una diferencia significativa entre Cca e Ig, donde la longitud promedio final
48 fue de $251,33 \pm 5,66 \mu\text{m}$ y $230,00 \pm 7,42 \mu\text{m}$, respectivamente. Para el segundo
49 experimento, con la dieta (Cca + Pl) se obtuvieron los mejores resultados para
50 crecimiento y supervivencia del cultivo de larvas de *A. brasiliiana* ($261,67 \pm 9,64 \mu\text{m}$ y
51 $31,50 \pm 0,87 \%$). Cuando las microalgas se usan de manera aislada *P. lutheri* y *C.*
52 *calcitrans* no presentan significativas diferencias. Sin embargo hay un efecto sinérgico
53 cuando se utilizan en combinación. El uso de la dieta específica de la especie de *P.*
54 *lutheri* combinada con *C. calcitrans*, se recomienda para proporcionar un rendimiento
55 satisfactorio en las larvas de *A. brasiliiana*, cultivadas a partir de la etapa trocófora paseo
56 pediveliger.

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58 **Palabras clave:** *Anomalocardia brasiliiana*, microalgas, larvas, crecimiento, molusco,
59 dieta, pediveliger

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INTRODUCTION

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68 In 2010, global production of marine molluscs reached 13.9 million tons, and
69 75.5% of all aquatic organisms produced by mariculture in the world were molluscs,
70 mostly bivalves, particularly oysters, mussels, clams and cockles (FAO, 2012). In
71 Brazil, mollusc culture is still limited to a few species including Mytilidae *Perna perna*,
72 Ostreidae *Crassostrea gigas*, *Crassostrea rhizophorae* and *Crassostrea brasiliiana* and
73 in the Pectinidae family *Nodipecten nodosus*. Considering the potential of various
74 species for cultivation, opportunity exists for the diversification of aquaculture.

75 Brazilian northeast region has favorable climatic conditions for the development
76 of mollusc culture, but hatcheries have been established for few native species and are
77 limited mainly to oysters, *C. rhizophorae* and *C. brasiliiana*. There is a need to develop
78 new technologies for aquaculture of other native species, such as the bivalve
79 *Anomalocardia brasiliiana* (Gmelin, 1791).

80 *Anomalocardia brasiliiana* is a resource of great importance for Brazilian coastal
81 communities (Silva-Cavalcanti & Costa, 2009; Oliveira *et al.*, 2011). Laboratory
82 production of its seed could be a management tool for restocking the heavily exploited
83 populations along the Brazilian coastline, as well as for diversification of mariculture.

84 One of the obstacles to establish successful larval cultures is the availability of an
85 appropriate diet (Ponis *et al.*, 2006; Liu *et al.*, 2009; Pettersen *et al.*, 2010). Microalgae
86 have been the main food source for larvae and seeds in bivalve hatchery (Helm &
87 Bourne, 2004). In addition, microalgae can provide improved water quality by
88 absorbing toxic nitrogen products (ammonia and nitrite) and combating pathogenic
89 bacteria for produce antibiotic substances (Lavens & Sorgeloos, 1996).

90 It is important that the microalgae are of the appropriate size to allow ingestion,
91 and that they meet the nutritional requirements of the cultivated species. The cost of the
92 microalgae must also be competitive with other species and be easy to produce on a
93 commercial scale (Heasman *et al.*, 2000; Ponis *et al.*, 2008). Most microalgae used in
94 bivalve hatcheries are species from the genera *Isochrysis*, *Pavlova*, *Chaetoceros*,
95 *Phaeodactylum*, *Skeletonema*, *Thalassiosira*, *Dunaliella*, *Nannochloris*, *Tetraselmis* and
96 *Rhodomonas* (Coutteau & Sorgeloos, 1992; Moueza *et al.*, 1999; Brown, 2002; Helm &
97 Bourne, 2004; Cragg, 2006; Ponis *et al.*, 2008; Liu *et al.*, 2009).

98 Precise knowledge of microalgae as well as proper management is essential to
99 provide greater shellfish larvae survival and growth. There isn't studies on the use of

100 microalgae in hatcheries of this species, and the purpose of this study was to evaluate
101 survival and growth of larvae of *A. brasiliiana* when fed different microalgal diets.

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MATERIALS AND METHODS

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105 A total of 500 adults longer than 20 mm were collected on the beach of Mangue
106 Seco (07°49' 44,19"S, 035°50' 03,06"W), Igarassu municipality, 30 km from Recife,
107 Pernambuco state, Brazil, transported to the Laboratory of Sustainable Mariculture
108 (LAMARSU). and acclimated during 24 h in 500 liter tanks at 25 °C, 30 g L⁻¹ salinity
109 and 6 mg L⁻¹ mean dissolved oxygen. .

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After this period, they were fed twice daily a mixture of *Isochrysis galbana* and
Chaetoceros calcitrans at a cell ratio of 1:1, with a total ration of 20x10⁴ cell. mL⁻¹ .day⁻¹.
In the first experiment, after 10 days, spawning occurred spontaneously, and the
fertilized eggs were filtered 50 µm mesh opening and kept in a 500 L tank. Yet to the
second experiment, spawns occurred on the same day that the breeding arrived at the
laboratory, through the induction, release of gametes and gradual temperature increase
(1 °C.h⁻¹)

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After 24 hours D-veliger larvae (n = 30) had an average length of 69.94 ± 1.54
mm and were stocked with an initial density of five larvae.mL⁻¹, in triplicate for each
treatment, in plastic containers with two liters of seawater (three µm filtered and UV-
treated).

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Temperature and salinity were measured daily, oxygen twice a day and the water
of larval cultures was renewed completely every three days. Feeding was once a day,
and microalgae supplied to the larvae were from three days-old cultures, in exponential
growth.

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The microalgae used in the experiment were obtained from LAMARSU stock
strains. The seawater with salinity of 32 ± 2 g L⁻¹ was filtered through paper with 1 µm
pores, autoclaved, and enriched with a Conway sterilized medium (Walne, 1966),
supplemented with sodium metasilicate (40 mg L⁻¹) to provide a silica source for the
diatom *C. calcitrans* and *P. tricornutum*.

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The effect of microalgal diets on larval growth of *A. brasiliiana* was evaluated in
two completely randomized experiments. The first experiment tested the microalgae *I.*
galbana (Ig), *Phaeodactylum tricornutum* (Phaeo) and *Chaetoceros calcitrans* (Cca)
and the combinations (Ig + Cca), (Ig + Phaeo) and (Cca + Phaeo). The larval rearing

134 period was 15 days, starting with the D-veliger larval stage and ending with pediveliger
135 larvae. The total algal cell density provided was 30×10^3 cell mL⁻¹ and for bialgal diets a
136 1:1 ratio was used.

137 The second experiment evaluated the algae *C. calcitrans* (Cca), *P. lutheri* (Pl)
138 and the combination (Cca + Pl). *C. calcitrans* was assessed again because it had the
139 highest relative growth in the first experiment and was characterized as an excellent diet
140 for weight gain in molluscs (Rivero-Rodriguez *et al.*, 2007). The microalgae *P. lutheri*
141 has been described in some studies (Ponis *et al.*, 2006, 2008) as an excellent diet for
142 bivalve molluscs. The methodology and algal density provided in the second experimet
143 were the same adopted for the first experiment.

144 To assess larval survival at the end of experiment the entire volume of each
145 experimental unit was filtered. The larvae were concentrated in a 50 mL container, from
146 which one milliliter (1 mL) samples were drawn. The larval counting was done with a
147 Sedgewick-Rafter counting chamber and optical microscope, three samples of each
148 experimental unit were analyzed.

149 For the evaluation of larval growth, one milliliter samples of each experimental
150 units were removed on the first and last day of the experiment and images of 30 larvae
151 chosen at random were obtained using an optical microscope coupled to a camera lens,
152 and their length (maximum anterior-posterior dimension) and width (maximum dorsal-
153 ventral dimension) were measured using version 2.0 software ImageTool (Texas
154 University, Health Science Center, San Antonio, USA)..

155 Relative growth (K) was calculated using the equation $K = (\ln L2 - \ln L1) / t$
156 (Walen, 1963), where *L1*, *L2* represent the lengths at the beginning and end of the
157 experiment in μm , respectively, and *t* is the duration of the experiment in days.

158 Survival data, length, width and relative growth between diets, generated in both
159 experiments, were previously checked for normality using the Kolmogorov-Smirnov
160 test and for homogeneity of variance with Cochran's C test. Analysis of Variance
161 (ANOVA) was used to determine the effect of diets on the growth and survival of larvae
162 over time of cultivation. Duncan's test was performed to detect the mean levels which
163 differed significantly between treatments. The level of significance was $P < 0.05$.

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RESULTS

The variables of temperature (°C), salinity (g L⁻¹) and dissolved oxygen (mg L⁻¹) of the water were maintained within acceptable limits for shellfish growing. The temperature ranged from 25.05 °C to 25.60 °C, salinity from 29.88 g L⁻¹ to 30.18 g L⁻¹ and dissolved oxygen had a minimum value of 5.89 mg L⁻¹ and a maximum of 6.66 mg L⁻¹.

First Experiment

There was a significant difference in larval survival of *A. brasiliiana* as assessed at the end of cultivation, between diets (ANOVA). The Phaeo diet had the highest survival of approximately 25%, which was not significantly different of Cca, Ig + Phaeo and Ig + Cca diets. The lowest survival values were found in diets of Ig and Cca + Phaeo which averaged 6.83% and 5.27%, respectively, significantly lower than the Phaeo diet (Table 1).

No significant differences in shell width was found in larvae fed with the different algal diets tested (Table 1). Significant differences were observed between the relative growth rate in the diets tested (Table 1). The Cca and Ig + Phaeo diets had higher relative growth, than Ig diet.

Larval survival obtained in the first experiment reached a maximum value of 24.83 ± 3.03 % in the Phaeo diet. The survival of larvae fed with *C. calcitrans* (12.33%) was not significantly different from other diets tested (5.2- 4.83%). Larvae fed with microalgae *I. galbana* and a mixture of *C. calcitrans* with *P. tricornutum* had the lowest survival.

By analyzing the larval length as a growth factor, it was possible to determine significant differences between the diets tested ($P < 0.05$). The Cca diet had the greatest shell length at the end of cultivation (251.33 ± 5.66 µm), differing significantly from the Ig diet, which had the lowest growth in length (230.00 ± 7.42 µm). The other tested diets showed no significant difference regarding the growth in length (Fig. 1).

The microalgae *I. galbana* when given alone had lower survival and growth when compared to other diets tested. The use of diatoms *P. tricornutum* and *C. calcitrans* in this bivalve diet alone or combined other microalgae achieved better values for survival and growth.

202 Insert Table 1.

203

204 Insert Figure 1

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206 **Second experiment**

207 Larval survival of *A. brasiliiana* assessed at the end of cultivation was
208 significantly different among the three diets tested (Table 2). The Cca + Pl diet had the
209 highest survival with a mean of 31.50%.

210 When analyzing the width of the shell as a growth parameter, no significant
211 differences were found between the larvae fed with Cca + Pl and Cca diets (Table 2).
212 The combined use of microalgae *C. calcitrans* and *P. lutheri* led to the highest growth
213 in width after 15 days of culture with $242.00 \pm 10.02 \mu\text{m}$.

214 There were significant differences in the relative growth rate in larva with
215 different diets (Table 2). The Cca + Pl diet showed higher relative growth significantly
216 different than the Cca diet ($P < 0.05$), demonstrating its positive effect on the growth of
217 *A. brasiliiana* larvae when microalgae *C. calcitrans* and *P. lutheri* are used in
218 combination. Confirming the positive effect observed in larval survival, with a Cca + Pl
219 diet.

220 By analyzing the length of the larvae, significant differences were found between
221 the diets tested ($P < 0.05$). The diet containing both algae *C. calcitrans* and *P. lutheri*
222 had the highest growth in length of larvae at the end of the experiment, with a
223 significant difference from the Cca diet (Fig. 2).

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225 Insert Table 2.

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229 **DISCUSSION**

230

231 The water parameters, oxygen, temperature and salinity remained within the
232 limits recommended for bivalve culture during all the experimental period. The
233 absorption efficiency of some microalgae species, due to the maintenance of water
234 quality standards can explain good growth in bivalves (Grizzle *et al.*, 2001; Resgalla Jr.
235 *et al.*, 2007).

236 The use of a closed recirculation system, with daily water exchange can lead to
237 better survival of *A. brasiliiana* larvae. Liu *et al.* (2009) were able to increase survival to
238 68.0% by using a continuous water renewal system (50% daily and 100% every three
239 days) when *Clinocardium nutallii* larvae were fed some diet for nine days of cultivation.
240 Thus, the low survival rate (25%) observed in *A. brasiliiana* larvae may be related to the
241 use of closed cultivation without daily renewal, but with 100% renewal every three
242 days.

243 In addition to maintaining water quality, a second factor of great importance for
244 successful development of shellfish larvae is the nutritional value of the diet. Lourenço
245 (2006) revised the chemical composition of some marine microalgae species, *C.*
246 *calcitrans* (34% protein, 16% fat and 6% carbohydrate), *P. tricornutum* (30% protein,
247 14% fat and 8.4% carbohydrate), *I. galbana* (29% protein, 23% fat and 12.9%
248 carbohydrate) and *P. lutheri* (29% protein, 12% lipids and 9% carbohydrate), indicating
249 similar chemical compositions. However, slight differences in the concentration of fatty
250 acids, vitamins and amino acids supplied in the diet can compromise survival and larval
251 development.

252 The relative growth of larvae fed with Cca and Ig + Phaeo diets were higher only
253 than that of larvae fed an Ig diet, with no significant differences with the other diets
254 tested. The microalga *C. calcitrans* is considered the most suitable for feeding bivalve
255 larvae (Brown & Robet, 2002), not only because of its biochemical composition, but
256 also because of its cell size, digestibility and absence of toxins (Pettersen *et al.*, 2010).

257 Studies have found that the use of the microalgae *P. tricornutum* for feeding
258 other bivalves, causes slow growth (Epifanio *et al.*, 1981; Albentosa *et al.*, 1996;
259 Rivero-Rodriguez *et al.*, 2007), because it is difficult to digest (Rivero-Rodriguez *et al.*,
260 2007), probably due to its lack of tryptophan (Epifanio *et al.*, 1981). Tang *et al.* (2006)
261 achieved a relatively low growth in *Meretrix meretrix* larvae when fed *Phaeodactylum*
262 *tricornutum* (0.0388) and *Pavlova viridis* (0.0361).

263 A diet composed of the microalgae *C. calcitrans* showed the longest shell at the
264 end of the cultivation (251.33 ± 5.66 μm). Rivero-Rodriguez *et al.* (2007), evaluating
265 the growth of *Crassostrea corteziensis* seeds found significant growth when using *C.*
266 *calcitrans* alone or in combination with other diets, which was up to twice the growth
267 when this microalga is present in the diet. This growth was affirmed to be related to the
268 fact that *C. calcitrans* contains high levels of arachidonic acid (ARA). These authors

269 confirmed that the mixed use of *C. calcitrans* and *C. muelleri* is the second best diet in
270 terms of weight gain for *C. corteziensis*. Similar results were found by Liu *et al.* (2009)

271 In the second experiment in this study, there was an increase in survival of *A.*
272 *brasiliiana* larvae when *C. calcitrans* and *P. lutheri* microalgae were used in
273 combination, reaching average survival above 31%. Ponis *et al.* (2008), evaluating the
274 effect of *P. lutheri* in *Crassostrea gigas* larvae obtained survival above 78% when *C.*
275 *calcitrans* was added to the diet. This corroborates our findings that after 15 days of
276 culture, larvae fed a bialgal Cc + Pl diet had better survival that significantly differed
277 from those fed only monoalgal diets (Cca and Pl). Other studies have also achieved
278 good results when adding other species of diatom microalgae (Epifanio, 1979;
279 Romberger & Epifanio, 1981; Laing & Millican, 1986; O'Connor & Heasman, 1997),
280 which has been attributed to better essential nutrient balance (Webb & Chu, 1983).

281 The relative growth for the Cca + Pl diet (0.0867) was higher than the other diets
282 tested in both experiments. There was no significant difference in growth when the
283 microalgae *P. lutheri* and *C. calcitrans*, were used singly, however there is a synergistic
284 effect when they are used together. The final average length of *A. brasiliiana* larvae fed
285 a combination of *P. lutheri* and *C. calcitrans* was significantly higher than larvae fed
286 only *C. calcitrans*.

287 Bialgal diets are often used in feeding bivalve larvae, and it is common to
288 combine species, using a flagellate with a diatom, to maximize growth and larval
289 development (Galley *et al.*, 2010; Liu *et al.*, 2009; Martínéz-Fernandez & Southgate,
290 2007). The flagellated species commonly used are *Isochrysis galbana* and *Pavlova*
291 *lutheri* and the diatoms include *Chaetoceros calcitrans*. Spolaore *et al.* (2006) affirm
292 that the combination of different algae species offers a better nutritional balance and
293 improves animal growth compared to a monoalgal diet, which agrees with the result of
294 this experiment, which found that the bialgal diet (Cca + Pl) led to better growth at the
295 end of cultivation.

296 In hatchery of the mollusc *Pinctada margaritifera*, Martínéz-Fernandez &
297 Southgate (2007) had a higher growth rate in the D-larvae phase when cultured with the
298 ternary combination of *Pavlova* sp./*Pavlova salina*/*I. galbana*. However, the growth
299 rate was not significantly higher than those of larvae fed with the binary combination of
300 *P. salina*/*I. galbana* which has been used as a "standard diet" for *P. margaritifera* larvae
301 (Southgate & Beer, 1997). Martínéz-Fernandez & Southgate (2007) suggest that feeding
302 a single species of microalgae to *P. margaritifera* larvae may be more practical during

303 the first 10 days in a hatchery. However, the addition of diatom microalgae to the diet
304 increased growth rate and survival in umbonate larvae of *P. margaritifera* when
305 compared to treatments without diatoms.

306 Protein and vitamin content are important factors for determining the nutritional
307 value of microalgae. Furthermore, high amounts of polyunsaturated fatty acids (eg.
308 eicosapentaenoic [EPA], arachidonic acid [AA] and docosahexaenoic acid [DHA]),
309 (Hemaiswarya *et al.*, 2011), can lead to better growth and survival of larvae fed with the
310 microalgae *P. lutheri*, which is rich in DHA/EPA, and *C. calcitrans*, which is used to
311 increase vitamin levels (Hemaiswarya *et al.*, 2011).

312 *Pavlova lutheri* is used frequently in the diet of the reproductive stock of bivalves,
313 such as oysters, clams, mussels and scallops (Hemaiswarya *et al.*, 2011). *C. calcitrans* is
314 rich in fatty acids such as arachidonic acid (ARA) and vitamins, and is widely used in
315 bivalve hatchery (Hemaiswarya *et al.*, 2011; Rivero-Rodriguez *et al.*, 2007).

316 This study found that monoalgal and bialgal diets present satisfactory results in
317 terms of survival. Prymnesiophyceae *P. lutheri* is commonly used in aquaculture as live
318 food for marine invertebrates (molluscs, crustaceans, zooplankton) and particularly for
319 bivalves (larvae, juveniles and breeding stock) (Webb & Chu, 1983; Borowitzka, 1997;
320 Wikfors & Onho, 2001; Brow, 2002; Rico-Villa *et al.*, 2006), but its use alone may
321 present low growth when compared to use with other diatoms (Rico-Villa *et al.*, 2006;
322 Ponis *et al.*, 2008). The microalga *C. calcitrans* is used in some studies as a reference
323 diet that offers good results for growth and survival (Ponis *et al.*, 2006; Ponis *et al.*,
324 2008).

325 Our results confirm the potential of the microalgae *C. calcitrans* for offering
326 good growth of mollusc larvae. In hatching the mussel *Mytilus galloprovincialis*, was
327 observed an increase in larval mortality when a diet of *C. calcitrans* was replaced with
328 *C. muelleri*, with a consequent decrease in DHA content in the larvae (Pettersen *et al.*,
329 2010).

330 A comparison of the results of the first and second experiment in the current
331 study indicated that the Cca + Pl diet had the best results in both survival ($31.50 \pm$
332 0.87%) and growth ($261.67 \pm 9.64 \mu\text{m}$). The combination of the microalgae *C.*
333 *calcitrans* and *P. lutheri* proved to be an excellent diet for the larval culture of *A.*
334 *brasiliiana*. Used alone, the microalgae *C. calcitrans* and *P. lutheri* showed no significant
335 difference, however, there is a synergistic effect when they are used in combination.

336 The use of microalgae *C. calcitrans* combined with *P. lutheri* can provide good
337 growth in *Anomalocardia brasiliiana* larvae. The study found that the use of bialgal
338 diets leads to better growth and survival, and is more recommended due to better
339 nutritional balance. There is a need for new studies to improve dietary practices and
340 increase survival between the larval stages, trochophore and pediveliger.

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471 **Table 1.** Mean (\pm SE) survival, width and relative growth (K) of larvae of *A. brasiliana*
 472 fed different diets over 15 days of culture. Cca: *Chaetoceros calcitrans*, Ig: *Isochrysis*
 473 *galbana*; Phaeo: *Phaeodactylum tricornutum*, mixed diets Cca+Ig: *C. calcitrans* and *I.*
 474 *galbana*; Cca + Phaeo: *C. calcitrans* and *P. tricornutum*, and Ig + Phaeo: *I. galbana* and
 475 *P. tricornutum*. Different letters in the same column have a significant difference ($P <$
 476 0.05).

Diets	Survival (%)	Width (μm)	K ($\mu\text{m}\cdot\text{day}^{-1}$)
Ig	6.83 \pm 1.92 ^b	212.33 \pm 6.79	0.0779 \pm 0.0022 ^b
Cca	12.33 \pm 5.84 ^{ab}	229.00 \pm 5.19	0.0848 \pm 0.0015 ^a
Phaeo	24.83 \pm 3.03 ^a	222.33 \pm 5.27	0.0830 \pm 0.0020 ^{ab}
Ig + Cca	13.50 \pm 4.91 ^{ab}	226.33 \pm 5.04	0.0821 \pm 0.0020 ^{ab}
Ig + Phaeo	18.27 \pm 2.42 ^{ab}	226.00 \pm 7.81	0.0839 \pm 0.0019 ^a
Cca + Phaeo	5.27 \pm 1.36 ^b	220.00 \pm 6.49	0.0807 \pm 0.0020 ^{ab}

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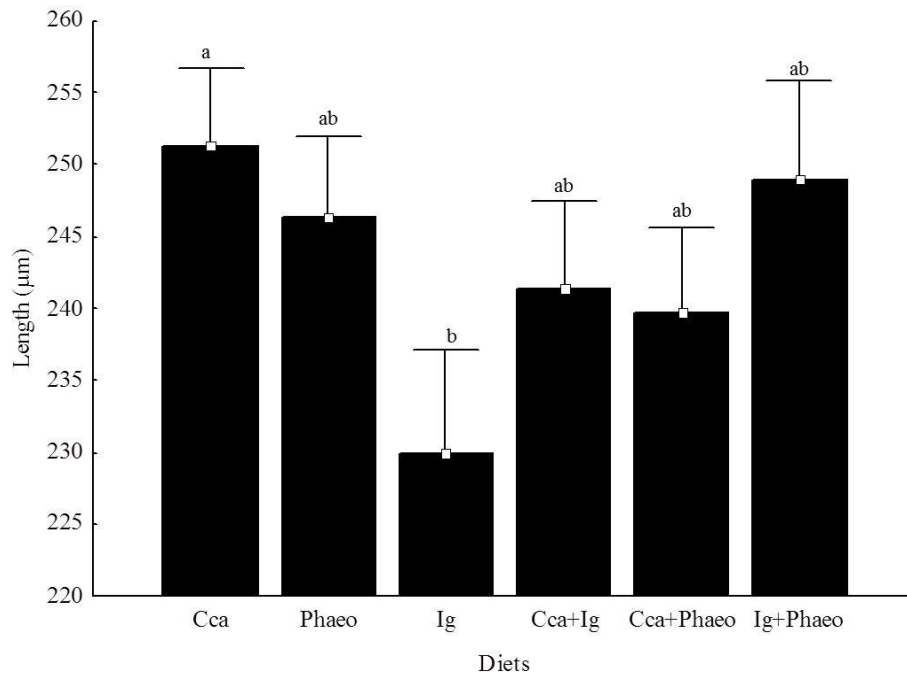
478

479 **Table 2.** Mean (\pm SE) survival, width and relative growth (K) of larvae of *A. brasiliana*
 480 fed different diets over 15 days of culture. Cca: *Chaetoceros calcitrans*, Pl: *Pavlova*
 481 *lutheri*, mixed diet Cca+Pl: *C. calcitrans* and *P. lutheri*. Different letters in the same
 482 column have a significant difference ($P <$ 0.05).

Diets	Survival (%)	Width (μm)	K ($\mu\text{m}\cdot\text{day}^{-1}$)
Cca	4.42 \pm 1.58 ^c	211.67 \pm 8.60 ^b	0.0781 \pm 0.0028 ^b
Pl	16.80 \pm 5.63 ^b	219.33 \pm 6,69 ^{ab}	0.0805 \pm 0.0021 ^{ab}
Cca+Pl	31.50 \pm 0.87 ^a	242.00 \pm 10.02 ^a	0.0867 \pm 0.0024 ^a

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486 **Figure 1.** Mean length of larvae *A. brasiliiana* fed different diets of single or mixed
487 algae over a period of 15 days of growth. Cca: *Chaetoceros calcitrans*, Ig: *Isochrysis*
488 *galbana*; Phaeo: *Phaeodactylum tricornutum*, mixed diets of Cca + Ig: *C. calcitrans* and
489 *I. galbana*; Cca + Phaeo: *C. calcitrans* and *P. tricornutum*, and Ig+Phaeo: *I. galbana*
490 and *P. tricornutum*. Means with different letters differ significantly ($P < 0.05$).

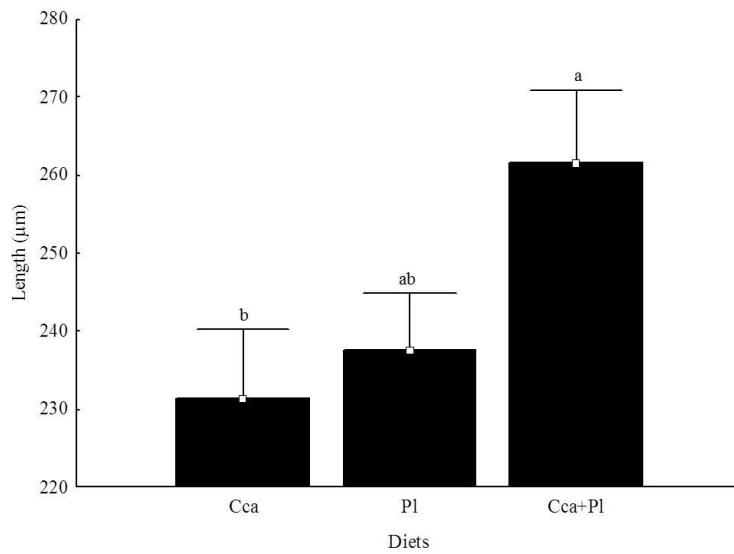
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497 **Figure 2.** Mean length of larvae of *A. brasiliana* fed microalgal diets for a period of 15498 days. Ig: *Isochrysis galbana*; Cca: *Chaetoceros calcitrans*; Pl: *P. lutheri*. Means with499 different letters have a significant difference ($P < 0.05$).

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5.2. - Artigo científico II

Artigo científico a ser encaminhado a Revista Aquaculture.

Todas as normas de redação e citação, deste capítulo, atendem as estabelecidas pela referida revista (em anexo).

1 Efeito da densidade de estocagem no crescimento e sobrevivência de pós-larvas de
2 *Anomalocardia brasiliana* (Bivalvia: Veneridae)

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26 Resumo

27 As pós-larvas de *Anomalocardia brasiliiana* foram cultivadas por 28 dias para avaliação
28 do efeito da densidade de estocagem no crescimento e sobrevivência, sendo alimentados
29 com a mistura das microalgas *Pavlova lutheri* e *Chaetoceros calcitrans*. Três
30 densidades de estocagem foram testadas 40, 80 e 160 ind.cm⁻², em unidades
31 experimentais de pequeno volume (2 litros) com sistema de cultivo estático e renovação
32 total da água feita a cada 48h. O delineamento experimental utilizado foi inteiramente
33 casualizado com três tratamentos (densidades) e três repetições cada. Os animais
34 cultivadas na densidade de estocagem de 40 ind.cm⁻² alcançaram 1mm de comprimento
35 aos 24 dias de cultivo. Ao final dos 28 dias de cultivo apenas 18% dos animais
36 cultivadas na densidade de 80 ind.cm⁻² apresentavam comprimento de 1mm, enquanto
37 que o tratamento com 160 ind.cm⁻² 100% não atingiram 1mm de comprimento. Ao
38 avaliarmos todo o período experimental (0-28 dias) a densidade de 40 ind.cm⁻²
39 apresentou a maior taxa de crescimento específico diária, 4,98±0,08 %.dia⁻¹. A taxa de
40 sobrevivência das pós-larvas para as menores densidades (40 e 80 ind.cm⁻²)
41 apresentaram maiores médias (53,24 ± 4,60 % e 52,95 ± 3,32 %) respectivamente,
42 diferindo significativamente da maior densidade de estocagem com 31,54 ± 0,70 %. Na
43 larvicultura das pós-larvas de *A. brasiliiana* deve-se realizar o manejo da densidade de
44 estocagem no decorrer do crescimento deste molusco. A densidade de 160 ind.cm⁻²
45 pode ser utilizada até que as pós-larvas alcancem 600µm de comprimento, pós-larvas
46 maiores que 600 µm devem ser cultivadas na densidade de 40 larvas.cm⁻² para manter a
47 taxa de crescimento máximo diária.

48 Palavras-chave: crescimento, sobrevivência, pós-larvas, larvicultura.

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52 1. Introdução

53 O marisco, *Anomalocardia brasiliana*, é um recurso pesqueiro de grande
54 importância para os estados brasileiros, principalmente, nas comunidades costeiras da
55 região nordeste (Nishida et al., 2004; Silva-Cavalcanti & Costa, 2009; Oliveira et al.,
56 2011; Lavander et al., 2011; Oliveira et al., 2014). Pertence à família Veneridae, vive
57 em profundidades de 0,5 a 1,5 m, superficialmente escavado na areia próximo ao
58 manguezal, é sensível a variações ambientais, com alta mortalidade devido às chuvas,
59 que causam grandes flutuações no tamanho e na distribuição das populações (Monti et
60 al., 1991; Mouëza et al., 1999; Oliveira et al., 2014).

61 A região nordeste do Brasil apresenta condições climáticas favoráveis para o
62 desenvolvimento da malacocultura, no entanto para as espécies nativas ainda é pouco
63 desenvolvida a larvicultura. A espécie *A. brasiliana* apresenta potencial para ser
64 cultivada, contudo o estabelecimento de um protocolo de produção larval para uma
65 nova espécie exige a avaliação de diversos fatores tais como: densidade de estocagem,
66 qualidade e quantidade da dieta a ser fornecida, e parâmetros de qualidade da água de
67 cultivo (temperatura e salinidade).

68 Foram realizados diversos estudos para estabelecimento de um protocolo de
69 produção de larvas de moluscos, tais como: bivalves sedentários (Liu et al, 2002; Liu et
70 al, 2006; Yan et al, 2006; Liu et al, 2008; Liu et al, 2010), mexilhões (Galley et al,
71 2010; Pettersen et al, 2010), ostras (Rico-Villa et al, 2006) e vieiras (Rupp e Parsons,
72 2004; Gouda et al, 2006). Observou-se que os fatores que influenciam o crescimento e a
73 sobrevivência das larvas é essencial para o sucesso de uma larvicultura.

74 Uma importante variável a ser considerada é a densidade de estocagem, pois afeta
75 no crescimento e sobrevivência de larvas, juvenis e adultos bivalves tem sido
76 amplamente estudado (Fréchette, 2005), principalmente para aquelas espécies com

77 importância comercial para aquicultura. A maioria dos estudos avaliaram o período de
78 crescimento nas fases de D véliger até a metamorfose para pé de véliger (Liu et al.,
79 2006; Yan et al., 2006; Liu et al., 2010), pois são fases críticas, onde as larvas são
80 frágeis e susceptíveis a doenças durante o manejo.

81 Quando as larvas realizam metamorfose para a etapa pé de véliger, ficam mais
82 resistentes ao manejo e passam a ser cultivadas em densidades mais elevadas, por um
83 período maior de tempo (semanas) até alcançarem o tamanho de semente (1mm) quando
84 poderão ser transferidas para o campo (Nicolas e Robert, 2001; Epelbaum et al., 2011;
85 Liu et al., 2011). A Avaliação da densidade de estocagem durante a fase pós-larval tem
86 como objetivo promover um ambiente de cultivo adequado para que estas alcancem a
87 fase de semente em um menor tempo possível.

88 Uma larvicultura comercial tem por meta melhorar os dados de cultivo,
89 aumentando a produção em um volume limitado de água com o menor custo possível.
90 Nas larviculturas de moluscos é comum utilizar os sistemas de fluxo contínuo de água,
91 diferenciados pelo sentido da corrente, de cima para baixo (Down-welling) e de baixo
92 para cima (Up-welling). Contudo, quando se utiliza unidades de pequeno volume, o
93 sistema de cultivo fechado com circulação da água também é bastante utilizado.

94 Neste sentido, este estudo tem como objetivo avaliar o efeito da densidade de
95 estocagem sobre o crescimento e a sobrevivência de pós-larvas de *Anomalocardia*
96 *brasiliiana* em um sistema de cultivo fechado.

97

98 **2. Material e Métodos**

99 Para obtenção de larvas de *A. brasiliiana* foi necessário à coleta de indivíduos
100 adultos no ambiente natural. Os reprodutores foram capturados na praia de Mangue
101 Seco (07° 49' 44,19"S e 035° 50' 03,06"O), litoral norte do estado de Pernambuco –

102 Brasil. Foram coletados 500 indivíduos adultos, com comprimento médio de $24,50 \pm 2,00$
103 mm e transportados ao Laboratório de Maricultura Sustentável (LAMARSU). No
104 laboratório os animais foram aclimatados em tanques de 500 litros com controle das
105 variáveis de qualidade da água, temperatura ($25\text{ }^{\circ}\text{C}$), salinidade (30 g.L^{-1}) e oxigênio
106 dissolvido médio de (6 mg.L^{-1}). Após a identificação dos mariscos que estavam com as
107 gônadas maduras foram realizados estímulos para desova tais como: aumento da
108 temperatura, oferta de alimento e lançamento de gametas na água. Após estes estímulos
109 foram obtidas as desovas. Os ovos fecundados foram filtrados através de uma malha
110 com abertura de $50\text{ }\mu\text{m}$ e mantidos em incubadoras com 30L de volume útil por 15 dias,
111 com troca total da água a cada 48h. Durante esse tempo as larvas foram alimentadas
112 com uma mistura das microalgas *Pavlova lutheri* e *Chaetoceros calcitrans*, na
113 proporção celular de 1:1 e a concentração algal foi diferenciada para cada estágio larval
114 e está descrito na Tabela 1.

115

116 Inserir Tabela 1

117

118 Larvas pé de véliger com 15 dias de vida foram mensuradas ($n=30$), obtendo-se
119 um tamanho médio de $307,89 \pm 50,92\text{ }\mu\text{m}$ de comprimento (medida entre as
120 extremidades ântero-posterior da concha). Em seguida, foram transferidas para as
121 unidades experimentais que eram constituídas de recipientes plásticos transparentes com
122 dois litros de volume útil, internamente suspenso um cilindro de PVC (diâmetro 10 cm;
123 área $78,5\text{ cm}^2$), sendo na base do cilindro colocado uma malha de $250\text{ }\mu\text{m}$ ficando a uma
124 distância de fundo de 5 cm e adicionado um airlift para circulação da água (Figura 1).

125

126 Inserir Figura 1

127

128 As pós-larvas foram cultivadas por um período de 28 dias, com avaliação de três
129 diferentes densidades de estocagem, 40 ind.cm⁻², 80 ind.cm⁻² e 160 ind.cm⁻², em
130 um sistema fechado de circulação da água com troca total a cada 48 horas. A água
131 utilizada na larvicultura foi previamente esterelizada por radiação ultravioleta, para
132 posterior utilização nas unidades experimentais. Diariamente foram aferidas as variáveis
133 temperatura, salinidade e oxigênio dissolvido. A qualidade da água nas unidades
134 experimentais foi mantida dentro dos limites aceitáveis para a larvicultura de bivalves,
135 com temperatura média de 25,89±0,50 °C, salinidade média de 28,5±0,88 g.L⁻¹ e com
136 média de oxigênio dissolvido em 5,66 ±0,55 mg.L⁻¹).

137 A alimentação foi realizada uma vez ao dia, com as microalgas *P. lutheri* e *C.*
138 *calcitrans*, fornecidas às pós-larvas a partir do terceiro dia de cultivo algal, na fase
139 exponencial da curva de crescimento, a uma concentração de 50x10³ cel.mL⁻¹. As
140 microalgas foram obtidas do banco de cepas do LAMARSU, e mantidas em tubos de
141 ensaio com água marinha de salinidade 32±2 g.L⁻¹ e enriquecida com meio Conway
142 esterelizado, sendo adicionado metassilicato de sódio (40 mg.L⁻¹) como fonte de sílica
143 no cultivo da diatomácea *C. calcitrans*.

144 Para a avaliação do crescimento larval, foram retiradas amostras de um mililitro
145 das unidades experimentais e com auxílio da câmara de Sedgewick-Rafter, imagens
146 foram tomadas utilizando uma câmara fotográfica objetiva acoplada a um microscópio
147 óptico. Foram realizadas fotografias aleatórias em 30 pós-larvas de cada amostra, a cada
148 semana. As medidas de comprimento (máxima dimensão antero-posterior) e largura
149 (máxima dimensão dorso-ventral) das larvas foram obtidas utilizando o software
150 ImageTool versão 2.0 (Texas University, Health Science Center, San Antonio, USA).

151 Durante o experimento a taxa de crescimento das pós-larvas foram estimadas
152 usando a seguinte fórmula:

$$153 \quad TCE = 100 (\ln C2 - \ln C1) / t,$$

154 Onde TCE é a taxa de crescimento específico (% dia⁻¹); *C1*, *C2* representam os
155 comprimentos no início e no fim do experimento em µm, respectivamente, e *t* é o tempo
156 de duração do experimento (dias).

157 Para a avaliação da sobrevivência larval no final do cultivo, foi realizada uma
158 filtragem de todo o volume de cada unidade experimental. As pós-larvas foram
159 concentradas em recipientes de 50 mL, e retiradas amostras de um mililitro (mL). A
160 contagem dos indivíduos foi realizada com auxílio da câmara Sedgewick-Rafter e
161 microscópio óptico, e analisado três amostras de cada unidade experimental.

162 Os dados de sobrevivência, comprimento e crescimento das densidades de
163 estocagem avaliadas, foram previamente checados quanto à normalidade dos dados
164 usando o teste de Kolmogorov-Smirnov e para homogeneidade de variância com o teste
165 de Cochran C. A Análise de Variância (ANOVA) foi usada para determinar o efeito das
166 densidades no crescimento e sobrevivência das pós-larvas ao longo do tempo de cultivo.
167 O teste de Tukey foi realizado para detectar qual média entre os tratamentos diferem
168 significativamente, a nível de significância de 5% ($p < 0,05$). Os dados estão
169 apresentados em média ± erro padrão.

170

171 **3. Resultados**

172 O comprimento médio das pós-larvas de *A. brasiliiana* estocadas em diferentes
173 densidades de estocagem, estão apresentados na Figura 2.

174 Ao término da primeira semana do experimento (7 dias) não foram encontradas
175 diferenças ($p > 0,05$) no comprimento dos animais entre as densidades testadas. A partir

176 da segunda semana o comprimento das pós-larvas foram menores a medida que se
177 aumentou a densidade de estocagem. Aos 21 dias de cultivo não foi identificado
178 diferença ($p>0,05$) no comprimento das pós-larvas cultivadas com as densidades 40
179 ind.cm⁻² e 80 ind.cm⁻². As pós-larvas de *A. brasiliiana* cultivadas na maior densidade de
180 estocagem (160 ind.cm⁻²) apresentaram menor média de comprimento ($p<0,05$),
181 tornando-se mais evidente ao final dos 28 dias de cultivo.

182 As pós-larvas cultivadas na densidade de 40 ind.cm⁻² alcançaram nos 24 dias de
183 cultivo 1mm de comprimento, e ao final dos 28 dias do experimento apenas 18% das
184 larvas cultivadas na densidade de 80 ind.cm⁻² apresentavam comprimento de 1mm,
185 enquanto que no tratamento com 160 ind.cm⁻² não atingiram este comprimento.

186

187 Inserir Figura 2

188

189 A tabela 2 apresenta a TCE (% dia⁻¹) das três diferentes densidades de estocagem
190 para cada período do cultivo. O rápido crescimento ocorreu para os primeiros sete dias
191 do experimento. A menor densidade (40 ind.cm⁻²) continuou apresentando o maior
192 crescimento até 14 dias de cultivo, diferindo significativamente das outras densidades
193 avaliadas ($p<0,05$). Aos 21 dias de cultivo as pós-larvas de todos os tratamentos
194 apresentaram uma queda acentuada na taxa de crescimento (Tabela 2).

195 As pós-larvas cultivadas na densidade de 40 ind.cm⁻² apresentam a maior taxa de
196 crescimento, diferindo significativamente das outras densidades testadas. Ao avaliarmos
197 todo o período experimental (0-28 dias) a densidade de 40 ind.cm⁻² permaneceu com a
198 maior taxa de crescimento específico diária.

199

200 Inserir Tabela 2

201

202 A sobrevivência larval das menores densidades (40 e 80 ind.cm⁻²) apresentaram
203 média de 53,24 ± 4,60 % e 52,95 ± 3,32 % respectivamente, diferindo
204 significativamente da maior densidade de estocagem com 31,54 ± 0,70 % (Figura 3).
205 Portanto, observa-se o efeito negativo na sobrevivência quando ocorre aumento na
206 densidade de estocagem de pós-larvas de *A. brasiliiana*.

207

208 Inserir Figura 3

209

210 4. Discussão

211 O alimento é um fator limitador do crescimento, de larvas de moluscos bivalves
212 quando são cultivadas em altas densidades. Estudos evidenciam que (Loosanoff e
213 Davis, 1963; Sprung, 1984a,b), o excesso de alimento pode causar um gasto energético
214 com a produção de pseudofazes e outros efeitos inibitórios relacionados à fisiologia de
215 digestão, resultando em uma diminuição do crescimento larval, além de ocasionar
216 contaminação bacteriana e toxidez alimentar pela acumulação de ectometabolitos
217 (Loosanoff e Davis, 1963; Sprung, 1984a,b).

218 Ao estudar o efeito da densidade de estocagem de uma espécie-alvo deve se
219 garantir que o alimento não ultrapasse a concentração máxima desejável (Liu et al.,
220 2010). No presente estudo, o residual algal foi verificado diariamente, sendo ajustado
221 quando necessário e mantido na mesma concentração da dieta em cada tratamento.
222 Desta forma, supõe-se que o alimento não tenha sido um fator limitante.

223 Altas densidades de estocagem no cultivo de larvas de bivalves podem ocasionar
224 redução no crescimento, isto pode ser atribuído além da escassez do alimentação,
225 também ao reduzido espaço de confinamento que causará deterioração na qualidade da

226 água (Reghavan e Gopinathan, 2008; Velasco e Barros, 2008). Larvas pelágicas
227 utilizam o velum para nadar e se alimentar, logo um aumento na densidade de cultivo
228 aumenta a possibilidade de colisão entre os indivíduos, que pode resultar numa inibição
229 da atividade alimentar (Liu et al., 2006).

230 Na metamorfose de larvas pelágicas para pé de véliger, quando estas passam a ter
231 hábitos bentônicos, há uma queda na taxa de crescimento (Liu et al., 2006). Além do
232 fator biológico, estudos evidenciam que a área de fundo pode ser um fator limitante de
233 densidade, para larvas com hábitos bentônicos. Em larvas pé de véliger há uma redução
234 no crescimento quando os indivíduos ocupam 100% ou mais da área de cobertura de
235 fundo (Liu et al., 2011). Heasman et al. (2002) observou que ocupar 70% da área de
236 cobertura de fundo na larvicultura de juvenis de vieiras *Pecten fumatus* é um fator
237 limitador da densidade para garantir a manutenção da máxima taxa de crescimento.

238 Usando a área de fundo (100%) como fator limitador da densidade para larvas pé
239 de véliger de *Clinocardium nuttallii*, foi possível determinar que 160 ind.cm⁻² seria o
240 recomendado para indivíduos com até 1mm de comprimento (Liu et al., 2011). O fator
241 área de cobertura de fundo não foi limitador de crescimento para *A. brasiliiana*, pois
242 utilizando a metodologia de Liu et al. (2011) obtivemos ao final do cultivo 26,74 %,
243 29,88 % e 25,29 % de cobertura de fundo para as densidades 40, 80 e 160 ind.cm⁻²,
244 respectivamente. Para pós-larvas de *A. brasiliiana* a densidade de 160 ind.cm⁻² seria
245 recomendado até alcançar 600 µm de comprimento e 40 ind.cm⁻² até alcançar 1mm,
246 mantendo o limite máximo de crescimento.

247 A taxa de crescimento específico em larvas de *Meretrix meretrix* foi inferior a 2%
248 quando estavam no período de metamorfose e assentamento, independente do densidade
249 de estocagem (Liu et al., 2006). No cultivo de pós-larvas de *A. brasiliiana*, aos 21 dias,
250 observou-se queda na TCE (<2%). Aos 24 dias de cultivo as pós-larvas do tratamento

251 com menor densidade (40 ind.cm⁻²) alcançaram 1mm de comprimento, aumentando a
252 taxa de crescimento. O aumento na taxa de crescimento não ocorreu nas outras
253 densidades, onde as larvas ainda não haviam alcançado 1mm de comprimento.

254 As pós-larvas cultivadas em altas densidades apresentaram o menor crescimento.
255 Essa baixa taxa de crescimento diário afetou o comprimento final, pois aos 28 dias de
256 cultivo as pós-larvas cultivadas na maior densidade apresentaram o menor
257 comprimento.

258 A sobrevivência final foi menor na densidade 160 ind.cm⁻². A densidade de
259 estocagem afetou diretamente na sobrevivência de pós-larvas de *A. brasiliiana*. A taxa
260 de sobrevivência de larvas de bivalves diminui continuamente ao longo do tempo,
261 durante o desenvolvimento das fases D-véliger para pé de véliger, mesmo quando as
262 condições de criação se mantiveram favoráveis (Gouda et al., 2006; Magnesen et al.,
263 2006; Saucedo et al.,2007; Velasco e Barros, 2008). No cultivo de larvas de *Argopecten*
264 *nucleus* há uma queda na sobrevivência quando as larvas são cultivadas em altas
265 densidades (Velasco e Barros, 2008). Variação na taxa de sobrevivência entre 50 % e 90
266 % é considerada satisfatória para o cultivo de larvas de moluscos (Utting e Spencer,
267 1991), o que corrobora com a sobrevivência obtida para as densidades de 40 ind.cm⁻² e
268 80 ind.cm⁻².

269

270 5. Conclusão

271 Na larvicultura de larvas assentadas de *A. brasiliiana* deve-se realizar o manejo na
272 densidade de estocagem no decorrer do crescimento. Para larvas pé de véliger de *A.*
273 *brasiliiana* a densidade de 160 larvas.cm⁻² seria recomendado até alcançar 600 µm de
274 comprimento e 40 larvas.cm⁻² até alcançar 1mm, mantendo o limite máximo de
275 crescimento.

276 A densidade de estocagem afeta diretamente o crescimento e a sobrevivência de
277 larvas assentadas de *A. brasiliiana*. Em um sistema de cultivo fechado a densidade de 40
278 larvas.cm⁻² pode ser utilizada mantendo a taxa de crescimento específico em torno de
279 5% ao dia.

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373 Tabela 1. Concentração algal fornecida às larvas de *A. brasiliana* até atingir o estágio
374 juvenil.

Etapas	Dias	Fases	Concentração Algal
1	1 - 7	D véliger / Umbonada / véliger	30.000 cél.mL ⁻¹
2	8 - 12	Pé de véliger	40.000 cél.mL ⁻¹
3	13 - 45	Pé de véliger / Juvenil	50.000 cél.mL ⁻¹

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378 Tabela 2. Taxa de crescimento específico de larvas pós-assentamento de *A. brasiliana*
379 em três diferentes densidades de estocagem para cada período cultivado (dias).

TCE (%.dia ⁻¹)					
Densidade de estocagem (Larvas.cm ⁻²)	Período do cultivo (dias)				
	0-7	7-14	14-21	21-28	0-28
40	8,21±0,33 ^a	6,00±0,32 ^a	0,37±0,32 ^b	5,02±0,32 ^a	4,98±0,08 ^a
80	8,44±0,27 ^a	3,91±0,30 ^b	1,47±0,26 ^a	2,27±0,30 ^b	4,08±0,07 ^b
160	8,52±0,23 ^a	2,73±0,23 ^c	1,19±0,34 ^{ab}	0,82±0,29 ^c	3,37±0,07 ^c

380 Médias com caracteres diferentes diferem significativamente (p<0,05).

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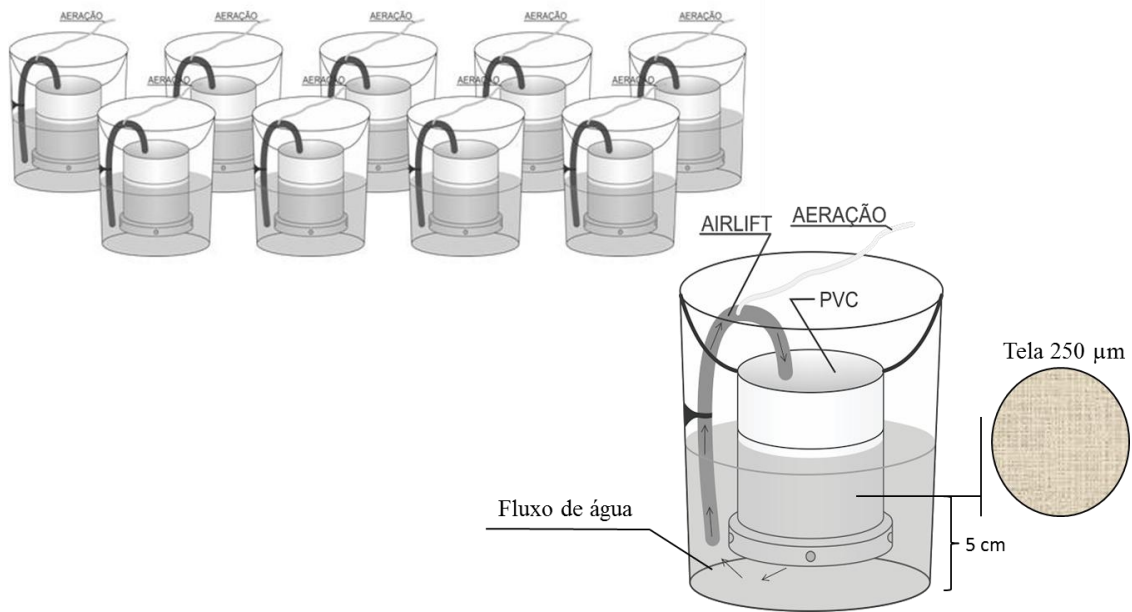
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394 Figura 1. Desenho esquemático das unidades experimentais utilizadas para avaliação do
395 efeito da densidade de estocagem no crescimento e sobrevivência de larvas pé de
396 véliger de *A. brasiliana*.

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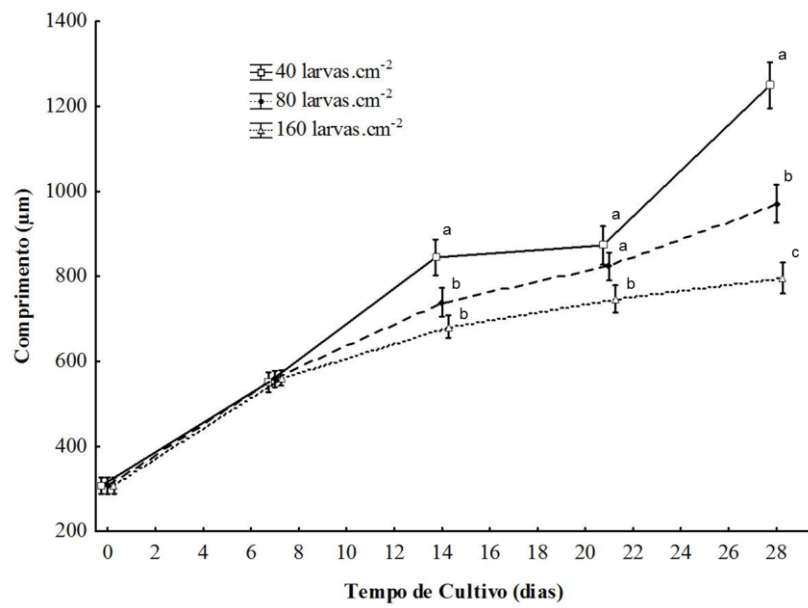
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411 Figura 2. Comprimento médio (\pm EP) de larvas pé de véliger de *A. brasiliana* para cada

412 período do cultivo (7, 14, 21 e 28 dias) em três diferentes densidades de estocagem.

413 Médias com letras sobreescritas diferentes diferem significativamente ($p < 0,05$).

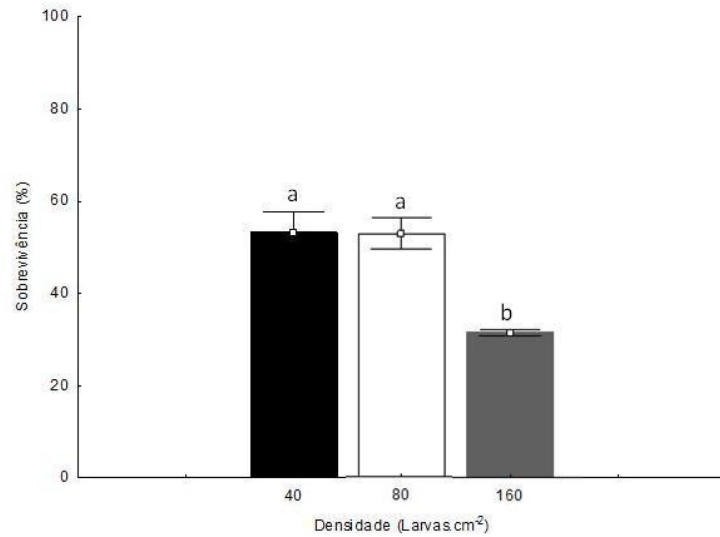
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420 Figura 3. Sobrevivência larval de *Anomalocardia brasiliana* cultivada por 28 dias em
421 diferentes densidades de estocagem. Médias com caracteres diferentes diferem
422 significativamente ($p < 0,05$).

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6. CONSIDERAÇÕES FINAIS

A larvicultura do molusco bivalve *Anomalocardia brasiliiana* pode ser realizada utilizando diversos tipos de dietas microalagais, porém a microalga *I. galbana* quando fornecida isoladamente não apresentou crescimento e sobrevivência larval satisfatório, devendo ser utilizada sempre combinada a uma outra espécie, principalmente, diatomáceas.

No cultivo de pós-larvas do molusco bivalve *A. brasiliiana* deve-se considerar o manejo na densidade de estocagem no decorrer do crescimento das pós-larvas. No início da metamorfose para pé-de-véliger pode-se utilizar a densidade de 160 ind.cm² até que estes alcancem 600µm de comprimento (eixo dorso ventral), posteriormente reduzir a densidade de estocagem para 40 ind.cm² até 1mm de comprimento.

A fim de melhorar os resultados obtidos neste estudo, novos trabalhos deverão ser realizados com objetivo de determinar o melhor sistema de cultivo, aberto ou fechado com recirculação, diferentes condições de temperatura e salinidade, além do uso de probióticos na larvicultura de *A. brasiliiana*.

ANEXOS

Normas da Revista Latin American Journal of Aquatic Research

Instructions for Authors

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- Coelho, V., R.A. Cooper & S. Rodrigues. 2000. Burrow morphology and behaviour of the mud shrimp *Upogebia omissa* (Decapoda, Thalassinidea, Upogebiidae). Mar. Ecol. Progr. Ser., 200: 229-240.

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- Thurman, H. & A. Trujillo. 2002. Essentials of oceanography. Prentice Hall, New Jersey, 524 pp.

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- Brummet, R.E. & B.A. Costa-Pierce. 2002. Village-based aquaculture ecosystems as a model for sustainable aquaculture development in Sub-Saharan Africa. In: B. Costa-Pierce (ed.). Ecological aquaculture: evolution of the blue revolution. Blackwell Science, Oxford, pp. 145-160.

d) Articles published on Internet should indicate: Author(s). Year of publication. Article title. Web site. Date reviewed.

- Walker, J.R. 1997. MLA-Style citations of Internet sources. [<http://www.cas.usf.edu/english/walker/janice.html>]. Reviewed: 24 January 2008.

e) References to articles or books published in CD-Rom should indicate: author(s), year of publication, (CD-ROM), article or book titles, editorial, city.

- Retamal, M.A. 2000. (CD-ROM). Decápodos de Chile. ETI-Universidad de Concepción. Springer-Verlag, Berlin.

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Normas da Revista Aquaculture



Introduction

Types of paper

Original Research Papers should report the results of original research. The material should not have been previously published elsewhere. Articles are expected to contribute new information (e.g. novel methods of analysis with added new insights and impacts) to the knowledge base in the field, not just to confirm previously published work.

Review Articles can cover either narrow disciplinary subjects or broad issues requiring interdisciplinary discussion. They should provide objective critical evaluation of a defined subject. Reviews should not consist solely of a summary of published data. Evaluation of the quality of existing data, the status of knowledge, and the research required to advance knowledge of the subject are essential.

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Technical Papers should present new methods and procedures for either research methodology or culture-related techniques.

The Letters to the Editor section is intended to provide a forum for discussion of aquacultural science emanating from material published in the journal.

Contact details for submission

Papers for consideration should be submitted via the electronic submission system mentioned below to the appropriate Section Editor:

Nutrition:

D.M. Gatlin

The Nutrition Section welcomes high quality research papers presenting novel data as well as original reviews on various aspects of aquatic animal nutrition relevant to aquaculture. Manuscripts addressing the following areas of investigation are encouraged:

- 1) determination of dietary and metabolic requirements for various nutrients by representative aquatic species. Studies may include environmental/stress effects on animal's physiological responses and requirements at different developmental stages;

- 2) evaluation of novel or established feedstuffs as well as feed processing and manufacturing procedures with digestibility and growth trials. Such studies should provide comprehensive specifications of the process or evaluated ingredients including nutrients, potential anti-nutrients, and contaminants;
- 3) comparison of nutrient bioavailability from various ingredients or product forms as well as metabolic kinetics of nutrients, food borne anti-nutrients or toxins;
- 4) identification of key components in natural diets that influence attractability, palatability, metabolism, growth reproduction and/or immunity of cultured organisms;
- 5) optimization of diet formulations and feeding practices;
- 6) characterization of the actions of hormones, cytokines and/or components in intracellular signaling pathway(s) that influence nutrient and/or energy utilization;
- 7) evaluation of diet supplementation strategies to influence animal performance, metabolism, health and/or flesh quality.

Manuscripts concerning other areas of nutrition using novel or advanced methods are also welcome. Please note that in regard to various diet additives such as probiotics, prebiotics, herbal extracts, etc., a very large number of papers have already been published. Therefore, Aquaculture will not continue to accept manuscripts that present initial and preliminary investigations of such additives. Manuscripts addressing these and other feed additives will be accepted for review only if they are of the highest scientific quality and they represent a significant advance in our knowledge of the mechanisms involved in their metabolism. Manuscripts may also be considered if they present clinical efficacy data generated in large-scale trials and economic cost-benefit analysis of these applications.

Aquaculture Production Science:

B.Costa-Pierce

AQUACULTURE PRODUCTION SCIENCE (PS) is one of 5 sections of the international journal AQUACULTURE dedicated to research on improvements and innovations in aquatic food production.

This section supports worldwide dissemination of the results of innovative, globally important, scientific research on production methods for aquatic foods from fish, crustaceans, mollusks, amphibians, and all types of aquatic plants. Contributions are encouraged in the following areas: 1) Improvement of production systems that results in greater efficiencies of resource usage and sustainability of aquaculture; 2) Effective applications of technologies and methods of aquaculture production for improved stocking regimes; 3) The use of new species and species assemblages; and, 4) Investigations to minimize aquaculture wastes and improve water quality, including technologies for nutrient recycling in aquaculture ecosystems, and potential synergy of aquaculture and other food production systems using methods such as polyculture and integrated aquaculture. Aspects of seafood processing and technology will not be considered in this section although aquaculture techniques that may influence the

nutritional value of aquatic food products may be considered in the Nutrition Section.

Physiology:

Fish: A. P. (Tony) Farrell

Invertebrates: J. Benzie

The Physiology Section welcomes high quality papers that present either novel research data or original reviews. The content must be relevant to solving aquaculture problems on all aspects of the physiology of cultured aquatic animals and plants.

Submitted manuscripts must have a valid hypothesis or objective, clearly state the relevance to aquaculture, have proper experimental design with appropriate controls and utilize appropriate statistical analysis. Mention of trade names is limited to the main text.

Relevant physiological topics include, but are not limited to:

- Reproductive and endocrine physiology, including control of development and sex differentiation, induced ovulation and spermiation, gamete quality, storage and cryopreservation, physiology of gynogenetic, and triploid and transgenic organisms
- Cardiorespiratory, muscle and exercise physiology
- Osmoregulatory physiology
- Digestive physiology, including endocrine and environmental regulation of growth
- Larval physiology and ontogeny, including metamorphosis, smolting and molting
- Performance under variable culture conditions, including temperature, water quality, rearing density, and stress and disease physiology
- Physiology of harvest and handling techniques

Genetics:

G. Hulata

The Genetics Section welcomes high-quality research papers presenting novel data, as well as critical reviews, on various aspects of selective breeding, genetics and genomics. Submitted manuscripts must have a valid hypothesis or objective, clearly state the relevance to aquaculture, have proper experimental design with appropriate sample size and controls and utilize appropriate statistical analysis.

Relevant genetics topics include, but are not limited to:

- Breeding programs using classic selection procedures, markers or combining marker assisted selection with classic selection
- Applications of crossbreeding and interspecific hybridization
- Evaluation of commercially important phenotypes among cultured strains, populations or stocks
- Applications of biotechnology and genetic manipulation methods
- Development of linkage maps, identification of QTL or association of commercially important traits with specific gene(s). Where appropriate, linkage maps should include co-dominant markers, such as microsatellite DNA and SNP

markers, to enable application to other populations and facilitate comparative mapping.

Aquaculture will NOT accept manuscripts dealing with the application of well-described techniques to yet another species, unless the application solves a specific biological problem important to aquaculture production; or manuscripts dealing with gene cloning, characterizing of microsatellites, species identification using molecular markers, EST papers with small collections, or mapping papers with a small number of markers, unless the papers also deal with solving a biological problem that is relevant to aquaculture production.

Aquaculture will not accept manuscripts focusing mainly on population genetics studies that are based on RAPD and AFLP markers, since the dominance and multilocus nature of the fingerprints are not suitable for making inferences about population genetic diversity and structure.

Sustainability and Society:

D.C. Little

The Sustainability and Society section of the journal Aquaculture invites articles at the interface of natural and social sciences that address the broader roles of aquaculture in global food security and trade.

Aims and scope of the Sustainability and Society section are the: global dissemination of interdisciplinary knowledge regarding the management of aquatic resources and resulting impacts on people. Interconnections with other sectors of food production; resource management and implications for societal impact. Going beyond a narrow techno-centric focus, towards more holistic analyses of aquaculture within well-defined contexts. Enquiry based on understanding trajectories of change amid the global challenges of climate change and food security. Mixed methods and approaches that incorporate and integrate both social and natural sciences. Relevance for the diverse range of policy makers, practitioners and other stakeholders involved. Articles that take a value chain approach, rather than being wholly production orientated, are encouraged.

Disease

B. Austin

The Disease sections welcomes critical reviews and high quality articles containing novel data on all aspects concerning diseases of farmed aquatic species. The aims of the section are: description of new and emerging diseases including characterization of the causal agent(s), development in the understanding of fish pathogens for example including new methods of growth where this has been a problem for fastidious organisms, pathogenicity and epizootiology, developments in the diagnosis of disease going beyond the use of standard well used methods, and methods of disease control, notably new developments in vaccines, immunostimulants, dietary supplements, medicinal plant products, probiotics, prebiotics and genetically-disease resistant stock. Relevance to aquaculture must be demonstrated. Articles, which adapt well known methods without further refinement of those methods, are unlikely to be accepted.



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- Aim to use the following fonts in your illustrations: Arial, Courier, Times New Roman, Symbol, or use fonts that look similar.
- Number the illustrations according to their sequence in the text.
- Use a logical naming convention for your artwork files.
- Provide captions to illustrations separately.
- Size the illustrations close to the desired dimensions of the printed version.
- Submit each illustration as a separate file.

A detailed guide on electronic artwork is available on our website:

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Regardless of the application used other than Microsoft Office, when your electronic

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EPS (or PDF): Vector drawings, embed all used fonts.

TIFF (or JPEG): Color or grayscale photographs (halftones), keep to a minimum of 300 dpi.

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Please do not:

- Supply files that are optimized for screen use (e.g., GIF, BMP, PICT, WPG); these typically have a low number of pixels and limited set of colors;
- Supply files that are too low in resolution;
- Submit graphics that are disproportionately large for the content.

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Text graphics may be embedded in the text at the appropriate position. See further under Electronic artwork.

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Number tables consecutively in accordance with their appearance in the text. Place footnotes to tables below the table body and indicate them with superscript lowercase letters. Avoid vertical rules. Be sparing in the use of tables and ensure that the data presented in tables do not duplicate results described elsewhere in the article.

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Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. Unpublished results and personal communications are not recommended in the reference list, but may be mentioned in the text. If these references are included in the reference list they should follow the standard reference style of the journal and should include a

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Reference to a book:

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Reference to a chapter in an edited book:

Mettam, G.R., Adams, L.B., 2009. How to prepare an electronic version of your article, in: Jones, B.S., Smith, R.Z. (Eds.), *Introduction to the Electronic Age*. E-Publishing Inc., New York, pp. 281–304.

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